

Volume 51

Issue-5

October 2024



THE INDIAN ECOLOGICAL SOCIETY

# THE INDIAN ECOLOGICAL SOCIETY

(www.indianecologicalsociety.com) Past President: A.S. Atwal and G.S. Dhaliwal (Founded 1974, Registration No.: 30588-74)

#### **Registered Office**

College of Agriculture, Punjab Agricultural University, Ludhiana – 141 004, Punjab, India (e-mail : indianecologicalsociety@gmail.com)

#### **Advisory Board**

Kamal Vatta

Chanda Siddo Atwal

Asha Dhawan

#### **Executive Council**

President

A.K. Dhawan

Vice-President

R. Peshin

**General Secretary** 

S.K. Chauhan

#### Joint Secretary-cum-Treasurer

Vijay Kumar

Councillors

Vikas Jindal

Vaneet Inder Kaur

#### **Editorial Board**

#### **Managing-Editor**

A.K. Dhawan

#### **Chief-Editor**

Sanjeev K. Chauhan

#### **Associate Editor**

S.S. Walia Gopal Krishan K.K. Sood

#### Editors

Neeraj Gupta	S.K. Tripathi	Ashalata Devi	Bhausaheb Tambat
V. Ravichandran	S. Sheraz Mahdi	N.S. Thakur	P. Siddhuraju
Sunny Agarwal	Anil Kumar Nair	Vikas Sharma	Debajit Sarma
Benmansour Hanane	Jawala Jindal	Anil Sharma	Prabjeet Singh
S.V.S. Gopala Swami	B.A. Gudade		

#### See detail regarding editorial board at web site (https://indianecologicalsociety.com/editorial-board/)

The Indian Journal of Ecology is an official organ of the Indian Ecological Society and is published bimonthly in February, April, June, August, October and December by Indian Ecological Society, PAU, Ludhiana. Research papers in all fields of ecology are accepted for publication from the members. The annual and life membership fee is Rs (INR) 1000 and Rs 8000, respectively within India and US \$ 100 and 350 for overseas. The annual subscription for institutions is Rs 8000 and US \$ 300 within India and overseas, respectively. All payments should be in favour of the Indian Ecological Society payable at Ludhiana. See details at web site. The manuscript registration is Rs 500.

#### **KEY LINKS WEB**

website: http://indianecologicalsociety.com

Membership:http://indianecologicalsociety.com/society/memebership/

Manuscript submission: http://indianecologicalsociety.com/society/submit-manuscript/

Status of research paper:http://indianecologicalsociety.com/society/paper-status-in-journal-2/

Full journal: http://indianecologicalsociety.com/society/full-journals/



### Trends of Forest Phenology Studies in India-Challenges and Opportunity for Satellite Remote Sensing and Near Surface Sensors

#### Dhruvi Sedha, Chandra Prakash Singh, Hitesh Solanki<sup>1</sup>, Jincy Rachel Mathew<sup>1</sup>, Mehul R. Pandya, Bimal K. Bhattacharya<sup>1</sup>, C. Jeganathan<sup>2</sup> and Siddhartha Khare<sup>3</sup>

EPSA, Space Applications Centre (SAC), ISRO, Ahmedabad-380 015, India <sup>1</sup>Department of Environmental Sciences, Gujarat University, Ahmedabad-380 009, India <sup>2</sup>Department of Remote Sensing, Birla Institute of Technology (BIT), Mesra, Ranchi-835 215, India <sup>3</sup>Department of Civil Engineering, Geomatics Engineering Division, Indian Institute of Technology, Roorkee-247 667, India *E-mail: dhruvisedha@gmail.com* 

**Abstract:** Phenology, for being pulse of the ecosystem, elucidates the complex relationship between climate and vegetation, serving as a vital indicator of climate change. India, a biodiversity hotspot, has forests classified into 5 major groups, 16 subgroups, and 255 subtypes based on climatic, edaphic, and successional variations, displaying varying phenological traits. Recent advancements in technology have revolutionized our ability to collect valuable data across various spatial and ecological scales. Despite the abundant observational data capturing phenological changes at various scales, remote sensing remains a relatively less explored tool for operational forest phenology monitoring in Indian forest ecosystems. This comprehensive review explores the latent potential of long-term satellite remote sensing data supported with near surface sensor system networks to reveal complex patterns of phenological dynamics within a spectrum of diverse forest types in India. We found that, majority of the research has predominantly focused on field-based observations limited to moist and dry tropical forests, followed by montane temperate and subtropical forests driven by their ecological importance, higher accessibility, and their sensitivity to climate change impacts. Through the utilization of advanced satellite sensors (e.g., MODIS, Landsat, and Sentinel) thorough investigations have successfully examined the dynamics of vegetation in forest phenological studies. Looking ahead, integrated studies with diverse monitoring methods are crucial for upscaling phenological observations robustly. These advances are necessary to precisely quantify the impacts of a changing world on forest phenology.

Keywords: Phenology, Forest dynamics, Phenometrics, Remote Sensing, Vegetation patterns

Phenology, as a holistic environmental discipline that integrates biometeorology, ecology, and evolutionary biology (Morellato et al 2016), functions as the pulse of ecosystems, elucidating the intricate relationship between climate and vegetation, acting as a vital indicator of climate change (Rosenzweig et al 2008). Phenology has served as an important diagnostic proxy for a wide variety of areas, such as climate change (Jin et al 2017, Brown et al 2017), food security (Lobell et al 2008, Alemu and Henebry 2017, Gao and Zhang 2021), drought (de Beurs and Henebry 2008, Chang et al 2021), forest fire risk (Chéret and Denux 2011, Bison 2022), frost hazard (Ge et al 2013, Hänninen 2006), landscape dynamics (Jeganathan et al 2014, Dronova and Taddeo 2022), and biogeochemical cycling (Gray et al 2014, Piao et al 2019), as well as input to a number of prognostic models that are frequently used to make predictions in these fields (Ma et al 2022). It is possible to demonstrate the consequences of environmental change independent of ecological observations based on long-term phenological data (Walther et al 2002, Rosenzweig et al 2007, Jiang et al 2014).

Over the past two decades, there has been a notable

growth in the development of multidisciplinary research frameworks for phenology, which incorporate technological advancements and aim to explore the intricate relationships between phenology, ecosystem function, and species across diverse locations and time periods (Cleland et al 2007, Morisette et al 2009, Pau et al 2011). The Intergovernmental Panel on Climate Change (IPCC) has prioritized the collection and analysis of phenological observations to enhance our comprehension of the biological effects of climate change in response to mounting evidence of early spring onsets (Ciais et al 2014). Moreover, phenology plays a role, as an aspect of Biodiversity Conservation particularly in achieving the goals outlined in objectives 13 (Climate action) and 15 (Life on land) of the Sustainable Development Goals (SDGs). It is essential to study patterns and their influencing factors to effectively develop strategies, for both adapting to and mitigating the impacts of climate change (Radeloff et al 2019).

The recent resurgence of phenology as an ecological specialization is driven by climate change implications, highlighting phenological variations as sensitive indicators of global warming's biological impact (Walther et al 2002).

Changes in vegetation attributes, coupled with the intricate connections to climate change, remain crucial, lacking a comprehensive mechanistic understanding (Richardson et al 2012). Technological advancements, particularly in remote sensing, have expanded phenological observation beyond manual methods, enabling efficient collection of substantial monitoring data from both ground-based and remote sensing technologies (Zhang et al 2003, Zhang et al 2016).

The study of vegetation phenology can provide an imperative knowledge about previous, present, and probable future ecosystem conditions (Wolkovich et al 2014). Alterations in the timing of phenological phenomena could desynchronize species interactions because different species respond to climate change differently (Thackeray et al 2016). The progression of vernal phenomena, the extension of the vegetative period, and the temporal displacement of the conclusion of the growing season (onset of senescence) are just a few of the important indicators of phenological response to the ongoing climate change which are responsible for change in carbon budget. This highlights the interest at the regional and global scales in understanding how climate influences forest ecosystems, with Indian forests serving as a notable case (Chaturvedi et al 2011, White et al 2014, Kumar et al 2019).

In the global context, Indian forests hold exceptional importance as mega-biodiversity hotspots and ecologically significant regions. Indian region and its environs is internationally recognized as one of the mega-diverse countries, harboring approximately 7-8% of the world's recorded species and encompassing four of the biodiversity hotspots identified globally: the Himalaya, Indo-Burma, Western Ghats and Sri Lanka, and Sunda land (Sivaperuman and Venkataraman 2018). These diverse ecosystems represent unique reservoirs of biodiversity and play a vital role in sustaining ecological processes and supporting numerous endemic species. Understanding the phenology of Indian forests becomes imperative in this broader global perspective of biodiversity conservation and sustainable ecosystem management.

#### **Brief Historical Overview**

#### In-situ observation

The commencement of efforts to observe the phenology of the tree species in India is believed to have taken place in the 1950s. Prior to that, India had undertaken several investigations pertaining to agricultural harvest, agro ecology of the flora (Misra 1946), classification of forests (Champion 1936), and silviculture (Chatterjee 1939). However, no explicit research has been documented on the phenology of forests. The eighth general conference of the United Nations Educational, Scientific, and Cultural Organization (UNESCO) convened in Montevideo in 1954. The conference aimed to promote global or regional initiatives on scientific matters, specifically those related to the humid tropical zone. The committee was established with the objective of examining issues related to research in the fundamental aspects of the natural sciences that are relevant to the humid tropical zones. The aim was to encourage and support the continuation of such research, and to take the necessary steps to initiate or promote research in those areas where significant problems were not adequately addressed. The symposium featured the participation of several Indian authors who presented their research on the humid tropics of India (Puri 1958).

The deep link between phenology and different aspects of our environment explains why, despite observing phenological events for thousands of years, it was officially acknowledged as a scientific discipline only in the 1900s (Fig. 1). Recognizing the vital role of phenology in various facets of our environment underscores its importance in understanding how our ecosystems work and managing resources, especially with changes in climate and habitats. The majority of phenology related studies in India are based on the field observations carried out during (1 to 3 years' timeframe), with a few longer-term studies (Datta and Rane 2013, Suresh and Sukumar 2018). Figure 1 depicts India's phenological study chronology. These community-level investigations provide a baseline understanding of plant behavior and how it responds to ecological conditions (Hag et al 2021). The limited period of temporal phenology variations hinders comprehending their causes and effects difficult. Few studies quantitatively relate these patterns to climate factors, whereas the majority describe the seasonal rhythms of leaf flushing, flowering, and fruiting. Earlier studies estimated phenophases time and extent by counting species in leaf, flower, or fruit (Kumar and Shahabuddin 2005).

Shukla and Ramakrishnan (1982) conducted the first formal phenological study in Meghalaya, in the northeastern region of India. The majority of phenological research have come from South (Prasad and Hegde 1986, Bhat 1992, Murali and Sukumar 1993, Sundarapandian et al 2005, Nanda et al 2011, Prabakaran et al 2013, Nanda et al 2015, Nanda 2017) and North-east India (Shukla and Ramakrishnan 1982, Boojh and Ramakrishnan 1983, Baruah and Ramakrishnan 1989a, Kikim and Yadava 2001, Paul et al 2018, Devi et al 2020) followed by Western (Jadeja and Nakar 2010), Northern (Ralhan et al 1985, Khare et al 2016), and North-Western India (Kushwaha and Singh 2005), with a few from Central India (Newton 1988) in later years. In India, the phenological research has mainly concentrated on dry deciduous (Prasad and Hegde 1986,



935

Murali and Sukumar 1993, Murali and Sukumar 1994, Kushwaha and Singh 2005, Nanda et al 2011, Nanda et al 2014) and evergreen forests (Krishna et al 2007, Mohandass et al 2016, Nanda 2017) in the south, with less attention paid to tropical (Baruah and Ramakrishnan 1989b, Sundarapandian et al 2005) and subtropical moist forests (Baruah and Ramakrishnan 1989a, Bhat et al 2000, Bhat and Murali 2001) in the northeast (Table 1). This underscores the need for further research and in-depth investigations into the phenological patterns and processes of these tree species to enhance our understanding of their reproductive strategies, ecological interactions, and conservation requirements.

To enhance our understanding of phenology in Indian forests, comprehensive studies encompassing a wide range of tree species are crucial (Kushwaha and Singh 2008). Such investigations yield valuable insights into the temporal patterns of key phenological events, including flowering, fruiting, and leaf emergence. Moreover, they unravel the underlying ecological mechanisms governing these events, shedding light on aspects such as reproductive success, pollination dynamics, seed dispersal mechanisms, and the potential implications of climate change on patterns of phenology. Nevertheless, conducting in-situ observations for phenological studies in Indian woods encounters inherent limitations. The vast geographic expanse and diverse forest types pose challenges in achieving representative species and location coverage. Additionally, logistical complexities, time-intensive nature, and associated costs further impede the execution of in-situ observations. These constraints accentuate the necessity of complementing in-situ data collection with remote sensing techniques and automated phenocam towers, leveraging a more comprehensive phenological observation strategy (Chandra et al 2022). By integrating multiple methodologies and expanding the scope of phenological research in Indian forests, valuable insights into ecological dynamics and conservation imperatives can be gleaned.

Few qualitative studies have been conducted to examine evolutionary concerns associated with ecological determinants of tree phenology (Murali and Sukumar 1993, Murali and Sukumar 1994). With the few exceptions of studies (Krishnan 2002, Joshi and Janarthanam 2004), most of the research conducted focuses on trees. Nautiyal et al (2001) examined the phenology of 171 grass, herb, forb, and shrub species in an alpine pasture at an elevation greater than 3000 m a.m.s.l. Based on the findings, it has been discovered that flowering takes place between May and August after snowmelt, with fruiting occurring between June and August before senescence takes hold before the start of winter. The study conducted by Vashistha et al (2009) investigated the phenology of a total of 103 species of graminoids, forbs, and shrubs in an alpine pasture situated at an altitude of 3200 m a.m.s.l. in the North-West Himalaya region of India. In their research Joshi and Janarthanam (2004) examined the flowering phenology of a total of 113 endemic species in Goa, encompassing a diverse array of life-forms such as monocots, dicots, herbs, shrubs, trees, and climbers. Moreover, it has been stated that the plateaus located in the northern region of the Western Ghats are distinctive owing to their copiousness of herbaceous native species. The distribution of these endemics is closely linked to precipitation patterns, thus any alteration in the long-term moisture regime would have an impact on their habitat. In a study conducted by Krishnan (2002), the reproductive phenology of sixty understorey species, comprising of herbs such as terrestrial shrubs, orchids, and small trees, was examined in a moist evergreen forest located in the southern Western Ghats. Few studies have been carried out on the phenology of specific tree species that are either rare or have significant ecological roles (Ganesh and Davidar 1997, Shankar 2001, Bhat and Kaveriappa 2009, Datta and Rane 2013, Borah and Devi 2014, Nath et al 2016, Upadhaya et al 2018, Devi et al 2020, Chauhan and Chauhan 2020, Nandy et al 2021, Subin et al 2022).

Effective forest management and biodiversity conservation rely on comprehensive long-term datasets on ecosystem dynamics. Unfortunately, in India, there is a limited number of long-term observation sites, and access to such datasets is currently restricted. To comprehensively understand seasonal variations and climatic fluctuations and to detect alterations in the initiation, duration, extent, timing, and synchronization of vegetative and reproductive phases, extensive multi-year investigations are essential. Monitoring changes in species-level biological responses to local climate change can be achieved by utilizing prolonged phenological records derived from plant and animal observation networks. Extended studies in various regions such as Southeast Asia, Australia, Neotropics, Europe/North, America/South, Africa, and America commonly span over 10 - 30 years, occasionally even longer. In an era when the anticipated impacts of climate change are under intense scrutiny, it would be judicious to evaluate the ecological significance and potential advantages of establishing a network of phenological stations.

#### Satellite Remote Sensing in Phenological Studies

Remote sensing is recognized as an integral component in the advancement of phenological studies, as it offers substantial potential for comprehending widespread seasonal phenomena over extensive areas (Reed et al 2003). By analyzing long term phenological

Study region/State	Vegetation type	Reported (V	Vith date)	Field or	Study	Reference
		Leaf flushing	Leaf fall	based	duration	
Meghalaya North-Eastern India	Sub-tropical humid forest	January-March	February-April	Field based	2 years	Shukla and Ramakrishnan (1982)
Meghalaya North- Eastern India	Deciduous Timber Tree	March- early April (From $1^{st}$ of March at lower altitude and from $20^{th}$ of March at higher altitude)	February-March	Field based	1 year	Boojh and Ramakrishnan (1983)
Kumaun Himalaya, Uttarakhand North India	Sub-tropical Evergreen	March-April	February-April	Field based	1 year	Ralhan et al (1985)
	Deciduous	March-April	October-January			
Bandipur, Karnataka South India	Tropical dry deciduous forest	April-May	December- January	Field based	4 years	Prasad and Hegde (1986)
Madhya Pradesh Central India	Moist Deciduous	April - July	February-April	Field based	14 months	Newton (1988)
Shillong, Meghalaya North-East India	Tropical trees	Early April - May	March-April	Field based	1 year	Baruah and Ramakrishnan 1989a)
Uttar Kannada, Karnataka South India	Tropical moist forest	February-March	November- January	Field based	2 years	Bhat (1992)
Mudumalai Sanctuary, Tamil Nadu South India	Tropical Dry Deciduous Forest	January-February	NA	Field based	2 years	Murali and Sukumar (1993)
Mudumalai, Tamil Nadu Southern India	Tropical Dry Forest	January-March	NA	Field based	2 years	Murali and Sukumar (1994)
Manipur North-east India	Subtropical deciduous and evergreen forest	March-April	November- February	Field based	1 year	Kikim and Yadava (2001)
Western Ghats Tamil Nadu, South India	Tropical moist deciduous forest	February-March	December- January	Field based	2 years	Sundarapandian et al (2005)
Vindhyan Plateau, Sonbhadra Uttar Pradesh North-west India	Tropical dry deciduous forest	March-June	January-March	Field based	2 years	Kushwaha and Singh (2005)
Similipal Biosphere Reserve, Orissa Eastern India	Tropical Moist Deciduous Forest	April	February-April	Field based	1 year	Mishra et al (2006)
Bala Fort Rajasthan Western India	Tropical dry- deciduous thorn forest	February-May	October- December	Field based	3 years	Yadav and Yadav (2008)
Girnar Reserve Forest, Junagadh Gujarat,Western India	Subtropical deciduous forest	February-March	January	Field based	1 year	Jadeja and Nakar (2010)
Bhadra wildlife sanctuary, Karnataka South India	, Tropical dry deciduous forest	April-May	January-February	Field based	2 years	Nanda et al (2010)
Katerniaghat wildlife sanctuary, Uttar Pradesh North-west India	Tropical moist deciduous forest	End of March- April	November	Field based	11 months	Bajpai et al (2012)
Uttar Kannada Kamataka	Evergreen forest	August	Мау	Satellite +	8 years	Prabakaran et al
South India	Semi-evergreen forest	June	February	available data		(2013)
	Moist deciduous forest	Мау	January			

Table 1. Phenological studies inclue	ng location, forest type:	s, methods employed, stu	dy duration and ke	y findings:
	., , , , , , , , , , , , , , , , , , ,		/	

Cont...

Study region/State	Vegetation type	Reported (V	Vith date)	Field or	Study	Reference
		Leaf flushing	Leaf fall	satellite based	duration	
Hollongapar Gibbon Wildlife Sanctuary, Assam North-east India	Moist Tropical Forest	December and in May (observed twice)	January- February	Field based	2 years	Borah and Devi (2014)
	Tropical dry deciduous forest	Beginning of May –end of May	Mid of January			
Bhadra wildlife sanctuary, Karnataka South India	, Tropical evergreen forest	Early January- end of February	Mid of February	Field based	2 years	Nanda et al (2015)
Western Himalayan region of Doon valley, Uttarakhand North India	Moist Deciduous Forest	April	January-March	Satellite based	2 years	Khare et al (2017)
Kemmanugundi, Bhadra wildlife sanctuary Karnataka South India	Tropical evergreen forest	December-early January	August to October	Field based	2 years	Nanda (2017)
Arunachal Pradesh North-east India	Temperate mixed broad-leaved forests	May-June	October- November	Field based	1 year	Paul et al (2018)
Arunachal Pradesh North-east India	Dry mixed deciduous forests	July- October	January-April	Satellite based	33 years	Mohapatra et al (2019)
India	Dry and moist teak forests	July-August	November-end of December	Satellite based	13 years	Ghosh et al (2019)
Nimaichandpur-II, Hailakandi district Assam North-east India	Tropical Deciduous tree- Parkia timoriana	April- May	November- January	Field based	2 years	Devi et al (2020)
Narmada, Gujarat Western India	Tropical dry deciduous forest from 2003-2004	28 July	13 February	Satellite + ground based available data	2 years	Malhi et al (2021)
	Tropical dry deciduous forest from 2013-2014	11 August	22 March			
Rajmahal hills, Jharkhand Eastern India	Deciduous broadleaf forest	March (JD 81)-April (JD 97)	March (JD 65)- March (JD 81)	Satellite based	19 years	Ranjan and Gorai (2022)

Table 1. Phenological studies including location, forest types, methods employed, study duration and key findings:

data from forests, important issues with climate change modelling and monitoring can be addressed (Prabakaran et al 2013). Observations of phenological occurrences on the ground can give an accurate assessment and can provide a precise understanding of phenological patterns (Nandy et al 2021). However, these conventional fieldbased methods have several limitations like, huge investment of time and effort to cover a vast region, and it's difficult to make daily observations (required during transition period). Furthermore, these observations face challenges in terms of spatial specificity and are hard to modify. There is also a lack of a substitute to model them for different scenarios or specific conditions. It is at this point that remote sensing emerges with its advantages to address these shortcomings. By utilizing the benefits of stratification, remote sensing mapping and stratification could cover a vast area with even low intensity sampling. With greater accuracy and precision thanks to the satellites' temporal revisits and long historical records, it is possible to evaluate and analyze altered scenarios. Freely available passive optical satellite imageries with high temporal frequency were widely employed for vegetation phenology study in India (Dash et al 2010, Ghosh et al 2019, Jeganathan et al 2010, Pillai et al 2019, Ayushi et al 2022) and the rest of the world to get over these restrictions (Heumann et al 2007, White et al 2014, Zhang et al 2003, Zheng et al 2022).

#### **Role of Spatial and Spectral Resolutions**

Accurate assessment of plant phenology's climate change sensitivity requires evaluating year-to-year spatial variations in leaf flush and fall timing over large areas and extended periods. Satellite imagery is optimal for discerning plant phenology across spatial scales (Czernecki et al 2018, Pastor-Guzman et al 2018, White et al 2009). Spatial and spectral resolutions of satellite sensors are crucial for phenology data quality. High spatial resolution provides detailed local views, while lower resolution offers broader perspectives. Higher spectral resolution enables precise vegetation characterization (Wingate et al 2015).

Commonly used sensors for plant phenology in India include AVHRR (Prasad et al 2008), MERIS MTCI (Jeganathan et al 2010), MODIS (Deka et al 2019), and Landsat (Mohapatra et al 2019). Synchronizing data between sensor families offers advantages like real-time monitoring, increased observation frequency, and more cloud-free observations in evergreen forests (Pastick et al 2018, Li et al 2019). Phenology detection depends on data availability and external factors like cloud cover (Younes et al 2021). The integration of various observation techniques (field, PhenoCam, satellite) at different levels (Species,



[FBSL: Field Based & Species Level, FBLL: Field Based & Landscape Level, FBRL: Field Based & Regional Level, PBSL: Phenocam Based & Species Level, PBLL: Phenocam Based & Landscape Level, PBRL: Phenocam Based & Regional Level, RBSL: Remote Sensing Based & Species Level, RBLL: Remote Sensing Based & Landscape Level, RBRL: Remote Sensing Based & Regional Level]

Fig. 2. Number of studies published using different techniques and level of observations

landscape, regional) contributes to comprehensive phenological studies.

# Long Term Remote Sensing-Based Phenological Studies Combining *In-Situ* and Remote Sensing Data

Efforts to observe the dynamics of the vegetated land surface from space commenced at the opening of Landsat era (Henebry and de Beurs 2013). Despite Landsat's continuous Earth surface data since 1970, its use for longterm ecosystem studies in India only started in the 2000s, initially limited by cost and processing constraints. The USGS's 2008 open access policy for Landsat data enabled broader time series research on ecology and biodiversity. Vegetation indices derived from remote sensing data contribute significantly to various aspects of forest monitoring, such as forest classification, analysis of vegetation phenology, change detection, and the retrieval of forest biophysical parameters (Glenn et al 2008, Lambert et al 2013).

The widely used normalized difference vegetation index (NDVI) is a key parameter for vegetation studies. A study on moist deciduous forests in the Western Himalaya, using Landsat 8 Operational Land Imager (OLI) and Thermal Infrared Sensor (TIRS) data, assessed phenological changes via NDVI and land surface temperature (LST) (Khare et al 2016). However, limitations in NDVI's ability to fully capture temporal vegetation development led to the introduction of the temporal normalized phenology index (TNPI), offering a valuable alternative to long-term monthly records for understanding forest phenology (Khare et al 2017, Khare and Rossi 2019).

Prasad et al (2008) used NOAA-AVHRR data (1990-2000) to study phenological variations in Indian forests, finding inverse correlations between NDVI and temperature, and positive correlations with precipitation. This aligns with the known influence of these variables on plant phenology, affecting energy, water, and CO2 fluxes (Vadrevu et al 2007). Few studies have used remote sensing to extract phenological variables in India's tropical and subtropical regions (Mohapatra et al 2019). Estimating phenological parameters for these vegetation types via remote sensing will enhance our understanding of their response to climate change.

A continental-scale vegetation map was created using NOAA AVHRR GAC data (1.1 km/4 km resolution), examining NDVI variations due to stress, leaf activity, vegetation amount, and seasonal changes. For finer regional-scale monitoring, the IRS Wide Field Sensor (WiFS) was developed to address NOAA AVHRR limitations (Roy and Joshi 2002). WiFS's temporal NDVI data effectively monitor vegetation cover and phenology (Singh et al 1999), suitable for tracking short-term vegetation changes regionally (Joshi et al 2006, Joshi et al 2001). With two radiometers, multiple spectral bands, and a wide sweep width, WiFS provides crucial vegetation information at 23.5 m (VNIR) and 70.5 m (SWIR) resolutions. It's valuable for vegetation discrimination, cover mapping, and environmental monitoring, offering sufficient temporal, spatial, and spectral resolutions for regional vegetation dynamics studies.

The low temporal resolution constitutes a significant hindrance to the in-depth examination of dynamic variations, emerging as a notable obstacle in the analysis. However, to overcome these challenges, since the early 2000s, MODIS (Terra and Aqua satellites) and VEGETATION (SPOT satellite) sensors have provided improved multispectral timeseries, extending remote sensing applications to phenology and vegetation monitoring over large areas.

While NDVI has been traditionally used for vegetation phenology analysis, recent research focuses on the Enhanced Vegetation Index (EVI), which accounts for background and atmospheric effects and shows increased sensitivity in high biomass areas. Gupta et al (2014) employed MODIS EVI data from 2003 to 2012 to track changes in the phenological transition dates of tropical deciduous broadleaf forests (DBF) in Eastern India. The objective of the study was to examine seasonal and interannual changes in vegetation, with a particular focus on the districts of West Singhbhum and Sundargarh. The results demonstrated that there was an upsurge towards forest degradation: (i) the EVI varied from 0.41 in early April to 0.71 in mid-October; and (ii) the overall pattern declined with a slope of 0.0022. The findings indicate that the end of season (EOS) varied between 1<sup>st</sup> of February and 7<sup>th</sup> March, while the start of season (SOS) varied between 2<sup>nd</sup> of May and 20<sup>th</sup> June. The length of the season decreased overall, peaking in 2007 at 302 days and falling to 235 days in 2010.

Ghosh et al (2019) used MODIS-derived EVI to ascertain that the temporal extent of the growing season in moist teak (Tectona grandis) forests have a 48-day longer growing season than dry teak forests, with significant LAI differences (~2.8). The study established a significant correlation between changes in phenology and shifts in rainfall patterns. In their study, Chakraborty et al (2018) examined the spatial patterns of a significant downward trend of forest seasonal greenness over a wide range of forest types in India (2001-2014) using data from an 8-day composite MODIS NDVI time series. Deka et al (2019) utilized a threshold-based approach for phenological parameter extraction, finding stronger NDVI-temperature correlations than NDVI-precipitation. Singh et al (2020a) reported significant NDVI-rainfall associations, while Deka et al (2019) noted strong NDVItemperature correlations in humid subtropical regions, suggesting that temperature could be a critical factor in the context of climate change.

Nandy et al (2021) examined the phenological changes of various Sal (*Shorea robusta*) forests in India and investigated the connection between phenology metrics and meteorological variables. Several phenological metrics such as start of season (SOS), length of the season (LOS), maximum of the season (MAX), and end of season (EOS) for teak growing in Narmada forests, Gujarat, India, were calculated using MODIS NDVI time series. Malhi et al (2021) used the derivative method to extract phenology metrics, identifying inflection points in NDVI time series. This method assumes the highest NDVI change rate indicates the start or end of the growing season, extracting key variables like SOS, peak of season (POS), and EOS.

Using a MODIS-EVI dataset, Ranjan and Gorai (2022); monitored phenological trends over the Indian Rajmahal Hills from 2001 to 2019. According to the study's findings, the SOS typically occurred between late, around 22<sup>nd</sup> March (Julian day (JD) 81) up till 7<sup>th</sup> April (JD 97), whereas EOS has primarily been recorded between early 6 March (JD 65) up till 22<sup>nd</sup> March (JD 81). Additionally, between 2001 and 2010, the SOS and EOS of many vegetation types showed a delayed tendency. On the other hand, from 2010 to 2019, the SOS and EOS of every type of plant exhibited an upward trend.

With the use of the SPOT-VGT NDVI time series data product and the TIMESAT tool, Prabhakaran et al (2013); obtained the forest phenological parameters and spatial calendar maps of the Uttar Kannada district, on the west coast of India. They found deciduous forests experience earlier greenness onset than evergreen forests, which have longer grown seasons. Meteorological factors, especially maximum temperature and rainfall, significantly influence forest tree phenology. The majority of satellite sensor-derived phenology studies have used the NDVI calculated from a variety of sensors, including the IRS WiFS (Joshi et al 2006), MODIS (Chakraborty et al 2018;), SPOT-VGT, and the NOAA-AVHRR (Prasad et al 2008). Nevertheless, NDVI has limitations such as saturation at high vegetation biomass and atmospheric influence. The MERIS Terrestrial Chlorophyll Index (MTCI) from the Medium Resolution Imaging Spectrometer (MERIS) sensor offers a potential solution (Dash and Curran 2007). MERIS, with 15 spectral bands and 300 m spatial resolution, making it a promising tool for extracting canopy chlorophyll content data.

The ENVISAT mission's programmable spectrometer MERIS, which operates in the solar reflective spectral band, was utilized to carry out India's first phenology study. The

study captured foliage onset correlating with latitude, with minor inter-annual differences except for 2003 (Dash et al 2010). The study's limitations posed challenges in pinpointing the specific month, week, and date of leaf flushing and leaf fall based on qualitative descriptions (e.g., spring season, early dry period, post-monsoon period). Additionally, it was not possible to link flowering or bud burst data to the MERIS calculations (Atkinson et al 2012).

Jeganathan et al (2010) used the MTCI data, obtained from the Multitemporal MERIS, to investigate the phenology of four significant forest types in India, namely moist deciduous, dry deciduous, evergreen, and semi-evergreen. Their results indicated that the most frequent dates for the onset of greenness (OG) activity for tropical evergreen, semievergreen, wet deciduous, and dry deciduous vegetation types were February-April, January-April, March-May, and February-May, respectively. Additionally, the study found that the most frequent dates for end of senescence (ES) activity for tropical evergreen, semi-evergreen, wet deciduous, and dry deciduous vegetation types were February-April, January-April, February-April, and December-April, respectively (Jeganathan et al 2014).

Optical remote sensing faces challenges like atmospheric interference and cloud contamination. Trade-offs exist between spatial, temporal, and spectral resolutions. Various satellites offer different resolutions and revisit times, each with strengths and limitations for phenology studies (Helman 2018, Misra et al 2018). Recent advancements in satellite technology have improved sensing quality, making it crucial for phenology studies. However, challenges persist, including data acquisition costs, lack of ground-based validation, and processing expertise (Zhang et al 2016). There's a scarcity of ground-based observations and PhenoCams in forests, especially in the southern hemisphere (Singh et al 2020b). Efforts are underway in India to understand plant patterns and their connections to phenology and ecosystem models. Integrating PhenoCam data with Dynamic Global Vegetation Models (DGVMs) could improve local-level predictions and management strategies for climate change impacts on forest ecosystems (Jose et al 2023, Wang et al 2020).

#### **Objective of Studying Vegetation Phenology**

Three major categories-characterization, explanation, and application-can be used to group together the various research goals for studying vegetation phenology (Caparros Santiago et al 2021). Studies that emphasize spatial or spatiotemporal descriptions of vegetation phenology are included in the characterization process. Numerous researchers have systematically examined the spatial heterogeneity of phenological vegetation dynamics across diverse scales. This exploration often involves the utilization of mean or median phenometrics derived from satellite data, such as MODIS-NDVI, spanning the study duration (Deka et al 2019, Ayushi et al 2022). Notably, studies conducted in India have revealed that, compared to their evergreen counterparts, deciduous forests show an earlier onset of greenness (Singh et al 2020a). Furthermore, several studies conducted in India have analyzed the year, to year variations specifically focusing on the changes in timing for spring and autumn events (Singh and Sahoo 2019, Negi et al 2022). These investigations utilized sensing data with spatial resolution and have significantly contributed to a deeper understanding of the intricate relationship between vegetation phenology metrics and climatic effects across different types of forests in India. The findings from these studies shed light on the patterns observed in distinct forest ecosystems providing valuable insights into how climatic fluctuations impact vegetation dynamics in the region. This category of research on vegetation phenology also includes investigations into the potential effects of various methodological concerns (Atkinson et al 2012) on phenological estimation, the creation of phenological products (Ganguly et al 2010), or novel techniques that might make it possible to characterize the phenological behavior of vegetation.

The second category of studies includes those whose purpose was to analyze vegetation dynamics and how they relate to environmental conditions (Khare et al 2016, Reddy and Prasad 2018). Numerous investigations looked into the reactions of plant biological cycles to various climate variables. Temperature plays a crucial role in regulating phenological changes, especially in the Dry Deciduous and Temperate zone forests of India, influencing the timing of the growing season onset, duration, and foliage fall (Joshi and Janarthanam 2004). Conversely, water availability and precipitation patterns are key determinants of phenology in tropical forests. Experimental studies linking environmental temperature to plant phenology consistently show that higher temperatures lead to earlier spring and delayed autumn, while lower temperatures result in delayed spring and accelerated autumn. Photoperiod, in conjunction with low temperatures, contributes to deciduous tree leaf fall from November to February. Although some studies explore the relationship between vegetation phenology and variables like the carbon cycle, a significant portion of research in this domain focuses on phenological dynamics and its correlation with various factors (Kale and Roy 2012).

The third category encompasses research utilizing phenometrics as environmental data for diverse applications; the primary objective was not merely to comprehend the phenological behavior of vegetation, but also to explain it. Most of this research primarily employed phenological data to categorize different types of land cover (Joshi et al 2006). Phenology-based investigations have significantly enhanced the classification of land covers, thereby advancing the comprehension of Earth's surface land cover maps (Qader et al 2016). These studies have proven instrumental in identifying potential changes in land cover, including agricultural expansion (Knauer et al 2017) and deforestation (Valderrama - Landeros et al 2016). This underscores the pivotal role of phenology in monitoring and detecting alterations in land cover, contributing valuable insights to our understanding of the Earth's surface dynamics.

# Limitations of the Optical Datasets in Long Term Phenology Studies

The potential for Satellite Remote Sensing to enhance natural resource management is significant, despite certain constraints (Sharma et al 2021). Indian forests, with their rich biodiversity, provide an ideal context for remote sensingbased phenological studies. Recent research has demonstrated the utility of multi-temporal remote sensing data in tracking vegetation phenology relative to climate changes (Zhang et al 2006, Jeganathan et al 2014). Advances in technology have enabled more refined phenology retrieval at higher spatiotemporal resolutions (Ma et al 2022). While precise phenological event detection via satellite data is challenging, broader "land surface phenology (LSP)" descriptors are derived (De Beurs et al 2004, Richardson et al 2013). LSP overcomes limitations of ground-based observations, connecting traditional records with global models (Garonna et al 2018) and offering insights into Indian forest ecosystems (Jeganathan et al 2010). As of now, satellites are the sole practical means of maintaining a constant eye on Earth's dynamics at local to global scales. Specifically, when using optical datasets for long-term phenology studies we encounter some challenges (Berra et al 2019). Both species-specific phenology and vegetation phenology detection using satellite imagery still face a number of obstacles. The main obstacles arise from issues such as pixels, imprecise ground control points, atmospheric distortion and unclear definitions of ground objects (Sharma et al 2022). These factors ultimately affect the accuracy and clarity of the results obtained. These constraints persist across diverse image types, sensors, and platforms, including multi-spectral (hyper-spectral) images, high spatial resolution images, and LiDAR images (Prabakaran et al 2013). Researchers have employed techniques like maximum value compositing (MVC) and masking to address cloud impacts, though these methods have limitations (Dash et al 2010, Atkinson et al 2012, Chakraborty et al 2018).

Ground-based phenological studies in India typically

observe nine plant development stages, which include leaves (Bud stage, mature, abscission), flowers (Bud stage, anthesis, abscission) and fruit (Bud stage, maturation, and abscission), but accurately monitoring all these stages via satellite data remains challenging. Moreover, there is a scarcity of ground-based observations in developing and underdeveloped countries that lack established networks for phenological monitoring. To address this gap, expanding the reach of PhenoCam networks to presently underrepresented regions hold the potential to enhance the comprehensive monitoring of vegetation phenology globally. Such an expansion could offer a more holistic perspective on the ramifications of climate change and contribute to a deeper understanding of ecological processes on a global scale (Jose et al 2023).

#### CONCLUSION

The review has highlighted several critical areas that warrant further attention to maximize the potential of satellite remote sensing in supporting forest phenological studies in India. The key observations are summarized as follows:

- The immense potential of satellite remote sensing data to support forest phenological studies is evident, but significant challenges remain that must be addressed.
- Expanding the spatial scale and improving the spatiotemporal resolution of phenological monitoring is critical for accurately assessing the impacts of environmental changes across diverse forest ecosystems.
- 3. The limited availability of long-term phenology datasets hinders the understanding of temporal patterns and trends, while the inherent complexity of these ecosystems poses analytical challenges.
- Integrating in-situ data from local experts with remote sensing analysis is essential to maximize the value and application of satellite data for phenological research.
- 5. Combining SAR and optical satellite data can provide a more comprehensive dataset to enhance the understanding of woody vegetation dynamics.
- Exploring alternative vegetation indices beyond the commonly used NDVI and EVI can offer valuable insights into phenological changes.
- 7. Strengthening interdisciplinary collaboration among remote sensing experts, conservationists, and environmental managers is crucial for shaping a unified research agenda and driving more effective outcomes.
- Adopting emerging satellite technologies and innovative analytical methods, including machine learning, can advance the mechanistic understanding of phenological processes across multiple scales.
  - By strategically addressing these key focus areas, the

research community can significantly enhance the accuracy, reliability, and comprehensive application of satellite remote sensing in support of forest phenological studies in India, ultimately driving more impactful and informed decisionmaking for the sustainable management of these critical ecosystems.

#### ACKNOWLEDGMENTS

The first author is thankful to DST, New Delhi, for the INSPIRE fellowship. The authors are thankful to Shri. N.M. Desai, Director, Space Applications Centre (SAC), ISRO for his constant encouragement and guidance.

#### **AUTHOR'S CONTRIBUTIONS**

Dhruvi Sedha: Writing-Original Draft preparation, Conceptualization, Methodology, Figure Preparation and Editing. Chandra Prakash Singh: Conceptualization, Formal Analysis, Supervision, Reviewing and Editing. Hitesh Solanki: Supervision. Jincy Rachel Mathew: Visualization and Editing. Mehul R Pandya: Supervision and reviewing. Bimal Bhattacharya: Resources and Supervision. C Jeganathan: Reviewing and Editing. Siddhartha Khare: Reviewing and Editing. All authors have read and agreed to the published version of the manuscript.

#### REFERENCES

- Alemu WG and Henebry GM 2017. Land surface phenology and seasonality using cool earthlight in croplands of eastern Africa and the linkages to crop production. *Remote Sensing* **9**(9): 914.
- Atkinson PM, Jeganathan C, Dash J and Atzberger C 2012. Intercomparison of four models for smoothing satellite sensor timeseries data to estimate vegetation phenology. *Remote Sensing of Environment* **123**: 400-417.
- Ayushi K, Babu KN, Reddy CS, Mayamanikandan T, Barathan N, Debabrata B and Ayyappan N 2022. Remote sensing-based characterisation of community level phenological variations in a regional forest landscape of Western Ghats, India. *Geocarto International* 37(27): 16620-16635.
- Bajpai O, Kumar A, Mishra AK, Sahu N, Behera SK and Chaudhary LB 2012. Phenological Study of Two Dominant Tree Species in Tropical. *International Journal of Botany* 8(2): 66-72.
- Baruah U and Ramakrishnan P 1989a. Leaf dynamics of early versus late successional shrubs of sub-tropical moist forests of north-eastern India, pp. 431-436. In: <u>Proceedings: Plant</u> <u>Sciences 99</u>, Indian Academy of Sciences.
- Baruah U and Ramakrishnan P 1989b. Phenology of the shrub strata of successional sub-tropical humid forests of north-eastern India. *Vegetatio* **80**: 63-67.
- Berra EF, Gaulton R and Barr S 2019. Assessing spring phenology of a temperate woodland: A multiscale comparison of ground, unmanned aerial vehicle and Landsat satellite observations. *Remote Sensing of Environment* **223**: 229-242.
- Bhat D 1992. Phenology of tree species of tropical moist forest of Uttara Kannada district, Karnataka, India. *Journal of Biosciences* 17: 325-352.
- Bhat D and Murali K 2001. Phenology of understorey species of tropical moist forest of Western Ghats region of Uttara Kannada district in South India. *Current Science* 799-805.
- Bhat D, Naik M, Patagar S, Hegde G, Kanade Y, Hegde G, Shastri C,

Shetti D and Furtado R 2000. Forest dynamics in tropical rain forests of Uttara Kannada district in Western Ghats, India. *Current Science* 975-985.

- Bhat PR and Kaveriappa K 2009. Ecological studies on myristica swamp forests of Uttara Kannada, Karnataka, India. *Tropical Ecology* **50**(2): 329.
- Bison NN, Partelli-Feltrin R and Michaletz ST 2022. Trait phenology and fire seasonality co-drive seasonal variation in fire effects on tree crowns. *New Phytologist* **234**(5): 1654-1663.
- Boojh R and Ramakrishnan P 1983. The growth pattern of two species of Schima. *Biotropica* 142-147.
- Borah M and Devi A 2014. Phenology, growth and survival of *Vatica lanceaefolia* Bl.: A critically endangered tree species in moist tropical forest of Northeast India. *Tropical Plant Research* 1(3): 1-12.
- Brown LA, Dash J, Ogutu BO and Richardson AD 2017. On the relationship between continuous measures of canopy greenness derived using near-surface remote sensing and satellite-derived vegetation products. Agricultural and Forest Meteorology 247: 280-292.
- Caparros-Santiago JA, Rodriguez-Galiano V and Dash J 2021. Land surface phenology as indicator of global terrestrial ecosystem dynamics: A systematic review. *ISPRS Journal of Photogrammetry and Remote Sensing* **171**: 330-347.
- Chakraborty A, Seshasai M, Reddy CS and Dadhwal V 2018. Persistent negative changes in seasonal greenness over different forest types of India using MODIS time series NDVI data (2001–2014). *Ecological Indicators* **85**: 887-903.
- Champion HG 1936. A preliminary survey of forest types of India and Burma. Indian Forestry Record (NS) Silviculture 1: 1.
- Chandra S, Singh A, Mathew JR, Singh C, Pandya MR, Bhattacharya BK, Solanki H, Nautiyal M and Joshi R 2022. Phenocam observed flowering anomaly of *Rhododendron arboreum* Sm. in Himalaya: a climate change impact perspective. *Environmental Monitoring and Assessment* **194**: 877.
- Chang Q, Novick K, Ficklin D and Jiao W 2021. A phenology-based improvement for early agricultural drought detection. In: AGU Fall Meeting Abstracts, pp B13B-02.
- Chatterjee D 1939. Studies on the endemic flora of India and Burma. Annexe Thesis Digitisation Project 2018 Block 18.
- Chaturvedi R K, Gopalakrishnan R, Jayaraman M, Bala G, Joshi N, Sukumar R and Ravindranath N 2011. Impact of climate change on Indian forests: A dynamic vegetation modeling approach. *Mitigation and Adaptation Strategies for Global Change* **16**: 119-142.
- Chauhan SVS and Chauhan S 2020. Climate changes and trends in phenology of Cassia fistula L. in Agra (India)-1965-2019. *Journal of Native and Alien Plant Studies* **16**: 23-31.
- Chéret V and Denux J-P 2011. Analysis of MODIS NDVI time series to calculate indicators of Mediterranean forest fire susceptibility. *GIScience & Remote Sensing* **48**: 171-194.
- Ciais P, Sabine C, Bala G, Bopp L, Brovkin V, Canadell J, Chhabra A, DeFries R, Galloway J and Heimann M 2014. Carbon and other biogeochemical cycles. In: Climate change 2013: the physical science basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, pp 465-570.
- Cleland E E, Chuine I, Menzel A, Mooney HA and Schwartz MD 2007. Shifting plant phenology in response to global change. *Trends in Ecology & Evolution* **22**: 357-365.
- Czernecki B, Nowosad J and Jabłońska K 2018. Machine learning modeling of plant phenology based on coupling satellite and gridded meteorological dataset. *International Journal of Biometeorology* 62: 1297-1309.
- Dash J and Curran P 2007. Evaluation of the MERIS terrestrial chlorophyll index (MTCI). *Advances in Space Research* **39**: 100-104.

- Dash J, Jeganathan C and Atkinson P 2010. The use of MERIS Terrestrial Chlorophyll Index to study spatio-temporal variation in vegetation phenology over India. *Remote Sensing of Environment* **114**: 1388-1402.
- Datta A and Rane A 2013. Phenology, seed dispersal and regeneration patterns of Horsfieldia kingii, a rare wild nutmeg. *Tropical Conservation Science* **6**:674-689.
- De Beurs KM and Henebry GM 2004. Land surface phenology, climatic variation, and institutional change: Analyzing agricultural land cover change in Kazakhstan. *Remote Sensing* of Environment 89: 497-509.
- de Beurs KM and Henebry GM 2008. Northern annular mode effects on the land surface phenologies of northern Eurasia. *Journal of Climate* 21: 4257-4279.
- Deka J, Kalita S and Khan ML 2019. Vegetation phenological characterization of alluvial plain Shorea robusta-dominated tropical moist deciduous forest of northeast India using MODIS NDVI time series data. *Journal of the Indian Society of Remote Sensing* 47: 1287-1293.
- Devi NL, Singha D and Tripathi SK 2020. Phenology, population structure and carbon sequestration potential of Parkia timoriana: a heirloom tree in traditional Meitei homegarden of northeast India. *Vegetos* **33**: 222-228.
- Dronova I and Taddeo S 2022. Remote sensing of phenology: Towards the comprehensive indicators of plant community dynamics from species to regional scales. *Journal of Ecology* **110**: 1460-1484.
- Ganesh T and Davidar P 1997. Flowering phenology and flower predation of Cullenia exarillata (Bombacaceae) by arboreal vertebrates in Western Ghats, India. *Journal of Tropical Ecology* 13: 459-468.
- Ganguly S, Friedl MA, Tan B, Zhang X and Verma M 2010. Land surface phenology from MODIS: Characterization of the Collection 5 global land cover dynamics product. *Remote Sensing of Environment* **114**: 1805-1816.
- Gao F and Zhang X 2021. Mapping crop phenology in near real-time using satellite remote sensing: Challenges and opportunities. *Journal of Remote Sensing* **2021**(2): 1-14.
- Garonna I, de Jong R, Stöckli R, Schmid B, Schenkel D, Schimel D and Schaepman ME 2018. Shifting relative importance of climatic constraints on land surface phenology. *Environmental Research Letters* **13**: 024025.
- Ge Q, Wang H and Dai J 2013. Shifts in spring phenophases, frost events and frost risk for woody plants in temperate China. *Climate Research* **57**: 249-258.
- Ghosh S, Nandy S, Mohanty S, Subba R and Kushwaha S 2019. Are phenological variations in natural teak (Tectona grandis) forests of India governed by rainfall? A remote sensing based investigation. *Environmental Monitoring and Assessment* **191**: 1-10.
- Glenn EP, Huete AR, Nagler PL and Nelson SG 2008. Relationship between remotely-sensed vegetation indices, canopy attributes and plant physiological processes: What vegetation indices can and cannot tell us about the landscape. Sensors 8: 2136-2160.
- Gray JM, Frolking S, Kort EA, Ray DK, Kucharik CJ, Ramankutty N and Friedl MA 2014. Direct human influence on atmospheric CO2 seasonality from increased cropland productivity. *Nature* 515: 398-401.
- Gupta U 2014. Study of temporal variation of vegetation indices and phenology of tropical deciduous broadleaf forest in eastern India. The International Archives of the Photogrammetry, Remote Sensing and Spatial Information Sciences 40: 569-572.
- Hänninen H 2006. Climate warming and the risk of frost damage to boreal forest trees: Identification of critical ecophysiological traits. *Tree Physiology* 26(7): 889-898.
- Haq SM, Hamid M, Lone FA and Singh B 2021. Himalayan Hotspot with Alien Weeds: A Case Study of Biological Spectrum, Phenology, and Diversity of Weedy Plants of High Altitude Mountains in District Kupwara of J&K Himalaya, India, pp. 139-

152. In: Proceedings of the National Academy of Sciences, India Section B: Biological Sciences **91**, National Academy of Sciences, India.

- Helman D 2018. Land surface phenology: What do we really 'see' from space? Science of the Total Environment 618: 665-673.
- Henebry GM and de Beurs KM 2013. Remote Sensing of Land Surface Phenology: A Prospectus, pp. 385-411. In: Schwartz M (ed), *Phenology: An Integrative Environmental Science*, Springer, Dordrecht.
- Heumann BW, Seaquist J, Eklundh L and Jönsson P 2007. AVHRR derived phenological change in the Sahel and Soudan, Africa, 1982–2005. *Remote Sensing of Environment* **108**(4): 385-392.
- Jadeja B and Nakar R 2010. Phenological studies of some tree species from Girnar reserve forest Gujarat India. *Plant Archives* 10(2): 825-828.
- Jeganathan C, Dash J and Atkinson PM 2010. Mapping the phenology of natural vegetation in India using a remote sensingderived chlorophyll index. *International Journal of Remote Sensing* **31**(22): 5777-5796.
- Jeganathan C, Dash J and Atkinson P 2014. Remotely sensed trends in the phenology of northern high latitude terrestrial vegetation, controlling for land cover change and vegetation type. *Remote Sensing of Environment* **143**: 154-170.
- Jiang DH, Huang S and Han D 2014. Monitoring and modeling terrestrial ecosystems' response to climate change. Advances in Meteorology 2016: 5984595.
- Jin H, Jönsson A M, Bolmgren K, Langvall O and Eklundh L 2017. Disentangling remotely-sensed plant phenology and snow seasonality at northern Europe using MODIS and the plant phenology index. *Remote Sensing of Environment* **198**: 203-212.
- Jose K, Chaturvedi RK, Jeganathan C, Behera MD and Singh CP 2023. Plugging the gaps in the global PhenoCam monitoring of forests-The need for a PhenoCam Network across Indian Forests. *Remote Sensing* 15(24): 5642.
- Joshi P, Singh S, Agarwal S and Roy P 2001. Land cover assessment in Jammu & Kashmir using phenology as discriminant: An approach of wide swath satellite (IRS–WiFS). *Current Science* **81**(4): 392-399.
- Joshi PK, Roy P, Singh S, Agrawal S and Yadav D 2006. Vegetation cover mapping in India using multi-temporal IRS Wide Field Sensor (WiFS) data. *Remote Sensing of Environment* **103**(2): 190-202.
- Joshi VC and Janarthanam M 2004. The diversity of life-form type, habitat preference and phenology of the endemics in the Goa region of the Western Ghats, India. *Journal of Biogeography* **31**(8): 1227-1237.
- Kale MP and Roy P 2012. Net primary productivity estimation and its relationship with tree diversity for tropical dry deciduous forests of central India. *Biodiversity and Conservation* **21**: 1199-1214.
- Khare S, Latifi H and Ghosh K 2016. Phenology analysis of forest vegetation to environmental variables during pre-and postmonsoon seasons in western Himalayan region of India, pp. 15-19. In: International Archives of Photogrammetry, Remote Sensing and Spatial Information Sciences, Vol. 41.
- Khare S, Ghosh SK, Latifi H, Vijay S and Dahms T 2017. Seasonalbased analysis of vegetation response to environmental variables in the mountainous forests of Western Himalaya using Landsat 8 data. *International Journal of Remote Sensing* **38**(15): 4418-4442.
- Khare S and Rossi S 2019. Phenology analysis of moist decedous forest using time series Landsat-8 remote sensing data, pp. 127-131. In: Proceedings of IEEE International Workshop on Metrology for Agriculture and Forestry (MetroAgriFor), October 24-26, 2019, Portici, Italy.
- Kikim A and Yadava P 2001. Phenology of tree species in subtropical forests of Manipur in north eastern India. *Tropical Ecology* **42**(2): 269-276.

- Knauer K, Gessner U, Fensholt R, Forkuor G and Kuenzer C 2017. Monitoring agricultural expansion in Burkina Faso over 14 years with 30 m resolution time series: The role of population growth and implications for the environment. *Remote Sensing* **9**(2): 132.
- Krishna Prasad V, Badarinath K and Eaturu A 2007. Spatial patterns of vegetation phenology metrics and related climatic controls of eight contrasting forest types in India–analysis from remote sensing datasets. *Theoretical and Applied Climatology* 89: 95-107.
- Krishnan RM 2002. Reproductive phenology of a wet forest understorey in the Western Ghats, South India. *Global Ecology and Biogeography* **11**(2): 179-182.
- Kumar R and Shahabuddin G 2005. Effects of biomass extraction on vegetation structure, diversity and composition of forests in Sariska Tiger Reserve, India. *Environmental Conservation* 32(3): 1-12.
- Kushwaha C and Singh K 2005. Diversity of leaf phenology in a tropical deciduous forest in India. *Journal of Tropical Ecology* 21(1):47-56.
- Kushwaha C and Singh K 2008. India needs phenological stations network. *Current Science* **95**(7): 832-834.
- Lambert J, Drenou C, Denux J-P, Balent G and Cheret V 2013. Monitoring forest decline through remote sensing time series analysis. GIScience & Remote Sensing 50(4): 437-457.
- Li X, Zhou Y, Meng L, Asrar G R, Lu C and Wu Q 2019. A dataset of 30 m annual vegetation phenology indicators (1985–2015) in urban areas of the conterminous United States. *Earth System Science Data* **11**(2): 881-894.
- Lobell DB, Burke MB, Tebaldi C, Mastrandrea MD, Falcon WP and Naylor RL 2008. Prioritizing climate change adaptation needs for food security in 2030. *Science* **319**(5863): 607-610.
- Ma X, Zhu X, Xie Q, Jin J, Zhou Y, Luo Y, Liu Y, Tian J and Zhao Y 2022. Monitoring nature's calendar from space: Emerging topics in land surface phenology and associated opportunities for science applications. *Global Change Biology* 28(24): 7186-7204.
- Malhi RKM, Kiran GS, Shah MN, Mistry NV, Bhavsar VH, Singh CP, Bhattarcharya BK, Townsend PA and Mohan S 2021. Applicability of smoothing techniques in generation of phenological metrics of *Tectona grandis* L. using NDVI time series data. *Remote Sensing* **13**(17): 3343.
- Mishra R, Upadhyay V, Bal S, Mohapatra P and Mohanty R 2006. Phenology of species of moist deciduous forest sites of Similipal biosphere reserve. *Lyonia* **11**(1): 5-17.
- Misra G, Buras A, Heurich M, Asam S and Menzel A 2018. LiDAR derived topography and forest stand characteristics largely explain the spatial variability observed in MODIS land surface phenology. *Remote Sensing of Environment* **218**: 231-244.
- Misra R 1946. A study in the ecology of low-lying lands. Indian Ecologist 1:27-46.
- Mohandass D, Hughes AC and Davidar P 2016. Flowering and fruiting patterns of woody species in the tropical montane evergreen forest of southern India. *Current Science* **110**: 404-416.
- Mohapatra J, Singh CP, Tripathi OP and Pandya HA 2019. Remote sensing of alpine treeline ecotone dynamics and phenology in Arunachal Pradesh Himalaya. *International Journal of Remote Sensing* **40**(20): 7986-8009.
- Morellato LPC, Alberton B, Alvarado ST, Borges B, Buisson E, Camargo MGG, Cancian LF, Carstensen DW, Escobar DF and Leite PT 2016. Linking plant phenology to conservation biology. *Biological Conservation* 195: 60-72.
- Morisette JT, Richardson AD, Knapp AK, Fisher JI, Graham EA, Abatzoglou J, Wilson BE, Breshears DD, Henebry GM and Hanes JM 2009. Tracking the rhythm of the seasons in the face of global change: Phenological research in the 21st century. *Frontiers in Ecology and the Environment* 7(5): 253-260.

Murali K and Sukumar R 1993. Leaf flushing phenology and

herbivory in a tropical dry deciduous forest, southern India. *Oecologia* **94**: 114-119.

- Murali K and Sukumar R 1994. Reproductive phenology of a tropical dry forest in Mudumalai, southern India. *Journal of Ecology* 82: 759-767.
- Nanda A, Prakasha H, Murthy YK and Suresh H 2011. Phenology of leaf flushing, flower initiation and fruit maturation in dry deciduous and evergreen forests of Bhadra Wildlife Sanctuary, Karnataka, southern India. Our Nature 9(1): 89-99.
- Nanda A, Suresh HS and Krishnamurthy YL 2014. Phenology of a tropical dry deciduous forest of Bhadra wildlife sanctuary, southern India. *Ecological Processes* 3: 1-12.
- Nanda A, Suresh H and Krishna Murthy Y 2015. Leafing phenology of tropical forests of Bhadra wildlife sanctuary, Karnataka, India. *Applied Science Reports* **12**(1): 33-40.
- Nanda A, Prakasha HM and Murthy YLK 2017. Leafing patterns and seasonality at community level in a tropical dry deciduous forests of Bhadra Wildlife Sanctuary, Karnataka, Southern India. *Tree and Forestry Science and Biotechnology* 82-87.
- Nandy S, Ghosh S and Singh S 2021. Assessment of sal (Shorea robusta) forest phenology and its response to climatic variables in India. *Environmental Monitoring and Assessment* **193**(9): 616.
- Nath S, Nath AJ and Das AK 2016. Vegetative and reproductive phenology of a floodplain tree species Barringtonia acutangula from North East India. *Journal of Environmental Biology* **37**(2): 215.
- Nautiyal M, Nautiyal B and Prakash V 2001. Phenology and growth form distribution in an alpine pasture at Tungnath, Garhwal, Himalaya. *Mountain Research and Development* **21**(2): 168-174.
- Negi G, Joshi S, Singh P and Joshi R 2022. Phenological response patterns of forest communities to annual weather variability at long-term ecological monitoring sites in Western Himalaya. *Trees, Forests and People* **8**: 100237.
- Newton PN 1988. The structure and phenology of a moist deciduous forest in the Central Indian Highlands. *Vegetatio* **75**: 3-16.
- Pastick NJ, Wylie BK and Wu Z 2018. Spatiotemporal analysis of Landsat-8 and Sentinel-2 data to support monitoring of dryland ecosystems. *Remote Sensing* **10**(5): 791.
- Pastor-Guzman J, Dash J and Atkinson PM 2018. Remote sensing of mangrove forest phenology and its environmental drivers. *Remote Sensing of Environment* **205**: 71-84.
- Pau S, Wolkovich EM, Cook BI, Davies TJ, Kraft NJ, Bolmgren K, Betancourt JL and Cleland EE 2011. Predicting phenology by integrating ecology, evolution and climate science. *Global Change Biology* **17**(12): 3633-3643.
- Paul A, Khan ML and Das AK 2018. Phenological characteristics of Rhododendron species in temperate mixed broad-leaved forests of Arunachal Himalaya, India. *Journal of Forest and Environmental Science* 34(6): 435-450.
- Piao S, Liu Q, Chen A, Janssens IA, Fu Y, Dai J, Liu L, Lian X, Shen M and Zhu X 2019. Plant phenology and global climate change: Current progresses and challenges. *Global Change Biology* 25(6): 1922-1940.
- Pillai ND, Nandy S, Patel N, Srinet R, Watham T and Chauhan P 2019. Integration of eddy covariance and process-based model for the intra-annual variability of carbon fluxes in an Indian tropical forest. *Biodiversity and Conservation* 28: 2123-2141.
- Prabakaran C, Singh C, Panigrahy S and Parihar J 2013. Retrieval of forest phenological parameters from remote sensing-based NDVI time-series data. *Current Science* **105**(6): 795-802.
- Prasad SN and Hegde M 1986. Phenology and seasonality in the tropical deciduous forest of Bandipur, South India. Proceedings: *Plant Sciences* **96**: 121-133.
- Prasad VK, Badarinath K and Eaturu A 2008. Effects of precipitation, temperature and topographic parameters on evergreen vegetation greenery in the Western Ghats, India. International

Journal of Climatology: A Journal of the Royal Meteorological Society **28**(13): 1807-1819.

- Puri G 1958. Problems in the ecology of the humid tropics. Study of the Tropical Vegetation: 19-27.
- Qader SH, Dash J, Atkinson PM and Rodriguez-Galiano V 2016. Classification of vegetation type in Iraq using satellite-based phenological parameters. *IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing* **9**(1):414-424.
- Radeloff V, Dubinin M, Coops N, Allen A, Brooks T, Clayton M, Costa G, Graham C, Helmers D and Ives A 2019. The dynamic habitat indices (DHIs) from MODIS and global biodiversity. *Remote Sensing of Environment* **222**: 204-214.
- Ralhan P, Khanna R, Singh S and Singh J 1985. Phenological characteristics of the tree layer of Kumaun Himalayan forests. *Vegetatio* **60**: 91-101.
- Ranjan AK and Gorai AK 2022. Evaluating phenological trends of different vegetation types in response to climate change over the Rajmahal Hills in India during 2001-2019. *Remote Sensing Letters* **13**(9):898-911.
- Reddy DS and Prasad PRC 2018. Prediction of vegetation dynamics using NDVI time series data and LSTM. *Modeling Earth Systems* and Environment **4**: 409-419.
- Reed BC, Brown JF, VanderZee D, Loveland TR, Merchant JW and Ohlen DO 1994. Measuring phenological variability from satellite imagery. *Journal of Vegetation Science* **5**(5): 703-714.
- Richardson AD, Anderson RS, Arain MA, Barr AG, Bohrer G, Chen G, Chen JM, Ciais P, Davis KJ and Desai AR 2012. Terrestrial biosphere models need better representation of vegetation phenology: results from the North American Carbon Program Site Synthesis. *Global Change Biology* **18**(2): 566-584.
- Richardson AD, Keenan TF, Migliavacca M, Ryu Y, Sonnentag O and Toomey M 2013. Climate change, phenology, and phenological control of vegetation feedbacks to the climate system. *Agricultural and Forest Meteorology* **169**: 156-173.
- Rosenzweig C, Casassa G, Karoly DJ, Imeson A, Liu C, Menzel A, Rawlins S, Root TL, Seguin B and Tryjanowski P 2007. Assessment of observed changes and responses in natural and managed systems. DOI: 10.5167/uzh-33180.
- Rosenzweig C, Karoly D, Vicarelli M, Neofotis P, Wu Q, Casassa G, Menzel A, Root TL, Estrella N and Seguin B 2008. Attributing physical and biological impacts to anthropogenic climate change. *Nature* **453**(7193): 353-357.
- Roy P and Joshi P 2002. Forest cover assessment in north-east India--the potential of temporal wide swath satellite sensor data (IRS-1C WiFS). *International Journal of Remote Sensing* **23**(22):4881-4896.
- Shankar U 2001. A case of high tree diversity in a sal (Shorea robusta)-dominated lowland forest of Eastern Himalaya: Floristic composition, regeneration and conservation. Current Science: 81(7): 776-786.
- Sharma S, Richardson DC, Woolway RI, Imrit MA, Bouffard D, Blagrave K, Daly J, Filazzola A, Granin N and Korhonen J 2021. Loss of ice cover, shifting phenology, and more extreme events in Northern Hemisphere lakes. *Journal of Geophysical Research: Biogeosciences* **126**(10): e2021JG006348.
- Sharma S, Filazzola A, Nguyen T, Imrit MA, Blagrave K, Bouffard D, Daly J, Feldman H, Feldsine N and Hendricks-Franssen H-J 2022. Long-term ice phenology records spanning up to 578 years for 78 lakes around the Northern Hemisphere. *Scientific Data* 9(1): 318.
- Shukla R and Ramakrishnan P 1982. Phenology of trees in a subtropical humid forest in north-eastern India. *Vegetatio* **49**(2): 103-109.
- Singh B, Jeganathan C and Rathore V 2020a. Improved NDVI based proxy leaf-fall indicator to assess rainfall sensitivity of deciduousness in the central Indian forests through remote sensing. *Scientific Reports* **10**(1): 17638.

Singh K and Kushwaha C 2005. Paradox of leaf phenology: Shorea

robusta is a semi-evergreen species in tropical dry deciduous forests in India. *Current Science*: 1820-1824.

- Singh LA, Whittecar WR, DiPrinzio MD, Herman JD, Ferringer MP and Reed PM 2020b. Low cost satellite constellations for nearly continuous global coverage. *Nature Communications* **11**(1): 200.
- Singh S, Agarwal S, Joshi P and Roy P 1999. Biome level classification of vegetation in western India: An application of wide field view sensor (WiFS). In: *Joint Workshop of ISPRS Working Groups* I/1, I/3 and IV/4: Sensors and Mapping from Space, Hanover (Germany), pp 27-30.
- Singh SL and Sahoo UK 2019. Shift in phenology of some dominant tree species due to climate change in Mizoram, North-East India. *Indian Journal of Ecology* **46**(1): 132-136.
- Sivaperuman C and Venkataraman K 2018. Indian Hotspots: Vertebrate Faunal Diversity, *Conservation and Management* Volume 2. Springer.
- Subin K, Jose P and Vivek A 2022. Rarity analysis of an endangered tropical tree species of the Western Ghats, India. *Current Science* **123**(1): 67-72.
- Sundarapandian S, Chandrasekaran S and Swamy P 2005. Phenological behaviour of selected tree species in tropical forests at Kodayar in the Western Ghats, Tamil Nadu, India. *Current Science* **88**(5):805-810.
- Suresh H and Sukumar R 2018. Phenology of Ficus spp. in a tropical dry forest, Mudumalai, south India. *Journal of Forestry Research* 29: 1129-1138.
- Thackeray SJ, Henrys PA, Hemming D, Bell JR, Botham MS, Burthe S, Helaouet P, Johns DG, Jones ID and Leech DI 2016. Phenological sensitivity to climate across taxa and trophic levels. *Nature* **535**(7611): 241-245.
- Upadhaya K, Mir AH and Iralu V 2018. Reproductive phenology and germination behavior of some important tree species of Northeast India. In: Proceedings of the National Academy of Sciences, India Section B: Biological Sciences 88: 1033-1041.
- Vadrevu K, Badarinath K and Anuradha E 2007. Evaluation of vegetation greenness metrics for characterizing tropical forest cover types–a case study using NOAA AVHRR datasets. *Geocarto International* 22(1): 29-48.
- Valderrama-Landeros LH, España-Boquera ML and Baret F 2016. Deforestation in Michoacan, Mexico, from CYCLOPES-LAI time series (2000–2006). *IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing* **9**(12): 5398-5405.
- Vashistha RK, Rawat N, Chaturvedi AK, Nautiyal B, Prasad P and Nautiyal M 2009. An exploration on the phenology of different growth forms of an alpine expanse of North-West Himalaya, India. *New York Science Journal* **2**(6): 29-41.
- Walther G-R, Post E, Convey P, Menzel A, Parmesan C, Beebee TJ, Fromentin J-M, Hoegh-Guldberg O and Bairlein F 2002. Ecological responses to recent climate change. *Nature* **416**(6879): 389-395.
- Wang H, Jia G, Epstein HE, Zhao H and Zhang A 2020. Integrating a PhenoCam-derived vegetation index into a light use efficiency model to estimate daily gross primary production in a semi-arid grassland. Agricultural and Forest Meteorology 288: 107983.
- White K, Pontius J and Schaberg P 2014. Remote sensing of spring phenology in northeastern forests: A comparison of methods, field metrics and sources of uncertainty. *Remote Sensing of Environment* 148:97-107.
- White MA, de Beurs KM, Didan K, Inouye DW, Richardson AD, Jensen OP, O'keefe J, Zhang G, Nemani RR and van Leeuwen WJ 2009. Intercomparison, interpretation, and assessment of spring phenology in North America estimated from remote sensing for 1982–2006. *Global Change Biology* **15**(10): 2335-2359.
- Wingate L, Ogée J, Cremonese E, Filippa G, Mizunuma T, Migliavacca M, Moisy C, Wilkinson M, Moureaux C, Wohlfahrt G,

Hammerle A, Hörtnagl L, Gimeno C, Porcar-Castell A, Galvagno M, Nakaji T, Morison J, Kolle O, Knohl A, Kutsch W, Kolari P, Nikinmaa E, Ibrom A, Gielen B, Eugster W, Balzarolo M, Papale D, Klumpp K, Köstner B, Grünwald T, Joffre R, Ourcival JM, Hellstrom M, Lindroth A, George C, Longdoz B, Genty B, Levula J, Heinesch B, Sprintsin M, Yakir D, Manise T, Guyon D, Ahrends H, Plaza-Aguilar A, Guan JH and Grace J 2015. Interpreting canopy development and physiology using a European phenology camera network at flux sites. *Biogeosciences* **12**(20): 5995-6015.

- Wolkovich EM, Cook BI and Davies TJ 2014. Progress towards an interdisciplinary science of plant phenology: Building predictions across space, time and species diversity. *New Phytologist* **201**(4): 1156-1162.
- Yadav R and Yadav A 2008. Phenology of selected woody species in a tropical dry deciduous forest in Rajasthan, India. *Tropical Ecology* **49**(1): 25-34.
- Younes N, Joyce KE and Maier SW 2021. All models of satellitederived phenology are wrong, but some are useful: A case study

Received 02 July, 2024; Accepted 30 September, 2024

from northern Australia. International Journal of Applied Earth Observation and Geoinformation **97**: 102285.

- Zhang J, Hu J, Lian J, Fan Z, Ouyang X and Ye W 2016. Seeing the forest from drones: Testing the potential of lightweight drones as a tool for long-term forest monitoring. *Biological Conservation* **198**: 60-69.
- Zhang X, Friedl MA, Schaaf CB, Strahler AH, Hodges JC, Gao F, Reed BC and Huete A 2003. Monitoring vegetation phenology using MODIS. *Remote Sensing of Environment* 84(3): 471-475.
- Zhang X, Friedl MA and Schaaf CB 2006. Global vegetation phenology from Moderate Resolution Imaging Spectroradiometer (MODIS): Evaluation of global patterns and comparison with in situ measurements. *Journal of Geophysical Research: Biogeosciences* **111**(G4).
- Zheng W, Liu Y, Yang X and Fan W 2022. Spatiotemporal variations of forest vegetation phenology and its response to climate change in northeast China. *Remote Sensing* **14**(12): 2909.



Indian Journal of Ecology (2024) 51(5): 948-953 DOI: https://doi.org/10.55362/IJE/2024/4334 Manuscript Number: 4334 NAAS Rating: 5.38

# Carbon Sequestration Potential of Woody Species in Thiagarajar College Campus, Madurai, India

K. Saraswathi and P. Sneha

Department of Botany and National Centre of Excellence (MHRD) Thiagarajar College, Madurai-625 009, India E-mail: krish.saras31@gmail.com

**Abstract:** Thiagarajar College Campus had 56 tree species and 5 shrub species with a total of 502 individuals during 2023. Azadirachta indica had a maximum number of 109 individuals. Samanea saman had the highest biomass (12923.72 kg/tree), Carbon storage (6461.80 kg/tree) and carbon sequestration (23456.56 kg/tree and 2148.70 kg/tree). The other potential tree species on the campus with high carbon sequestration were *Wrightia tinctoria, Parkia benghalensis, Ficus biglandulosa* and *Terminalia catappa*. The carbon level (497-538 ppm CO<sub>2</sub>) in the surrounding environment was within the permissible limit due to the woody cover in the campus. The study recommends the potential tree species for disturbed and polluted urban space.

#### Keywords: Azadirachta indica, Samanea saman, Biomass, Carbon storage, CO<sub>2</sub> level

The heat-trapping gases released by human activities have become one of the great concerns of the world today as they cause global warming and its allied problems (https://www.epa.gov/climatechange-science/causes-climatechange). Carbon dioxide is the major greenhouse gas of global climate change (Dubal et al 2013). Around 87% of CO<sub>2</sub> is released from fossil fuel burning. The remaining is from forest clearing, industrial processes etc. Nearly 45% of CO<sub>2</sub> released during anthropogenic activities remains in the atmosphere.  $CO_2$  trapping is essential for reducing our carbon footprint.

Plants act as a major sink of carbon as sequester carbon through photosynthesis. The forests, grasslands or rangelands play a significant role in carbon sequestration. They store 25% of carbon in their biomass. Aquatic systems store another 25% of CO<sub>2</sub>. The bogs, peat and swamps can also store carbon as carbonate. Apart from that, carbon is sequestered in rocks through geological carbon sequestration (https://www.nationalgrid.com/stories/energyexplained/what-carbon-sequestration). The Kyoto Protocol and the United Nations Framework Convention on Climate Change gave directions to raise carbon sequestration. In the terrestrial landscape, trees play several important roles, especially carbon sequestration (Veeramani et al 2023). Trees offer several services for the sustenance of rural and urban ecosystems. They provide a significant contribution to climate change mitigation (Lahoti et al 2020). They trap heat and enable the purity of air, control the microclimate to regulate the urban heat island phenomenon (Jennings et al 2016). It is essential to maintain the urban green belt as it maintains air quality (Deshmukh et al 2020), ecosystem equilibrium (Anjali et al 2020) and to avoid climate change impact (Velasco and Chen 2019). Besides, they enhance socioeconomic and cultural values (Chaudhary and Tewari 2010).

The study by the Indian School of Mines, Dhanbad stated that the study of carbon sequestration in vegetation, litter and soil is important for ecorestoration of mining sites (https://bcclweb.in/files/2011/02/Eco\_restoration2015.pdf). The short duration trees grown through agroforestry insist the significance of tree species in climate regulation (Singh and Gokhale 2024). Urban trees store carbon equal to that of tropical forest (https://www.fastcompany.com/ 40589994 /urban-trees-can-store-almost-as-much-carbon-as-tropical-rainforests). The amount of carbon stored by tree species is influenced by the edaphic features and land use changes (Moussa et al 2018, Agarwal et al 2021a, Vineeta et al 2023).

In India, except in a few cities, studies on urban green cover are limited. The study conducted at Amity University Campus highlighted the need of urban green space for climate change mitigation (Sharma et al 2021). The study conducted near adjacent areas of Kolkata city pointed out the relationship between biomass and carbon sequestration and the importance of species selection as trees provide health security to urban residents (Agarwal et al 2021b). The present study was designed to assess the tree cover and its carbon sequestration potential in educational institution namely Thiagarajar College situated in the disturbed and polluted urban space of Madurai in Tamil Nadu, India.

#### MATERIAL AND METHODS

**Study area:** The study area, Thiagarajar college campus is in Teppakulam, Madurai, Tamil Nadu, India located at 9.90108 latitude and 78.151 longitudes (Fig. 1). The average rainfall in Madurai is 849 mm with a minimum of 28°C and a maximum of 42°C. The campus is 13.1 acres in size with 6.16 acres of built-up area. The study area is the connecting link between National Highway NH87 and the main part of Madurai city. It is amidst a residential area with a dense population and heavy vehicular movement. The study was conducted from January 2023 to April 2023.

**Enumeration of woody plant species:** The trees and shrubs in the campus were documented. The number of individuals of each species was listed. The scientific name of the trees and shrubs along with the author's citation were verified using Gamble Flora (1921-1935) and Mathew Flora (1991), International Plant Name Index (https://www.ipni. org/) and Plants of the World Online of Kew Botanical Garden (https://powo.science.kew.org/).

**Estimation of morphometric parameters:** The biomass of woody species was calculated using a non-destructive method. An allometric model with tree diameter, height and wood specific gravity was used (Saral et al 2017). Wood specific gravity was used to reduce the error of biomass estimate (Beets et al 2012). The tree height was measured using a GLM40 Distometer. The girth was measured at 1.3 m from the base of the tree using measuring tape. The diameter and radius were calculated from the girth. Wood Density of the tree species was obtained from http://db.worldag roforestry.org/wd.

Above ground biomass of the woody species (AGB): The above ground biomass of a tree includes the whole shoot, branches, leaves, flowers and fruits.

AGB (kg) = Volume of tree  $(m^3)$  X Wood Density kg/ $m^3$ .

 $V = \pi r^2 H$ , H = height of the tree in meters, R = radius of the tree in meters

**Below ground Biomass (BGB):** The below ground biomass (BGB) includes all biomass of live roots excluding fine roots. The BGB was calculated by using the following formula (Hangarge et al 2012)

 $BGB = AGB \times 0.26$ 

**Total biomass (TBM):** The total biomass is the sum of the above and below ground biomass.

Total biomass = AGB + BGB

**Estimation of carbon storage:** Generally, for any plant species, 50% of its biomass is considered as carbon content.



Fig. 1. Map showing the study area Thiagarajar College in Madurai, Tamil Nadu, India

Carbon = Total biomass x 50%

#### Estimation of carbon sequestration

Weight of  $CO_2$  = carbon content x 3.6663

Weight of  $CO_{2}$  is  $C + 2 \times O = 43.99915$ .

Hence, the ratio of  $CO_2$  to C is calculated as: 43.99915/12.001118 = 3.6663.

In order to determine the weight of carbon dioxide sequestered in the tree, the weight of carbon in the tree was multiplied by 3.6663.

**Estimation of CO<sub>2</sub> level in the atmosphere:**  $CO_2$  levels (ppm) within the campus and outside the campus were measured using a  $CO_2$  detector (Gas Analyzer Monitor Air Quality Meter for Home Indoor Outdoor/ B08FCK4GB7) during April 2023.

#### **RESULTS AND DISCUSSION**

The study area, Thiagarajar College Campus had total of 61 woody species (Table 1). Among them, 56 were trees and the rest of the 5 were shrubs. The total number of trees in the college campus was 503. *Azadirachta indica* had maximum number of individuals (109) followed by *Polyalthia longifolia*. Seventeen species were being represented by a single individual. Among the 61 species, 34 species (56%) were native species and 27 (44%) were exotic species. The tallest tree in the campus was *Polyalthia longifolia* (14.6 m). The smallest tree species was *Punica granatum* (2m).

Samanea saman had the largest trunk with 1.43 m diameter and the highest biomass (12923.72 kg/tree) and carbon storage 6461.80 kg/tree (Table 2). It sequestered 18616.13 kg/tree carbon in its aboveground components and 4840.24 kg/tree carbon in belowground components with a total of 23456.56 kg/tree (Table 3). It sequestered about 31% of the carbon in a tree among all the other species. It was followed by *Wrightia tinctoria, Parkia biglandulosa* and *Ficus benghalensis*. All these species had tall trunk, large diameter and wood density. Height, diameter and wood density influence the biomass, carbon storage and sequestration potential (Chave et al 2014, Mensha et al 2016, Saral et al 2017, Prasadan and Jithila 2018, Yumnam and Ronald 2022).

Besides the tree growth and morphometric parameters the relative abundance, tree cover and its diversity also influence the carbon storage in an area. Mouna et al (2019) observed correlation between carbon stock and number of individuals. High tree density could be one of the reasons for high carbon storage (Vineeta et al 2023). In the study site, *Azadirachta indica* had low biomass when compared to other species, due to its high density it sequestered 25.14 % of the total carbon. Channali et al (2022) observed that *Azadiracta indica* had high carbon sequestration capacity in Karnataka. *Azadirachta indica* and *Samanea saman* were the trees preferred to grow in urban environment due to their fast growth rate and wide range of adaptations to variety of soils (Rahman et al 2014). Besides

Table 1. Woody species with their morphometric parameters at Thiagarajar College Campus in Maduraj Tamil Nadu India

Name of the woody species	Family	No. of	Habit/	Height (m)	Diameter (m)	Wood density**
Adina cordifolia (Poxh ) Brandie	Pubiacaaa	1	Troc Nativo	3 37	0.18	(g/cm)
Adiria corditolia (Roxb.) Brandis	Rubiaceae	1	Tree Native	5.57	0.10	0.01
Albizia lehheck (L.) Benth	Mimosaceae	7	Tree Native	0.02 11 18+2 17	0.12	0.78
A procera(Roxh) Benth	Mimosaceae	17	Tree Native	10 39+2 06	0.36+0.1	0.68
Ailanthus excelsa Roxh	Simaroubaceae	1	Tree Native	8 99	0.3010.1	0.00
Artocarnus beterophyllus Lam	Moraceae	1	Tree Native	5.02	0.0	0.4
Azadirachta indica A Juss	Meliaceae	109	Tree Native	8 87+2 88	0.10	0.39
Raubinia purpurea I	Caesalniniaceae	2	Tree Native	5 66+0 90	0.21+0.13	0.72
B tomentosa l	Caesalpiniaceae	2	Tree Exotic	4 94+2 67	0.06+0.04	0.75
B variegata	Caesalpiniaceae	1	Tree Native	5 56	0.07	0.75
B rufescens Lam	Caesalpiniaceae	1	Tree Exotic	3 75	0.05	0.75
Bombax ceiba L.	Bombacaceae	12	Tree Native	9.27±2.0	0.37±0.13	0.35
Caesalpinia coriaria (Jacq.) Willd.	Caesalpiniaceae	14	Tree Exotic	6.53±1.33	0.36±0.14	1.13
C. pulcherima (L.) Sw.*	Caesalpiniaceae	1	Shrub Exotic	4.16	0.05	0.84
Cassia siamea Lam.	Caesalpiniaceae	6	Tree Exotic	9.22±1.72	0.33±0.97	0.74
Casuarina equisetifolia L.	Casuarinaceae	1	Tree Native	10.66	0.31	0.91
Cocos nucifera L.	Arecaceae	1	Tree Exotic	13.20	0.36	0.61
Crateva religiosa G.Forst.	Capparaceae	4	Tree Native	8.58±2.05	0.29±0.14	0.33
Dalbergia latifolia Roxb.	Fabaceae	3	Tree Native	8.57±2.95	0.16±0.01	0.75
<i>Delonix regia</i> (Bojer ex Hook.) Raf.	Fabaceae	13	Tree Exotic	9.39±2.36	0.3±0.01	1.15
Duranta versicolor L.*	Verbenaceae	8	Shrub Exotic	2.12±.0.03	0.09±0.02	0.56
<i>Elaeis guineensis</i> Jacq.	Palmae	1	Tree Native	11.84	0.27	0.33
Ficus benghalensis L.	Moraceae	4	Tree Exotic	8.83±5.30	0.69±0.41	0.49
F. religiosa L.	Moraceae	7	Tree Native	8.12±2.70	0.35±0.19	0.44
Gliricidia sepium (Jacq.) Kunth	Fabaceae	2	Tree Exotic	9.35±1.09	0.29±0.01	0.68
<i>Gmelina arborea</i> Roxb. ex Sm.	Verbenaceae	2	Tree Native	2.9±0.82	0.75±0.03	0.43
Guazuma tomentosa Kunth	Sterculiaceae	4	Tree Exotic	6.98±1.10	0.16±0.06	0.05
Holoptelea integrifolia (Roxb.) Planch.	Ulmaceae	2	Tree Native	10.89±1.19	0.52±0.24	0.51
Hibiscus rosa-sinensis L.*	Malvaceae	1	Shrub Exotic	7.51	0.05	0.47
Hyophorbe verschaffeltii (W.Bull ex J.Dix) H.Wendl.	Arecaceae	20	Tree Exotic	6.57±1.06	0.38±0.08	0.64
Lannea coromandelica (Houtt.) Merr.	Anacardiaceae	3	Tree Native	6.92±0.98	0.16±0.05	0.58
Leucaena leucocephala (Lam.) de Wit	Fabaceae	2	Tree Native	12.03±1.33	0.15±0.02	0.72
Madhuca longifolia (L.) J.F.Macbr.	Sapotaceae	5	Tree Native	9.01±5.95	0.37±0.23	0.79
Mangifera indica L.	Anacardiaceae	1	Tree Native	5.79	0.10	0.55
Millingtonia hortensis L.f	Bignoniaceae	16	Tree Exotic	8.66±3.33	0.22±0.12	0.48
Mimusops elengi L.	Sapotaceae	13	Tree Native	6.38±2.62	0.22±0.16	0.88
Morinda tinctoria Roxb.	Rubiaceae	3	Tree Native	7.00±1.04	0.19±0.04	0.54
Manilkara zapota (L.) P.Royen	Sapotaceae	1	Tree Exotic	6.28	0.06	0.95
Muntingia calabura L.	Muntingiaceae	3	Tree Native	5.50±1.04	0.11±0.04	0.3
Parkia biglandulosa Wight & Arn.	Mimosaceae	5	Tree Exotic	12.01±1.06	0.61±0.14	0.47
Peltophorum pterocarpum (DC.) Backer ex K.Heyne	Caesalpiniaceae	9	Tree Exotic	10.54±1.48	0.39±0.04	0.63
Pithecellobium dulce (Roxb.) Benth.	Mimosaceae	2	Tree Exotic	9.97±2.16	0.41±0.22	0.66
Plumeria rubra L.	Apocynaceae	1	Tree Exotic	5.73	0.1	0.64
Polyalthia longifolia (Sonn.) Thwaites	Annonaceae	90	Tree Native	14.6±2.64	0.21±0.13	0.58
Pongamia pinnata (L.) Pierre	Fabaceae	1/	Tree Native	7.18±2.45	0.2±0.12	0.61
Psidium guajava L.	Myrtaceae	2	Tree Exotic	6.22±2.32	0.12±0.02	0.85
Punica granatum L.	Lythraceae	1	Tree Exotic	2.00	0.006	0.7
Samanea saman(Jacq.) Merr.	Nimosaceae	3	Tree Exotic	12.82±2.16	1.43±0.05	0.52
	Santalaceae	2		4.86±2.26	0.15±0.002	0.93
	Sapindaceae	5		0.14±1.55	0.10±0.11	0.71
Simaroupa giauca DC.	Simaroupaceae	3		9.28±0.9	0.15±0.02	0.46
	Murtaceae	2	Tree Native	9.33±4.77	0.19±0.13	0.55
Syzygium cummi (L.) Skeels	Cooselpiniooooo	1	Tree Native	0.00±1.00	0.19±0.07	0.7
Tahahuja rasaa (Portal ) DC	Dianoniaceae	42	Tree Exolic	0.9913.20	0.30±0.25	0.99
Tabernaemontana divaricata (L.) D.B.r. ov Doom . 9 Sobult :		1		0.30 2 78±0 00	0.04 0.1+0.02	0.55
Tecome stans (L.) luce or Kunth*	Bigoniaceae	+	Shrub Evotio	2.7 0±0.90	0.11.0.02	0.75
Tectona staris (L.) Juss. ex Nullill	Vorbonaceae	0		3.19±U.13	0.00±0.00	0.40
Terminalia ariuna (Royh, ex DC \ Might & Arn	Combretaceae	∠ 1	Tree Native	0.00±1./1 10.17	0.10±0.02	0.01
T catanna I	Combretaceae	1		10.17	0.19	0.0
Wrightia tinctoria (Royh) R Br	Anocynaceae	1	Tree Native	R 11	0.54	0.34
Values – Mean±SD, n= 1 to 10 depends on the species:	*Shrub **Wood D	ensitv Sourc	e: http://db.worl	dagroforestry.or	a/wd	0.77

#### Carbon Sequestration Potential of Woody Species

Table 2. Biomass and carbon storage potential of woody species (kg/tree) in Thiagarajar College Campus in Madurai, Tamil Nadu, India

Name of the woody species	Above-ground	Below-ground	Total biomass	Above-ground	Below-ground	Total carbon
	biomass	biomass	(kg/tree)	carbon storage	storage	storage
	(kg/tree)	(kg/tree)		(kg/tree)	(kg/tree)	(kg/tree)
Adina cordifolia (Roxb.) Brandis	52.28	6.79	59.08	26.14	3.39	29.54
Aegle marmelos (L.) Corrêa	46.99	6.11	53.1	23.50	3.06	26.55
Albizia lebbeck (L.) Benth.	167.80	21.81	189.60	83.88	10.90	94.79
A procera(Roxb) Benth	179 70	23.36	203 10	89.85	11 68	101.50
Ailanthus excelsa Royh	247 70	32.20	270.00	123.80	16.10	130.00
Artocarnus beteronbyllus Lam	45.57	5 924	51 /0	22 70	2.96	25 75
	40.07	05.07	747.20	22.19	2.90	23.73
Azadiracina indica A.Juss.	1001.30	00.97	141.50	62.00	42.90	373.00
Bauminia purpurea L.	120.00	10.03	144.0	03.90	0.32	72.30
B. tomentosa L.	10.47	1.36	11.83	5.24	0.68	5.91
B. variegata L.	11.78	1.53	13.32	5.89	0.77	6.66
<i>B. rufescens</i> Lam.	3.53	0.46	3.99	1.77	0.23	1.99
Bombax ceiba L.	353.80	46.00	399.80	176.90	23.00	199.90
Caesalpinia coriaria (Jacq.) Willd.	763.50	198.50	962.00	381.80	99.26	481.00
C. pulcherima (L.) Sw.	4.39	1.14	5.53	2.19	0.57	2.77
Cassia siamea Lam.	576.80	150.00	726.80	288.40	74.98	363.40
Casuarina equisetifolia L.	741.70	192.80	934.50	370.80	96.42	467.20
Cocos nucifera L.	818.60	212.80	1031.00	409.30	106.40	515.70
Crateva religiosa G.Forst.	191.00	49.66	240.70	95.50	24.83	120.30
Dalbergia latifolia Roxb.	128.00	33.29	161.30	64.01	16.64	80.66
Delonix regia (Bojer ex Hook.) Raf.	776.60	201.90	978.50	388.30	101.00	489.20
Duranta versicolor L.	5.96	1.55	7.52	2.98	0.78	3.76
Elaeis quineensis Jacq.	219.40	57.05	276.50	109.70	28.52	138.20
Ficus benghalensis I	1623.00	421 90	2044 00	811 30	210.90	1022 00
F religiosa I	338.20	87 94	426.20	169 10	43.97	213 10
Gliricidia senium (Jacq.) Kunth	410.00	106 60	516.60	205.00	53 30	258 30
Cmelina arborea Roxh, ex Sm	536.00	139.40	675.40	268.00	69.69	337 70
Guazuma tomentosa Kunth	6.68	1 74	9 /1	200.00	09.09	4 20
	0.00	1.74	0.41	5.34	0.07	4.20
Holoplelea Integniolia (Roxb.) Planch.	1169.00	309.10	1496.00	594.50	154.60	749.10
HIDISCUS rosa-sinensisL.	4.43	1.15	5.59	2.22	0.58	2.79
Hyophorbe verschaffeltii (W.Bull ex J.Dix) H.Wendl.	482.30	125.40	607.70	241.10	62.69	303.80
Lannea coromandelica (Houtt.) Merr.	76.81	19.97	96.79	38.41	9.99	48.39
<i>Leucaena leucocephala</i> (Lam.) de Wit	158.90	41.31	200.20	79.44	20.66	100.10
Madhuca longifolia (L.) J.F.Macbr.	775.80	201.70	977.50	387.90	100.90	488.80
Mangifera indica L.	22.84	5.9	28.78	11.42	2.97	14.39
Millingtonia hortensis L.f	153.00	39.79	192.80	76.52	19.89	96.41
Mimusops elengi L.	219.00	56.95	276.00	109.50	28.47	138.00
Morinda tinctoria Roxb.	112.00	29.12	141.10	56.00	14.56	70.56
Manilkara zapota (L.) P.Royen	19.00	4.94	23.94	9.50	2.47	11.97
Muntingia calabura L.	16.11	4.19	20.29	8.05	2.09	10.15
Parkia biglandulosa Wight & Arn.	1674.00	435.20	2109.00	836.80	217.60	1054.00
Peltophorum pterocarpum (DC.) Backer ex K.Heyne	813.10	211.40	1025.00	406.60	105.70	512.30
Pithecellobium dulce (Roxb.) Benth.	871.80	226.70	1098.00	435.90	113.30	549.20
Plumeria rubra L.	28.06	7.29	35.35	14.03	3.65	17.68
Polvalthia longifolia (Sonn.) Thwaites	302 70	78 69	381.30	151 30	39.34	190 70
Pongamia pinnata (L.) Pierre	134 20	34.89	169 10	67 10	17 45	84 54
Psidium quaiava l	60 79	15.80	76 59	30.39	7 90	38.29
Punica granatum l	0.04	0.01	0.05	0.02	0.06	0.03
	10056 00	0.01	10000 70	0.0Z	1222.40	0.03
Santalina Santali(Jacq.) Men.	76.05	2000.00	12923.72	0120.40	0.01	401.00
	70.25	19.62	90.07	30.12	9.91	46.04
Sapindus emarginatus vani	149.50	38.88	188.40	/4.//	19.44	94.21
Simarouba glauca DC.	78.35	20.37	98.72	39.17	10.18	49.36
Sterculia foetida L.	147.20	38.27	185.50	73.59	19.13	92.73
Syzygium cumini (L.) Skeels	114.10	29.68	143.80	57.07	14.84	71.91
Tamrindus indica L.	889.00	231.10	1120.00	444.50	115.60	560.10
Tabebuia rosea (Bertol.) DC.	1413.00	367.30	1780.00	706.30	183.60	890.00
Tabernaemontana divaricata (L.) R.Br. ex	16.37	4.26	20.62	8.18	2.13	10.31
Tecoma stans (L.) Juss. ex Kunth	8.01	2.08	10.09	4.00	1.04	5.04
Tectona grandis L.f.	135.20	35.14	170.30	67.59	17.57	85.16
<i>Terminalia arjuna</i> (Roxb. ex DC.) Wight & Arn.	233.20	60.64	293.90	116.60	30.32	146.90
T. catappa L.	1547.00	402.30	1949.00	773.60	201.10	974.70
Wrightia tinctoria (Roxb.) R.Br.	1884.00	489.90	2374.00	942.20	245.00	1187.00
Total	33110 81	8360 65	41469 55	16554 45	4179 93	20734 54

#### K. Saraswathi and P. Sneha

Table 3. Carbon sequestration potential of woody species in Thiagarajar College Campus in Madurai, Tamil Nadu, India

Name of the woody species	Above-ground carbon sequestration (kg/tree)	Below-ground carbon sequestration (kg/tree)	Total carbon sequestration (kg/tree)	Total carbon sequestration of all the individuals of the species (kg/ha)
Adina cordifolia (Roxb.) Brandis	94.89	12.34	107.23	3.27
Aegle marmelos (L.) Corrêa	85.29	11.088	96.38	2.94
Albizia lebbeck (L.) Benth.	304.49	39.58	344.08	73.54
A. procera(Roxb.) Benth.	326.14	42.39	368.55	191.31
Ailanthus excelsa Roxb.	449.56	58.44	508.00	93.07
Artocarpus heterophyllus Lam.	82.71	10.752	93.46	2.85
Azadirachta indica A.Juss.	1200.25	156.03	1356.29	4514.10
Bauhinia purpurea L.	232.25	30.19	262.44	16.02
B. tomentosa L.	19.00	2.47	21.47	1.31
B. variegata L.	21.39	2.78	24.16	0.74
<i>B. rufescens</i> Lam.	6.41	0.833	7.24	0.22
Bombax ceiba L.	642.20	83.48	725.68	265.90
Caesalpinia coriaria (Jacq.) Willd.	1385.76	360.29	1746.06	746.41
C. pulcherima (L.) Sw.	7.96	2.07	10.03	0.31
<i>Cassia siamea</i> Lam.	1046.88	272.19	1319.07	241.66
Casuarina equisetifolia L.	1346.12	349.99	1696.10	51.79
Cocos nucifera L.	1485.76	386.29	1872.05	57.16
Crateva religiosa G.Forst.	346.68	90.13	436.81	53.35
Dalbergia latifolia Roxb.	232.36	60.41	292.78	26.82
<i>Delonix regia</i> (Bojer ex Hook.) Raf.	1409.47	366.46	1775.97	704.95
Duranta versicolor L.	10.83	2.81	13.64	3.33
Elaeis guineensis Jacq.	398.22	103.54	501.77	15.32
Ficus benghalensis L.	2944.91	765.68	3710.59	453.20
F. religiosa L.	613.92	159.62	773.53	165.34
<i>Gliricidia sepium</i> (Jacq.) Kunth	744.19	193.49	937.68	57.26
<i>Gmelina arborea</i> Roxb. ex Sm.	972.92	252.96	1225.88	74.86
Guazuma tomentosa Kunth	12.12	3.15	15.27	1.87
Holoptelea Integritolia (Roxb.) Planch.	2158.05	561.09	2/19.15	166.05
Hibiscus rosa-sinensisL.	8.05	2.09	10.14	0.31
Hyophorbe Verschaffeitil (W.Bull ex J.Dix) H.Wendi.	875.31	227.58	1102.89	673.52
Lannea coromandelica (Houtt.) Merr.	139.42	30.25	1/5.0/	16.09
Madhuca longifolia (L.) LE Machr	200.30	74.90	1774 21	22.19
Magnuca longitolia (L.) J.F.MacDi.	1400.11	10.78	52.24	1 59
Millingtonia hortensis   f	277 76	72.22	349 97	170.98
Mimusons elengi l	397.55	103.36	500.91	198.83
Morinda tinctoria Roxb	203 28	52 85	256 14	23.46
Manilkara zapota (L.) P.Roven	34 48	8.97	43 45	1.33
Muntingia calabura L.	29.23	7.60	36.83	3.37
Parkia biglandulosa Wight & Arn.	3037.76	789.82	3827.58	584.36
Peltophorum pterocarpum (DC.) Backer ex K.Hevne	1475.85	383.72	1859.57	511.03
Pithecellobium dulce (Roxb.) Benth.	1582.37	411.41	1993.78	121.76
Plumeria rubra L.	50.93	13.24	64.17	1.96
Polyalthia longifolia (Sonn.) Thwaites	549.31	142.82	692.13	1902.00
Pongamia pinnata (L.) Pierre	243.57	63.33	306.89	159.30
Psidium guajava L.	110.33	28.68	139.01	8.49
Punica granatum L.	0.07	0.01	0.09	0.002
Samanea saman (Jacq.) Merr.	18616.32	4840.24	23456.56	2148.70
Santalum album L.	138.39	35.98	174.37	10.64
Sapindus emarginatus Vahl	271.42	70.57	341.99	52.21
Simarouba glauca DC.	142.19	36.97	179.16	16.41
Sterculia foetida L.	267.14	69.45	336.59	20.55
Syzygium cumini (L.) Skeels	207.17	53.86	261.03	55.794
Tamrindus indica L.	1613.57	419.53	2033.10	2607.3
Tabebuia rosea (Bertol.) DC.	2563.99	666.64	3230.63	98.64
Tabernaemontana divaricata (L.) R.Br. ex	29.71	7.72	37.43	4.57
<i>Tecoma stans</i> (L.) Juss. ex Kunth	14.53	3.78	18.31	3.35
Tectona grandis L.f.	245.33	63.79	309.12	18.88
Terminalia arjuna (Roxb. ex DC.) Wight & Arn.	423.29	110.06	533.35	16.29
T. catappa L.	2808.20	730.13	3538.33	108.04
Wrightia tinctoria (Roxb.) R.Br.	3420.12	889.23	4309.35	131.58
Total	60095.38	15174.4	75269.78	17949.45

these two tree species the other species such as *Delonix regia*, *Tamarindus indica*, *Ficus religiosa*, *Peltophorum pterocarpum*, *Albizia lebbeck*, *Terminalia catappa*, *Ficus benghalensis* and *Terminalia arjuna* growing in Thiagarajar College campus were observed as preferable species for road side plantation (Ragula and Chandra 2020). The outside environment of the college campus had an average of 537.87 ppm CO<sub>2</sub>and within the college campus 496.65 ppm CO<sub>2</sub> in the atmosphere. There was no large difference in the CO<sub>2</sub> level in and around the campus. From the study it was inferred that the CO<sub>2</sub> level was maintained within the permissible limit by the woody tree species, though the campus had high anthropogenic disturbances (https://www.health.state.mn.us/communities /environment/air/toxins/co2.html, Sharma et al 2021).

#### CONCLUSION

Samanea saman, Wrightia tinctoria, Parkia biglandulosa, Ficus benghalensis, and Terminalia cattapa have high sequestration potential. Azadirachta indica is able to thrive well in all kinds of soil. The study suggests that these species are suitable for urban spaces with high anthropogenic disturbances and heavy vehicular movement.

#### REFERENCES

- Agarwal S, Chakrabarty SP, Zaman S, Pramanick P, Biswas P, Mukhopadhyay N, Dutta J and Mitra A 2021a. Assessment of Carbon Storage Potential of Dominant Tree Species in an Educational Campus, Kolkata, West Bengal. *Ela Journal of Forestry and Wildlife* **10**(2): 869-878.
- Agarwal S, Mitra A, Pramanick P and Mitra A 2021b. Stored Carbon in Urban trees Ground Zero Observation from the Konnagar Area of West Bengal, India, pp 1-22. In: Filho WL, Luetz JM and Ayal D (eds). Handbook of Climate Change Management. Springer Nature Switzerland AG.
- Anjali K, Khuman, YSC and Sokhi J 2020. A Review of the interrelations of terrestrial carbon sequestration and urban forests. AIMS Environmental Science 7(6): 464-485.
- Beets PN, Kimberley MO, Oliver GR, Pearce SH, Graham D and Brandon A 2012. Allometric equations for estimating carbon stocks in Natural Forest in New Zealand. *Forests* 3(3): 818-839.
- Channalli SPS, Salunke RS, Jadhav V, Patil PL and Dolli S 2022. An assessment of seasonal variation of carbon emission and carbon sequestration in rural areas of Belagavi District, Karnataka. *International Journal of Plant and Soil Science* **34** (24): 827-844.
- Chaudhary P and Tewari VP 2010. Managing urban parks and gardens in developing countries: A case from an Indian city. *International Journal of Leisure and Tourism Marketing* **1**(3): 248-256.
- Chave J, Réjou-Méchain M, Búrquez A, Chidumayo E, Colgan MS, Delitti WB, Duque A, Eid T, Fearnside PM, Goodman RC, Henry M, Martínez-Yrízar A, Mugasha WA, Muller-Landau HC, Mencuccini M, Nelson BW, Ngomanda A, Nogueira EM, Ortiz-Malavassi E, Pélissier R, Ploton P, Ryan CM, Saldarriaga JG and Vieilledent G 2014. *Global Change Biology* **20**(10): 3177-3290.
- Deshmukh A, Shaikh N, Soni N and Kumavat U 2020. Carbon sequestration potential of tree species of VPM campus, Thane. *J-BNB: A Multidisciplinary Journal* 9:57-60.

Dubal K, Ghorpade P, Dongare M and Patil S 2013. Carbon

Received 05 July, 2024; Accepted 21 September, 2024

sequestration in the standing trees at campus of Shivaji University, Kolhapur. *Nature Environment and Pollution Technology* **12**(4): 725-726.

- Hangarge LM, Kulkarni DK, Gaikwad VB, Mahajan DM and Chaudhari N 2012. Carbon Sequestration potential of tree species in Somjaichi Rai (Sacred grove) at Nandghur village, in Bhor region of Pune District, Maharashtra State, India. *Annuals* of *Biological Research* **3**(7): 3426-3429.
- Jennings V, Larson L and Yun J 2016. Advancing sustainability through urban green space: cultural ecosystem services, equity, and social determinants of health. *International Journal of Environment, Research* and *Public Health* **13**: 196-210.
- Lahoti S, Lahoti A, Joshi RK and Saito O 2020. Vegetation structure, species composition, and carbon sink potential of urban green spaces in Nagpur City, India. *Land* **9**(4): 107.
- Mensha S, Veldtman R, Ben du Toit, Kakaï RG and Seifert T 2016. Aboveground biomass and carbon in a South African Mistbelt Forest and the relationships with tree species diversity and forest structures. *Forests* **7**(79): 1-17.
- Mouna S, Jabeen FTZ, Dakshayini J, Haleshi J and Seetharam YN 2019. Carbon sequestration and tree diversity of Thimmalapura Reserve Forest, Tumakuru, Karnataka. *Indian Journal of Ecology* 46(1): 116-125.
- Moussa S, Kyereh B, Tougiani AA and Saadou M 2018. Carbon stocks of neem tree (*Azadirachta indica* A. Juss.) in different urban land use and land cover types in Niamey city, Niger, West Africa. South Asian Journal of Biological Research 1(2): 153-165.
- Prasadan PK and Jithila PJ 2018. Carbon storage and sequestration by trees: A study in Western Ghats Wayanad Region. *Indian Journal of Ecology* **45**(3): 479-482.
- Ragula A and Chandra KK 2020. Tree species suitable for roadside afforestation and carbon sequestration in Bilaspur, India. *Carbon Management* **11**: 369-380.
- Rahman AK Danial MMS Faiz MK et al 2014. *Inventory of tree biomass and volume allometric equations for South East Asia.* FRIM, Kepong, FAO of the United Nation, Rome Italy, p41.
- Saral AM, Selcia SS and Devi K 2017. Carbon storage and sequestration by trees in VIT University Campus. *IOP Conference Series: Materials Science and Engineering* **263**: 1-5.
- Sen S 2020. Green'-ing Kolkata: Creating a sustainable city: An overview. International Journal of Research and Analytical Reviews 7(2): 743-752.
- Sharma R, Pradhan L, Kumara M and Bhattacharya P 2021. Assessment of carbon sequestration potential of tree species in Amity university campus, Noida. *Environmental science* proceedings 3(52): 1-9.
- Singh N and Gokhale Y 2024. Potential and performance of poplars under short rotation agroforestry models: A case study of Yamunanagar district of Haryana. *Indian Journal of Ecology* 51(2):281-285.
- Velasco E and Chen KW 2019. Carbon storage estimation of tropical urban trees by an improved allometric model for aboveground biomass based on terrestrial laser scanning. *Urban Forestry and Urban Greening* **44**: 126387.
- Veeramani S, Anoop V, Babu MR, Subashrao PS and Suhyb PJ 2023. Carbon stock assessment and prediction in the Periyar Tiger Reserve, India, using Markov Chain and Invest Model. *Indian Journal of Ecology* **50**(6): 1965-1971.
- Vineeta, Sarkar BC, Tamang M, Shukla G, Shahina NN, Nath AJ and Chakravarty S 2023. Variation in tree density, biomass and carbon stock across an altitudinal gradient under large cardamom agroforestry system of Darjeeling Himalaya. *Research Square Platform LLC* DOI: https://doi.org/10.21203 /rs.3.rs-3330722/v1
- Yumnam JY and Ronald K 2022. Disparity in phytosociology, biomass and carbon stock of trees in primary and secondary temperate Broadleaf Forest of Indian Himalayas. *Indian Journal* of Ecology 49(5): 1613-1620.



Indian Journal of Ecology (2024) 51(5): 954-960 DOI: https://doi.org/10.55362/IJE/2024/4335 Manuscript Number: 4335 NAAS Rating: 5.38

# Ethnobotanical Exploration of Polypetalous Riparian Flora along Beas River in Himachal Pradesh Utilized by Traditional Practitioners in the Management of Hypertension

Neha Thakur and Nitesh Kumar<sup>\*</sup>

Department of Biosciences, Himachal Pradesh University, Shimla-171 005, India \*E mail: niteshchauhan7@gmail.com

**Abstract:** Polypetalous plants of Riparian vegetation along the Beas River of Himachal Pradesh were utilized to treat various diseases. Extensive fieldwork was carried out to document the traditional use of ethnomedicinal plants, and a survey was conducted from March 2024 to June 2024 to assess the significance of these plants in healthcare practices. A total of 143 informants, including 65 men and 78 women, were randomly chosen for interviews via semi-structured questionnaires. The study computed several quantitative indices: frequency citation, Jaccard index, family importance value, and relative frequency of citation. Thirty polypetalous plant species distributed among 15 different families were identified for their use in treating hypertension in Himachal Pradesh. The predominant life form documented in the study was herbs (17 species). The most typical preparation mode was powder (11 species), and plant parts were root (8 species). Among the families, Fabaceae showed the highest family importance value of 20.97, and the relative frequency of citation varied between 0.10 and 0.41 across the reported plant species. This research bridges the conventional knowledge of polypetalous plants offering a promising avenue for developing novel therapeutics for hypertension.

#### Keywords: Ethnobotany, Hypertension, Traditional Medicine, Polypetalous, Jaccard index

Cardiovascular conditions are significantly high in human populations, making their avoidance and management a top priority in numerous countries (Aumeeruddy and Mahomoodally 2020). In current years, the practice of therapeutic herbs for treating diseases has seen a substantial increase. High blood pressure can harm the blood vessels that supply blood to the heart, eyes, brain, and kidneys (Malik et al 2018). It plays a significant role in the growth of cardiovascular incidents, heart disease, and stroke. The mortality rate from hypertension is double as high as that of the common populace. It raises the risk of rapid death in patients with cardiovascular disease. Hypertension, described by high BP levels, is a significant risk issue for circulatory diseases, stroke, and renal complications, contributing significantly to global morbidity and mortality (Baharvand-Ahmadi and Asadi-Samani 2017). The high BP affects around 1.27 billion adults aged 29-80 worldwide (Meresa et al 2017). This condition is most prevalent in lowand middle-income countries, where two-thirds of the affected populace resides. Approximately 10 million people die each year from hypertension-related causes, making it more lethal than all infectious diseases combined (Baharvand-Ahmadi et al 2015). Efforts to manage hypertension focus on lifestyle changes such as adopting a healthier diet, decreasing salt intake, boosting physical activity, giving up smoking, and limiting alcohol consumption. For many, medication is also essential to attain optimal blood pressure control (Diallo et al 2019). The alternative method to cure and manage hypertension is through the use of herbal remedies. These polypetalous plants are effective in treating high blood pressure and addressing various other health conditions (Ajayi et al 2019). Medicinal plants have been used for centuries in folk medicine to cure numerous ailments (Ozturk et al 2018). This ancient therapeutic medicine offers a practical and thorough approach to using herbal plants, and adhering to its principles can be advantageous in tackling public health challenges. Ethno-botanical surveys studies aimed at documenting therapeutic information and the usage of herbal plants. In modern times, pharmacology has made remarkable advances and herbal drugs are utilized alongside conventional pharmaceuticals to address a wide array of health conditions (Panmei et al 2019). Reports indicate that over 30% of modern medicines have their origins traced back to medicinal plants. Numerous herbal plants have been thoroughly researched for their medicinal properties thus far. Despite advances in conventional pharmacotherapy, many individuals worldwide rely on traditional medicinal practices, including the usage of plant-based medications, for the management of high BP (Hassaine et al 2019, Nunes et al 2015). The ethnobotanical knowledge embedded within traditional healing systems offers a rich repository of potential therapeutic agents that warrant systematic investigation and validation. Due to the widespread occurrence of hypertension globally, particularly in Himachal Pradesh, and

the pressing demand for the exploration of potent herbal remedies, this research aimed to compile information on polypetalous plants endorsed by local people in the riparian region of Beas River of Himachal Pradesh for managing hypertension.

#### MATERIAL AND METHODS

**Research area:** The research was carried out in the riparian area of the Beas River in Himachal Pradesh, India. The Beas basin originates in the Shivalik Hills of Himachal Pradesh and extends over 460 km. This area covers 12,130 square kilometers, situated between latitudes 31°41' N to 32°46' N and longitudes 75°84' E to 77°88' E.

#### Methodology

**Ethnobotanical survey**: Direct interviews with traditional healers were carried out from March 2024 to June 2024 using a semi-structured questionnaire. The structured questionnaire was administered to traditional healers across diverse geographical regions known for their rich biodiversity. Information regarding the plants' identity, preparation methods, dosage, and perceived efficacy was documented.

**Collection of plants and identification**: Plant specimens cited by traditional healers were collected, identified, and authenticated by botanists and taxonomists from the Himalayan Forest Research Institute (HFRI), Shimla. Voucher specimens were deposited in the herbaria of Himachal Pradesh University, Shimla.

#### **Quantitative Analysis**

**Relative frequency of citation (RFC):** It is determined to assess the extent of indigenous information about the usage of therapeutic herbs in the studied regions (Nadaf et al 2019):

Where "Fc" represents the number of participants who mentioned using the species, whereas "N" represents the overall number of participants.

**Family importance value (FIV):** These values indicate the respondents' information on the plant families used. The FIV values of the therapeutic herb were measured (Nadaf et al 2019):

$$FIV = \frac{No. \text{ of families cited by authors}}{Total \text{ no. of authors}} \times 100$$

A high value indicates that there is extensive information and several respondents are well-known, whereas a low value suggests limited awareness about the usage of that plant family (Nadaf et al 2019).

Jaccard similarity index (JI): It is computed by assessing previous research publications across national, international, and worldwide scopes. This calculation involves determining the proportions of the mentioned plant species and their therapeutic usage (Faruque et al 2018, Kayani et al 2015):

$$JI = \frac{c}{a+b+c}$$

In this formula, "a" symbolizes the count of plants in Area 1, "b" symbolizes the count of plants in Area 2, and "c" symbolizes the count of floras common to both Areas 1 and 2. **Statistical analysis:** The Excel 2016 spreadsheet was utilized for basic calculations and to ascertain plant frequencies.

#### **RESULTS AND DISCUSSION**

Diversity of polypetalous plants: The riparian vegetation of the Beas River used 30 taxa from 15 families and 27 genera (Table 2, Fig. 3). The most participants were between the ages of 39 and 79 (Table 1). The majority of informants who are interested in traditional therapeutic information are from the older age group (Ahmed et al 2015; Kayani et al 2015; Malik et al 2018). The Fabaceae family, with 8 plants, followed by Ranunculaceae, was the most prevalent among medicinal plants (Fig. 4). Out of approximately 350,000 identified species of flowering plants, nearly 9% are part of the Fabaceae family, which can observed in almost every type of environment across various regions (Gbekley et al 2018). However, in the research site, more Ranunculaceae and Apiaceae species were employed, as previously reported. Informants identify a variety of herbal species within the same family, each with unique medical capabilities. Another possible explanation for the high citation of Fabaceae and Ranunculaceae is their higher occurrence in high-altitude regions, as observed in previous research.

The herbs were the preferred choice among therapeutic herb species due to their high natural richness in these regions and easy accessibility to local communities. Herbs are readily available and possess potent healing capabilities, producing secondary metabolites with therapeutic properties effective

Та	bl	e '	1.	D	emoar	aph	nic c	haract	terist	tics o	ft	he respondents
	~	•	••	-	onnogn			i lai ao				

Characteristics	Groups	Informants	Percentages
Gender	Male	65	45.45
	Female	78	54.54
Age	30-50	31	21.67
	51-65	46	32.16
	66-79	57	39.86
Ethnic group	Gaddi	72	50.34
	Gujjar	69	48.25
Education	Illiterate	41	28.67
	Matric	59	41.25
	Secondary school	52	36.36

Table 2. Ethnomec	licinal uses of po	olypetalous plants								
Botanical name	Local name	Family name	Habit	Parts used	Mode of preparation	Ethnobotanical uses	- C	RFC	Jaccard ndex (JI)	FIV
Acacia nilotica (L.)	Kikar	Fabaceae/Leguminosae	Tree	Root	Infusion	One glass of root infusion is drunk after	26	0.18	0.53	20.97
Argemone mexicana L.	Kanduri, Bharbhand	Papaveraceae	Herb	Seed and Root	Powder, Infusion	a mean to reduce high blood pressure. 1-2 grams of the dried seed powder is given with mildly heated water to reduce hypertension.	8	0.12	0.60	6.99
Bauhinia variegata L.	Karali	Fabaceae	Tree	Flower and Bud	Powder	night to reduce high blood pressure. One teaspoon of powder of dried flowers and buds is taken with water to	33	0.23	0.57	20.97
Berberis lycium Royle	Kashmal	Berberidaceae	Shrub	Root	Infusion	reduce hypertension. 4-5 ml of root infusion is drunk early in the morning to decrease high blood	39	0.27	0.58	6.99
Berberis aristata DC.	Kashmal	Berberidaceae	Shrub	Leaf and fruit	Cooked	pressure. Leaves and fruits are cooked to make an infusion and 2-3 ml is drunk two times a day to reduce high blood pressure.	52	0.15	0.55	6.99
Capsella bursa	Jangli sarson	Brassicaceae	Herb	Shoot	Extract	1-2 ml extract of the shoot is drunk	15	0.10	0.69	6.99
pastoris (L.) iviedik. Cassia fistula L.	Amaltas/ Aliah	Fabaceae/Leguminosae	Tree	Bark	Powder	orice a uay to decrease righer terision. 1-2 grams of the dried bark powder is	58	0.19	0.76	20.97
Cassia occidentalis	Bara elwan	Caesalpiniacae/Leguminosae	Shrub	Seed	Infusion	given with water to reactly percension. 2ml of the seed infusion seed is drunk offer a meal to reduce hypertension	31	0.21	0.77	6.99
Cassia tora L.	Reli	Caesalpiniaceae/Leguminosae	Shrub	Seed	Powder	1-2 grams of dried seed powder is drunk with warm water to reduce high BD.	56	0.20	0.70	6.99
Centella asiatica (L.) Urban	Brahmi	Apiaceae/ Umbelliferae	Herb	Whole plant	Decoction	The decoction of the entire plant is drunk early in the morning to reduce	54	0.16	0.63	6.99
Clematis grata Wall.	Dhand	Ranunculaceae	Climber	Flower	Powder	ngristood prostore. 1 gram powder of the dried flower is given with mildly heated water to treat by mortonsion	17	0.11	0.57	10.48
Fragaria nubicola	Kida-bhumla/	Rosaceae	Herb	Fruit	Fresh	Fresh fruits are consumed to lower	32	0.22	0.50	6.99
Euruu. <i>Fumaria indica</i> (Haussk) Pugsley.	Lai aanie Pitpapra	Papaveraceae	Herb	Whole plant	Decoction	The decoction of the entire plant is drunk after a meal to cure high blood	5	0.14	0.64	6.99
Geranium walichianum Oliv	Laljari	Geraniaceae	Herb	Root	Decoction	pressure. 3-4 ml of root decoction is utilized for the treatment of hynectension	27	0.18	0.52	3.49
wamonanan oliv. Impatiens balsamina L.	Tiur	Balsaminaceae	Herb	Leaf and Root	Decoction and Powder	The leaf decortion is given after a meal to cure High Blood pressure. The dried root powder is given with mildly heated water to treat hypertension.	50	0.20	0.60	3.49

956

#### Neha Thakur and Nitesh Kumar

Cont...

Table 2. Ethnomed	licinal uses of pol	ypetalous plants								
Botanical name	Local name	Family name	Habit	Parts used	Mode of preparation	Ethnobotanical uses	с Ч	RFC	Jaccard Index (JI)	FIV
Indigofera hamiltonii Graham	Kathi	Fabaceae	Shrub	Root	Decoction	5 ml of the decoction of the root is drunk two times a week to reduce hypertension.	39	0.27	0.58	20.97
Mangifera indica L.	Aam	Anacardiaceae	Tree	Stem, Bark	Decoction	25–30 mL of decoctions of stem and bark are drunk twice daily to cure high blood pressure.	51	0.35	0.67	3.49
Melilotus alba Medik.	Kutaik	Fabaceae	Herb	Leaf	Infusion	The infusion of the leaves is given daily to reduce high blood pressure.	30	0.20	0.57	20.97
Medicago polymorpha L.	Khokhani	Fabaceae	Herb	Whole plan	it Powder	The powder of the entire plant is given with warm water to lower hypertension.	42	0.29	0.54	20.97
<i>Murraya koenigii</i> (L.) Spreng.	Kadipata	Rutaceae	Shrub	Leaf	Powder	Dried leaf powder is taken with lukewarm water to cure hypertension	20	0.13	0.50	3.49
Nasturtium officinale W.T.Aiton	Chuch, Nadd	Brassicaceae	Herb	Shoot	Decoction	Shoots are cooked to make a decoction and taken early in the morning to decrease hypertension.	44	0.30	0.61	6.99
Nigella sativa L.	Kala jeera, kalonji	Ranunculaceae	Herb	Seed	Extract	3-4 ml extract of seed is utilized to reduce high blood pressure.	22	0.15	0.48	10.48
Oxalis comiculata L.	Malori	Oxalidaceae	Herb	Aerial part	Extract	The extract of the aerial part is utilized to reduce hypertension.	54	0.37	0.72	3.49
<i>Pimpinella</i> <i>acuminata</i> (Edgew.) C.B. Clarke	Tarpakhi	Apiaceae	Herb	Root	Powder	2-3 g powder of the dried root is drunk with mildly heated water to reduce hypertension.	17	0.11	0.41	6.99
Ranunculus muricatus L.	Jaldhar/ bimbi	Ranunculaceae	Herb	Stem an Leaf	d Infusion	The infusion of stem and leaves is drunk daily to decrease high blood pressure.	48	0.33	0.48	10.48
<i>Rosa brunonii</i> Lindl.	Kuje	Rosaceae	Shrub	Root	Decoction	5-6 ml decoction of the root is drunk early in the morning to reduce hypertension.	45	0.31	0.54	6.99
Sida cordifolia L.	Dragain	Malvaceae	Shrub	Seed	Powder	The powder of the seed is taken with lukewarm water to decrease hypertension.	32	0.22	0.55	3.49
Thalictrum foliolosum DC.	Barmot	Ranunculaceae	Herb	Leaf	Fresh	Fresh juice is used to treat pimples and boils.	56	0.39	0.74	10.48
<i>Trigonella emodi</i> Benth.	Kuchona	Fabaceae	Herb	Leaf	Fresh	Fresh leaves are eaten to decrease hypertension.	23	0.16	0.36	20.97
Viola canescens Wall.	Banfshah	Violaceae	Herb	Flower	Powder	One teaspoonful powder of dried flower is given with mildly heated water to decrease high blood pressure.	59	0.41	0.68	3.49

#### Ethnobotanical Exploration of Polypetalous Riparian Flora



Fig. 2. FIV of polypetalous plants



Fig. 3. Some polypetalous plants 1. Argemone mexicana 2. Berberis lycium 3. Cassia fistula 4. Cassia occidentalis 5. Fragaria nubicola 6. Nasturtium officinale 7. Oxalis corniculata 8. Rosa brunonii 9. Viola canescens



Fig. 4. Families of polypetalous plants



Fig. 5. Habit of polypetalous plants







Fig. 7. Plant parts of polypetalous plants

against numerous ailments (Yaseen et al 2015). This research focused mainly on the use of roots as the main parts of the plant for medicinal purposes. When comparing our results with previous studies, significant variations were observed in terms of the Jaccard Index (JI). The JI ranged from 0.41 to 0.77. In RFC, *Viola canescens, Thalictrum foliolosum,* and *Oxalis corniculata* were identified as the most significant plants. This study highlights numerous medicinal plant species that exhibit pharmacological activity but remain largely unexplored, emphasizing the need for further research.

**Plant parts used:** Different plant parts used in preparing herbal remedies include roots, stems, leaves, flowers, seeds, fruits, bark, aerial parts, and whole plants, especially when dealing with small herbal plants. Root was the most commonly used part (8), followed by leaf (6 each), flower, and whole plant (3 each), shoot, fruit, bark, and stem (2 species each), and bud, and aerial parts (only 1 species) (Fig. 7). Herbs constituted the main source of native medicines (17), followed by shrubs (8), trees (4), and climbers (1) (Fig. 5).

**Mode of preparation:** Herbal preparations were created using various methods such as infusion, powder, decoction, extract, and cooking. The most common usage method was powder (11), followed by decoction, infusion, extract, fresh, and cooked (Fig. 6). Local communities used varied quantities of herbal concoctions to cure a variety of illnesses.

#### **Quantitative Analysis**

**Relative frequency of citation (RFC):** *Viola canescens* had the maximum RFC value (0.41), followed by *Thalictrum foliolosum*, *Oxalis corniculata*, and *Clematis grata* (0.11) had the lowest RFC (Fig. 1).

**Family importance value (FIV):** Fabaceae was the most prevalent medicinal plant family in terms of both species and FIV index, with 8 species and an FVI of 20.97%. This was followed by Ranunculaceae (10.48%), and Apiaceae, Papaveraceae, Caesalpiniaceae, Berberidaceae, Brassicaceae, and Rosaceae (6.99%). The remaining families have fewer species such as Anacardiaceae, Geraniaceae, Balsaminaceae, Rutaceae, Malvaceae, Oxalidaceae, and Violaceae each containing one species with an FIV of 3.49% (Fig. 2).

#### CONCLUSION

The ethnobotanical survey presented herein provides a valuable understanding of the use of polypetalous herbs by tribal people to treat hypertension. The total of 143 informants, including 65 men and 78 women, were randomly chosen for interviews via semi-structured questionnaires. A total of 30 polypetalous plant species distributed among 15 different families were identified for their use in treating hypertension in Himachal Pradesh. The predominant was herbs. The most typical preparation mode was powder and plant parts were root. Among the families, Fabaceae showed the highest family importance value. RFC. These findings offer a foundation for further research aimed at connecting the healing potential of therapeutic herbs in the growth of harmless and effective treatment for hypertension.

#### REFERENCES

- Ahmed N, Mahmood A, Ashraf A, Bano A, Tahir SS and Mahmood A 2015. Ethnopharmacological relevance of indigenous medicinal plants from district Bahawalnagar, Punjab, Pakistan. *Journal of Ethnopharmacology* **175**: 109-123.
- Ajayi TO, Moody JO and Anthony CS 2019. Ethnobotanical survey of plants used in the management of hypertension in Ibadan North Local Government Area of Oyo State, Nigeria. *Nigerian Journal* of *Pharmaceutical Research* **15**(1): 61-73.
- Aumeeruddy MZ and Mahomoodally MF 2020. Traditional herbal therapies for hypertension: A systematic review of global ethnobotanical field studies. *South African Journal of Botany* **135**: 451-464.
- Baharvand-Ahmadi B and Asadi-Samani M 2017. A mini-review on the most important effective medicinal plants to treat hypertension in ethnobotanical evidence of Iran. *Journal of Nephropharmacology* **6**(1): 3-12.
- Baharvand-Ahmadi B, Bahmani M, Eftekhari Z, Jelodari M and Mirhoseini M 2015. Overview of medicinal plants used for cardiovascular system disorders and diseases in ethnobotany of different areas in Iran. *Journal of HerbMed Pharmacology* **5**(1): 39-44.
- Diallo MST, Traore MS, Balde MA, Camara AK, Baldé ES, Traore S and Balde AM 2019. Prevalence, management and

ethnobotanical investigation of hypertension in two Guinean urban districts. *Journal of Ethnopharmacology* **231**: 73-79.

- Faruque MO, Uddin SB, Barlow JW, Hu S, Dong S, Cai Q and Hu X 2018. Quantitative ethnobotany of medicinal plants used by indigenous communities in the Bandarban District of Bangladesh. *Frontiers in pharmacology* **9**: 40.
- Gbekley HE, Karou SD, Katawa G, Tchacondo T, Batawila K, Ameyapoh Y and Simpore J 2018. Ethnobotanical survey of medicinal plants used in the management of hypertension in the maritime region of Togo. African Journal of Traditional, Complementary and Alternative Medicines 15(1): 85-97.
- Hassaïne S, Saïdi A and Belhadj O A 2019. Ethnobotanical study of medicinal plants used in the treatment of high blood pressure in the region of Tlemcen (Northwestern Algeria). *Journal of Pharmacy & Pharmacognosy Research* **7**(1): 1-11.
- Kayani S, Ahmad M, Sultana S, Shinwari ZK, Zafar M, Yaseen G and Bibi T 2015. Ethnobotany of medicinal plants among the communities of Alpine and Sub-alpine regions of Pakistan. *Journal of Ethnopharmacology* **164**: 186-202.
- Malik K, Ahmad M, Bussmann RW, Tariq A, Ullah R, Alqahtani AS and Shah SN 2018. Ethnobotany of anti-hypertensive plants used in northern Pakistan. *Frontiers in Pharmacology* **9**: 789.

Received 23 July, 2024; Accepted 06 October, 2024

- Meresa A, Fekadu N, Degu S, Tadele A and Geleta B 2017. An ethnobotanical review on medicinal plants used for the management of hypertension. *Journal of Clinical and Experimental Pharmacology* **7**(2): 1-16.
- Nadaf M, Joharchi M and Amiri MS 2019. Ethnomedicinal uses of plants for treating nervous disorders at the herbal markets of Bojnord, North Khorasan Province, Iran. Avicenna Journal of Phytomedicine 9(2): 153.
- Nunes MGS, Bernardino A and Martins RD 2015. Use of medicinal plants by people with hypertension. *Northeast Network Nursing Journal* **16**(6): 775-781.
- Ozturk M, Altay V, Latiff A, Shareef S, Shaheen F and Iqbal Choudhry M 2018. Potential medicinal plants used in hypertension in Turkey, Pakistan, and Malaysia. *Plant and Human Health, Volume 1: Ethnobotany and Physiology* 595-618.
- Panmei R, Gajurel PR and Singh B 2019. Ethnobotany of medicinal plants used by the Zeliangrong ethnic group of Manipur, northeast India. *Journal of Ethnopharmacology* **235:** 164-182.
- Yaseen G, Ahmad M, Sultana S, Alharrasi AS, Hussain J and Zafar M 2015. Ethnobotany of medicinal plants in the Thar Desert (Sindh) of Pakistan. *Journal of Ethnopharmacology* **163**: 43-59.



# Carbon Stock Quantities of *Shorea robusta* Gaertn. along Altitudinal Gradient in Shivalik Hills of Western Himalaya

Himshikha Gusain

Department of forestry and Natural Resources HNB Garhwal University Srinagar Garhwal-246 174, India E-mail: Himshikha.hnb@gmail.com

**Abstract:** The study explains altitude wise carbon stock quantities of *Shorea robusta* Gaertn. in the Bhabhar transition zone of Western Himalaya, an important zone from biodiversity conservation and climate change point of view in Uttarakhand. The area was divided into three zones viz. zone I (400-700 m), II (701-1000 m) and III (1001-1300 m). 10 sample plots (total 30 plots) were laid randomly for tree investigation and soil sampling. Based on the generated data through non-harvesting method, a gradual reduction was observed in studied parametric values from 96 trees. Tree volume and carbon stock (105.12-457.38 t ha-<sup>1</sup>) were recorded highest at lower elevation. Contrary to this, soil carbon content increased from 33.33 t ha-<sup>1</sup> to 40.96 t ha-<sup>1</sup> as the elevation went-up from 700 to 1300 m., indicating a positive correlation with altitude. MC and BD showed weak positive correlation at mid-elevation. The findings suggested altitude as a detrimental factor for carbon stocking in trees and soil. It also provide a preliminary idea on influence of other factors such as slope, grazing pressure, choice of management practices and certain climatic factors like humidity and temperature on altitude wise reduction in carbon stocking of tree species in transitional zones.

#### Keywords: Altitude, Bhabhar zone, Biomass, Carbon stock, Shorea robusta

Carbon sequestration is often considered as a leading process reducing carbon dioxide (CO<sub>2</sub>) emissions under changing climatic scenario. Trees are known to store the atmospheric carbon by sequestrating it in the growth of woody biomass through the process of photosynthesis. Since trees store and exchange carbon through vital physiological processes, they can be effectively managed to sequester significant quantities of carbon from the atmosphere. Management of Shorea robusta Gaertn., commonly known as sal, can contribute to global sustainability as the species holds immense potential in context to forest productivity and climate change. In India, the species is extended from foothills of south of Himalaya to far regions of Satpura and Eastern Ghats. In Uttarakhand, mostly found in Terai and Bhabhar regions, especially in the Shivalik hills. As a tree species in natural forests, S. robusta not only holds higher economic value but also serves as an important ecological benefit in the form of slackening global warming and climate change through sequestering atmospheric carbon dioxide (Shrestha et al 2023). It contributes to approximately 10.87% of total growing stock in country's forests (ISFR, 2021). However, its capacity to store carbon can vary along the elevation gradient which may, furthermore influence the amount of greenhouse gases from the atmosphere, an important phenomenon to slowdown the process of global warming.

Forests store approximately half of total carbon of earth and therefore, their role in climate change mitigation is widely

recognized (Ekholm 2016, Gren and Zeleke 2016). Generally considered that as a complex terrestrial ecosystem, forests play a profound role in reducing ambient CO<sub>2</sub> levels as they contribute from 20 to 100 times more carbon sequestration per unit area than agrarian lands. In addition to this, mechanism to regulate regional and global carbon (C) cycle through carbon sequestration requires information on tree carbon stock for different species in different regions with varied local environmental conditions. Due to different climatic conditions, forests are markedly different in their individual types. In India, the diversity level of subtropical forests of Northern Himalayan region in Uttarakhand is different from the subtropical forests of Arunachal Pradesh with varied rate of ecological functioning (Himshikha et al 2022). Forest biomass consists of above ground and below ground biomass (Nonini and Fiala 2019, Bhandari and Chhetri 2020). On average, 50% of the biomass is estimated as the carbon content for all species of trees. Empirical measurements of below ground biomass are time consuming, costly and difficult. Therefore, BGB is often estimated using a constant proportion of AGB. In milieu of assessing sequestration potential of any tree species, AGB and BGB need to be measured to enable better calculations of total forest carbon particularly in different geographic conditions. There have been relatively few studies on below ground biomass and carbon stock, particularly in mountainous region. There is an information gap on the altitude wise differential amount of carbon stocking

especially in *S. robusta* forests in Bhabhar area, a transitional zone of Shivalik Himalaya in Uttarakhand, India.

Among locality factors, altitude is the major factor affecting the properties of the soils in different forest ecosystems. In forests, soil is also an important pool for storing a huge amount of atmospheric carbon and twice soil organic carbon (SOC) as vegetation and two-thirds as much as the atmosphere (Shreshtha et al 2023). Soil carbon plays a critical role in the global carbon cycle and is an important tool for mitigating the effects of climate change (Fageria 2012). Sakin (2012) suggested that about 55-58% of the soil organic matter (SOM) contains SOC. Soil C stocks and forests in general, act as a carbon sink, removing CO<sub>2</sub> from the atmosphere and therefore mitigate climate change (Mauki et al 2023). The quantification from natural forests in vegetation transition zone becomes more important to estimate their potential in mitigating impacts of climate change as such sensitive zones are important for policy prescription and management planning and harboring ecological services from the species in the region. Species in transitional areas often display adaptive response and therefore, such areas may be crucial for long-term biodiversity conservation and mitigation of climate change. To assess the impact of deforestation and re-growth or regeneration rates on the global carbon cycle, it is necessary to know the stocks of carbon as biomass per unit area for forest types from different perspectives on locality factors under singular agro-climatic zone. The effective monitoring of forests in vegetation transition zone is essential to understand the carbon cycle and its' responses to climate change and anthropogenic activities. Additionally, can provide useful details on the structural and functional characteristics of forest ecosystems along the altitudinal gradient. The hypothesis is that tree and soil carbon stock in sal forest generally changes along with the change in altitude in Bhabhar zone of Uttarakhand.

#### MATERIAL AND METHODS

**Study area:** The experiment was conducted in three locations in sal (*S. robusta*) dominated forest along vertical elevation gradient (400-1300 m) in Duggada forest range from Pauri Garhwal Forest division, Uttarakhand. The area comes under tropical to sub-tropical forest zone of Shivalik Hills situated in Western Himalaya. All selected forest stands are managed by State Forest Department. The area lies between latitudes 78° 34' and 78° 37' E and longitudes 29° 46' and 29° 50' N. The elevation gradients were established through three elevation grids, the higher, mid and lower elevation as zone I (400-700 m), II (701-1000 m) and III (1001-1300 m).

Sample plot design and tree measurements: The study

employed a systematic random sampling method to collect data at each altitudinal zone. Total 30 sample plots sizes of 10 m×10 m, 10 sample plots at each altitudinal range were randomly selected for field measurement and sampling purpose of trees (Fig. 1).

For sampling of soil, 1 m x 1 m quadrates were dug to 30 cm depth to collect soil samples from each plot. Data were collected from the field survey that includes measurement of tree parameters (height and girth). Trees with  $\ge$  30 cm girth at breast height ( $\ge$ 10 cm DBH) at 1.37 m within sample plots were selected for measurement (Joshi et al 2021). For estimation of tree biomass, the quantitative data were obtained from 96 trees. Trees on the border were included if > 50% of their basal area falls within the plot and, excluded if < 50% of their basal area falls outside the plot. Determination of tree basal area and tree volume was carried out using tree form factor method.

#### **Estimation of Biomass and Carbon Content**

**Above ground biomass (AGB):** AGB is described by IPCC (2006).

AGB (t ha<sup>-1</sup>) = Stem Volume × wood density × biomass expansion factor





Fig. 1. Sample plot design for enumeration of trees and soil sampling

For the estimation of AGB, wood density scale was used as 0.72 and biomass expansion factor as 3.4 (IPCC, 2003).

**Below ground biomass (BGB):** It is recommended to estimate BGB as 26% of AGB (IPCC, 2003). IPCC (2006) has suggested carbon fraction as 0.50.

BGB (t ha<sup>-1</sup>) = AGB × conversion Factor (0.26)

**Total biomass:** After acquiring the biomass per ha, it was multiplied by a default carbon fraction of 0.5 to calculate the net carbon content (carbon stock density per ha).

Total biomass (t ha<sup>-1</sup>) = AGB + BGB

Carbon stock density (t ha<sup>-1</sup>) =Total biomass × carbon fraction

Tree/stand biomass, volume of tree stem or bole: This was calculated as:

Stem/ bole volume  $(m^3ha^{-1})$  = Form factor × Tree height × Tree basal area

Form factor: This was calculated as:

Form factor (F) = 2 H1/3H

Whereas, H1 = Tree height at which diameter is half of diameter at breast height;

H = Total height of tree

#### Soil analysis

**Soil moisture:** The soil moisture content (MC %) was calculated as follow:

**Soil organic carbon (SOC):** Walkley and Black Wet Digestion Method was used to determine SOC % (Abraham 2013).

Actual soil organic carbon: There is incomplete oxidation of organic matter in this procedure. Therefore, the organic carbon was multiplied by 1.32 on the assumption that there was 77% recovery.

Actual soil organic carbon (%) = Soil organic carbon × 1.32

Soil carbon stock = SOC (%) × bulk density × soil depth **Statistical analysis:** Data were analyzed using descriptive and inferential statistical tools on MS Excel 2010 and SPSS software.

#### **RESULTS AND DISCUSSION**

#### **Tree Carbon Stock**

**Basal area and volume:** The lowest average tree basal area and volume (7.37  $m^2ha^{-1}$  and 44.23 $m^2ha^{-1}$ ) was in the lower altitude (400-700m) followed by 11.47  $m^2ha^{-1}and$  108.58  $m^2ha^{-1}at$  middle (701-1000 m) and highest (20.27  $m^2ha^{-1}$  and 186.84  $m^2ha^{-1}$ ) in the uppermost side (1001-1300m) in the forest.

**AGB and BGB:** AGB and BGB was highest ( $288.15 \text{ t ha}^{-1}$  and  $118.92 \text{ t ha}^{-1}$ ) in the lower side, followed by mid ( $167.47 \text{ t ha}^{-1}$  and  $69.11 \text{ t ha}^{-1}$ ) and lowest values ( $66.77 \text{ t ha}^{-1}$  and  $28.14 \text{ t ha}^{-1}$ ) at higher elevations. Similarly, total tree carbon stock were as  $457.38 \text{ t ha}^{-1}$  (highest) in lower  $265.82 \text{ t ha}^{-1}$  in mid zone and,  $105.59 \text{ t ha}^{-1}$ in the uppermost elevation (Table 1).

The studied parameters decreased significantly with increasing altitude (Except tree basal area). Tree carbon stock showed strong inverse correlation with altitude, indicating a strong negative correlation with elevation gradient. It could be due to less anthropogenic disturbance and differences in temperature regime between lower (Tropical zone) and upper elevations (Subtropical zone) limiting to tree growth along the altitudinal gradient. The contribution of above-ground and below-ground biomass of trees to the total biomass in the present study was 70.79 and 29.21 % from zone I and II, while was slightly varied as 70.35 and 29.65%, respectively at zone III (Table 2). Nandal et al (2019) mentioned contribution of AGB and BGB as 81.4 to 85.9 and 14.1 to 18.6 percent in total tree biomass. The findings favor to the fact that carbon quantification varies at regional to local scale depending on microclimatic and other environmental factors followed by various silvicultural and forest management practices adopted by concerned authorities.

Table 2. Percentage of AGB and BGB for TB in study area

Zone	AGB (%)	BGB (%)	
l (400-700 m)	70.79	29.21	
ll (701-1000 m)	70.79	29.21	
III (1001-1300 m)	70.35	29.65	

<b>Table 1.</b> Average basal area, volume and total carbon stock in Shorea rol
---

Elevation zone	Basal area (m² ha⁻¹)	Volume (m³ ha⁻¹)	AGB (t ha⁻¹)	BGB (t ha <sup>-1</sup> )	TB (t ha⁻¹)	Total carbon stock (t ha⁻¹)
l (400-700 m)	20.27 ± 2.26	186.84 ± 27.73	288.15 ± 42.77	118.92± 17.65	407.07	457.38 ± 67.88
ll (701-1000 m)	11.47 ± 2.10	108.58 ± 23.57	167.47 ± 36.63	69.11 ± 15.12	236.58	265.82 ± 58.15
III (1001-1300 m)	7.37 ± 0.00	44.23± 6.51	66.77 ± 9.77	28.14 ± 4.15	94.91	105.59 ± 17.17
r value	0.99	0.99	0.99	0.99	0.99	0.99
p value (≤ 0.05)	0.13	0.04*	0.03*	0.04*	0.04*	0.03*

r = Correlation coefficient, p-value = Significance level\*

Elevation zone	Soil moisture (%)	BD (g cm) <sup>-3</sup>	SOC (%)	SOC Stock (t ha <sup>-1</sup> )
I (400-700 m)	1.94 ± 0.34	0.87 ± 0.5	1.24± 0.18	33.33 ± 6.23
II (701-1000 m)	1.43 ± 0.26	0.97 ± 0.03	1.22 ± 0.23	35.38± 5.99
III (1001-1300 m)	2.19 ± 0.23	$0.86 \pm 0.03$	1.55 ± 0.23	40.96 ± 6.66
r Value	0.32	0.08	0.84	0.97
p Value (≤ 0.05)	0.79	0.95	0.37	0.17

Table 3. Correlation of soil properties (Moisture, BD, SOC% and SOC stock) with altitude

r = Correlation coefficient, p-value = Significance level\*

#### **Soil Properties**

**Soil moisture, SOC% and BD:** The highest value of soil moisture (2.19%), soil carbon stock (40.96) and SOC% (1.55%) was at higher elevation (1001-1300m). The lowest value of soil moisture and SOC (1.43 % and 1.22%) was in the middle altitude respectively. Opposite to SOC%, the highest value of BD was in middle altitude as  $0.97g \text{ cm}^3$  whereas lowest BD (0.86g cm<sup>-3</sup>) was in higher elevation (Table 3).

SOC stock (t ha<sup>-1</sup>): Maximum amount of SOC stock (40.96 t ha<sup>-1</sup>) was estimated from upper elevation while minimum (33.33 t ha<sup>-1</sup>) at lower elevation. Soil samples from all three altitudinal gradients showed weak positive correlation between moisture, bulk density, SOC % and SOC with altitude. By depicting weak positive correlation, the statistical analysis confirmed that soil carbon stock increases as the elevation goes up. It might be possible due to change in temperature and at higher elevations, the soils generally exhibit low nutrient absorption due to increased slope. This may also happen because of the soil texture, constant input of carbon with less loss, decrease in the microbial and enzymatic activity due to the decreasing temperature (Imtimogla et al 2021). Jeyakumar et al. (2020) mentioned general trend of decreasing SOC and biomass carbon as altitude increases at higher side. Therefore, additional research efforts are required to explore effects of other locality factors on SOC content. The overall effect of elevated atmospheric CO<sub>2</sub> on soil structure is not very well understood yet and enhanced atmospheric CO<sub>2</sub> may result in the increase in photosynthesis and ensuing increase in photosynthate, roots and microbial communities and hence increased amount of SOC in certain species dominated forests such as S. robusta.

Other factors such as slope, grazing pressure, certain climatic factors such as humidity, precipitation aspect and temperature may accelerate the influence of altitude on carbon stocking in the species. One of the limiting factors might be that the upper altitude range selected for this study falls under the fire prone area of the Duggada forest range under state forest dept. The estimates acquired in the study suggest that natural population of Sal at lower elevations has the potential to store a large amount of  $CO_2$  than the

population found at upper elevations in the region. Overall, though altitude as alone, impacts negatively on carbon stocking potential of this species, the study suggest that other parameters such as stand structure, choice of management practices, slopes, vegetation dynamics, succession stages of the forest community, biological factors such as anthropogenic disturbance, management regime, grazing pressure, environmental factors such as temperature, precipitation and seasonal Indices etc. should also be considered while estimating the carbon stocking potential of tree species.

#### CONCLUSION

The findings confirmed that the elevation gradient factor significantly influences the rate of carbon sequestration in the natural population of S. robusta present at transition zone. The population from lower elevation showed potential to store a large amount of CO<sub>2</sub>. Low carbon stocking in trees present at higher side was probably due to certain limiting factors such as successional stages, stand structure, fire, and anthropogenic influence at higher side. Increased SOC content with altitude might be an outcome of decreasing temperature and less microbial activity in the forest soil present in the higher elevation. Overall, the variations in carbon stocking is attributed to altitudinal characteristics in transitional zones. Hence, periodic inclusive research with multidimensional evaluation of local factors is recommended considering each critical factor while determining carbon stock quantities of tree species in transition zones such as Bhabhar in Shivalik Himalaya.

#### REFERENCES

- Abraham J 2013. Organic carbon estimations in soils analytical protocols and their implications. *Rubber Science* **26**(1): 45-54.
- Bhandari SK and Chhetri B 2020. Individual based modeling for predicting height and biomass of juveniles of *Shorea robusta*. *Austrian Journal of Forest Science* **137**(2): 133-160.
- Ekholm T 2016. Optimal forest rotation age under efficient climate change mitigation. *Forest Policy Economics* **62**: 62–68.
- Fageria NK 2012. Role of soil organic matter in maintaining sustainability of cropping systems. Commun. *Soil science and plant analysis* **3**(16): 2063-2113.
- Gren IM and Zeleke AA 2016. Policy design for forest carbon

sequestration: A review of the literature. *Forest Policy Economics* **70**:128-136.

- Himshikha, Dobhal S, Ayate D and Lal P 2022. Influence of anthropogenic activities on the biological diversity of forest ecosystem, in: Towards Sustainable Natural Resourcesmonitoring and managing ecosystem biodiversity (M Rani, BS Chaudhary, S Jamal and P Kumar (Eds.) Springer Nature Publishers Switzerland, p 215-233.
- Imtimongla, Ariina MMS, Saya D and Phuncho T 2021. Properties of soil in relation to altitude. *Just Agriculture* **1**(12): 1-13.
- ISFR 2021. *Forest Survey of India*. (Ministry of Environment, Forest and Climate Change (GOI) **17**: 1-584.
- IPPC 2003. IPCC Good practice guidance for land use, land-use change and forestry. Institute for global environmental strategies (IGES), Japan, p 590.
- Jeyakumar SP, Dash B, Singh AK, Suyal DC and Soni R 2020. Nutrient cycling at higher altitudes in: Microbiological Advancements for Higher Altitude Agroecosystem and Sustainability R Goel, R Soni, DC Suyal (Eds.). Springer Nature, Singapore, p 293-305
- Joshi VC, Negi VS, Bisht D, Sundriyal RC and Arya D 2021. Tree biomass and carbon stock assessment of subtropical and temperate forests in the Central Himalaya, India. *Trees, Forests*

Received 11 July, 2024; Accepted 12 September, 2024

and People 6: 1-8.

- Mauki D, Richard U and Kilonzo M 2023. Influence of elevation gradient and plant species composition on soil organic carbon in mountain Rungwe Forest reserve, Tanzania. *Environmental and Sustainability Indicators* **19**
- https://doi.org/10.1016/j.indic.2023.100291 accessed on 12. 01.2024
- Nandal A, Singh N, Yadav SS, Rao S and Yadav V 2019. Carbon stock assessment of selected tree species n Maharashi Dayanand University Campus, Rohtak (Haryana) India. Indian Journal of Ecology 46 (2): 39-45
- Nonini L and Fiala M 2019. Estimation of carbon storage of forest biomass for voluntary carbon markets: preliminary results. *Journal of Forestry Research* **32**(1): 329-338
- Sakin E 2012. Organic carbon matter and bulk density relationships in arid-semi arid soils in southeast Anatolia region. *African Journal of Biotechnology* **11**(6): 1373-1377
- Shrestha S, Gautam TP, Raut JK, Goto BT, Chaudhary S and Mandal TN 2023. Edaphic factors and elevation gradient influence arbuscular mycorrhizal colonization and spore density in the rhizosphere of *Shorea robusta* Gaertn. *Acta Ecologica Sinica*.https://doi.org/10.1016/j.chnaes.2023.05.011 accessed on 10.01.2024.



Manuscript Number: 4337 NAAS Rating: 5.38

# Variation in Fruit and Seed Morphology of *Pyrus pashia* (Buch-Ham ex D. Don) in Alaknanda Valley of Garhwal Himalaya, India

#### Amreen, A.K. Negi and Himshikha Gusain

Department of Forestry and Natural Resources H.N.B. Garhwal University, Srinagar, Garhwal-246 174, India E-mail: amreenansari1990@gmail.com

**Abstract:** The study was conducted on fruit and seed morphology of *Pyrus pashia* populations. Seven characters (fruit length, width, thickness, seed length, width, thickness and seed number) were measured from 450 trees in 15 different locations at altitude ranging from 650 to 2190 metres. The altitudes were grouped as follows: the first group (650 m, 1790 m), third group (1260 m, 1460 m, 2100 m), and the second cluster containing the remaining altitude locations. The coefficient of variation confirmed significant difference of morphological characters. The correlation between morphological characteristics of fruits and seeds and geographic features showed a negative and weak correlation with altitude and latitude. Furthermore, altitude negatively correlates with fruit length, seed number and seed length morphological traits but positively with others, emphasizing the need to consider specific parameters when selecting germplasm collection sites. Among different locations, altitude of 1220 meter showed that the seed and fruit characteristics confirm it as the best germplasm collection site.

#### Keywords: Pyrus pashia, Himalaya, Fruits, Seeds, Morphology, Populations

Understanding the morphology of fruits and seeds is fundamental in identifying the Indian wild pear P. pashia (Buch-Ham ex D. Don), a highly valued deciduous tree species (Arya et al 2011). This significant intermediate species between the oriental and occidental groups of pears has played a crucial role in forming the Pyrus genus. The round, brown fruits are edible and have high nutritional and ethnomedicinal value. Depending on the topographic and climatic conditions, the fruit's colour changes from willow green to brown with light spots in the middle stage and eventually turns black when ripe (Prakash et al 2021a). Each fruit contains around five seeds, which are black in colour, tiny, light, and pyriform in shape. A mature tree may produce up to 45 kg of fruit per year, with a taste ranging from rigorous to sweet and gritty. Approximately 50 seeds are available per gram (Prakash et al 2021b). The oval, hairless and shiny leaves are accompanied by clusters of white flowers, adding to its aesthetic appeal. Pyrus species face threats worldwide that have caused the loss of 85% of numerous pear varieties in 19th century or even today (Sindelar 2002). Considering the variations in fruit and seed traits is difficult for determining the best geographic source of seeds (Mkwezalamba et al 2015). This study aimed to evaluate altitudinal variation in fruit and seed morphology of Pyrus pashia in Alaknanda valley of Garhwal Himalaya, India.

#### MATERIAL AND METHODS

Sampling location: The study was conducted during October to December 2020 and 2021 in the Alaknanda

Valley, covering an area of approximately 1400 km<sup>2</sup> in the Garhwal Himalayan region, India. Based on the altitudinal gradient, fifteen different locations (L1 to L15) ranged from 650 to 2190 m were randomly selected, all of which had wild populations (Table 1). Samples were collected from 30 randomly selected trees in each location, making a total of 450 trees.

The 5 kg of fully ripened fruits was collected from October to December 2020-21. Two to three branches per tree were harvested at mid-height to reduce individual variation and after the initial fruit screening, 100 ripened and undamaged fresh fruits were measured. After de-pulping, the seeds were extracted from the fruits and sun-dried for further tests. The comprehensive data on seven parameters, fruit length, width, and thickness, seed length, width, thickness, and number of seeds/fruits, were recorded.

**Statistical analysis:** The statistical was done using R.4.1.1 (Cluster analysis) and SPSS 4.9 (Correlation matrix). The Minitab-18 statistical software was used for one way analysis.

#### **RESULTS AND DISCUSSION**

**Morphological characteristics of fruits of** *P. pashia*: There was high degree of variation in its phenotypic characteristics. The fruit length varied among locations, ranged from 16.29 mm for L6 to 24.93 mm for L2 (1220 m). Similarly, the mean fruit width varied from 25.97 mm for L2 to 17.86 mm for L6 (1420 m). However, the fruit thickness ranged from 26.01 mm for L8 (1465 m) to 17.94 mm for L6.
The length and width of the fruit were lower at both higher elevations (2190 m) and the lowest elevation (650 m). The L2 (1220 m) has shown superior fruit size of 24.93 mm and maintained its superiority over other locations. L3 (1260 m) consistently underperformed in all traits of *P. pashia* fruit. Kishore et al (2017) also observed the smallest fruit size of *P. pashia* at 1350 m. The shape of the fruit (Round, oblong and pyriform) varied, with the maximum fruit length (24.93 mm) and fruit width (25.97 mm) at an elevation of 1220 m. These

findings suggest that the population from this area can be utilized for genetic and tree breeding programs (Table 2). Lone et al (2000) observed significant variation in fruit traits in temperate fruits. These findings suggest that the quantity of *P. pashia* fruit seeds tend to decrease with altitude. Shankar et al (2012) also observed considerable variation in the morphological traits of fruits and seeds of *P. nepaulensis* among different locations. The lowest diameter in the fruit at the middle altitude of 1420 m might be due to the fruit's

	Table 1. Geographical	coordinates of stud	y sites in Alaknanda	valley, Garhwal Himalaya
--	-----------------------	---------------------	----------------------	--------------------------

Location code	Name	Altitude (m)	Latitude	Longitude
L1	Luneta	650	30°12'32.39" N	78°40' 35.04" E
L2	Kund	1220	30°30'20.11" N	79°05' 29.87" E
L3	Silyara	1260	30°27'14.01" N	78°38' 56.62" E
L4	Kuhed	1300	30°22'35.51" N	79°19' 37.98" E
L5	Gadora	1400	30°24'56.35" N	79°25' 50.34" E
L6	Kauliyadhar	1420	30°17'37.18" N	78°50' 22.17" E
L7	Kandikhal	1460	30°20'41.71" N	78°37' 50.67" E
L8	Barsori	1465	30°09'26.68" N	78°48' 31.57" E
L9	Chunikhal	1630	30°18'10.75" N	78°48' 43.70" E
L10	Agroda	1700	30°07'08.46" N	78°47' 49.84" E
L11	Chandrabadni	1780	30°18'20.90" N	78°37' 38.93" E
L12	Gopeshwar	1790	30°25'12.53" N	79°19' 16.41" E
L13	Tapowan	1970	30°30'02.40" N	79°37' 31.34" E
L14	Nauti	2100	30°12'43.59" N	79°12' 26.96" E
L15	Tala	2190	30°30'32.69" N	79°10' 14.24" E

Table 2. Variation in morphological characteristics (Length, width and thickness) of fruits of *Pyrus pashia* (Mean ± SD)

Location code	Altitude (m)	Fruit length (mm)	Fruit width (mm)	Fruit thickness (mm)
L1	650	17.94±0.48	18.75±0.53	18.70±0.59
L2	1220	24.93±0.79	25.97±0.78	25.76±0.69
L3	1260	17.81±0.50	19.01±0.49	19.19±0.50
L4	1300	20.89±0.05	23.86±0.39	23.83±0.34
L5	1400	22.62±0.37	19.82±0.34	19.53±0.43
L6	1420	16.29±0.78	17.86±0.70	17.94±0.72
L7	1460	19.39±0.21	20.07±0.29	21.95±0.00
L8	1465	20.70±0.02	24.12±0.44	26.01±0.74
L9	1630	20.89±0.05	23.41±0.31	23.42±0.27
L10	1700	20.19±0.07	21.94±0.04	21.95±0.00
L11	1780	20.07±0.09	21.46±0.04	21.42±0.09
L12	1790	22.06±0.26	20.83±0.15	20.64±0.23
L13	1970	22.10±0.27	22.07±0.06	22.10±0.03
L14	2100	21.90±0.24	23.87±0.39	23.68±0.31
L15	2190	21.07±0.08	22.43±0.13	22.84±0.16
F		56.43**	63.45**	62.78**

Statistically significant difference, \*\*=P<0.05

maturity. Altitude and other local factors cause a net decline in the variation between day and night temperatures, thus modifying the characteristics of inland growing areas. However, it is suggested that altitude alone is not the only factor affecting these variations.

Morphological characteristics of seeds of P. pashia: The variation in the seed size and number can affect a species reproductive ability. In wild fruits, the mean seed length ranged from 1260 m to 1630 m. The maximum seed width was in L11 (1780 m- 3.83 mm), while the minimum seed width was in L9 (1630 m- 2.92 mm), followed by L1 and L14.The average number of seeds/fruits differed from 5.97 for L2 (1220 m) to 4.76 for L3 (1260 m). This confirmed a significant change in altitude-wise seed morphology in terms of mean seed width and thickness (Table 3). The increasing trend in the morphological values of seeds up to the middle elevation (1420 m) was observed except for 1220 m. This variation in seed morphology may also be due resources, which change from season to season. The observed variations in seed characteristics of wild pear accessions could also be due to hybridization, sexual reproduction, bud mutation, and diverse agroecological conditions. Similar observations were observed wild, primitive varieties and landraces of other temperate fruits (Zafar et al 2004).

Correlation between the morphological features of *P. Pashia* to geographical distribution: The size of the fruit and seed mass of *P. pashia* increased as longitude increased which differed from Baker's rule. The sample locations show a strong correlation between altitude and longitude, which suggesting that the fruit of *P. pashia* has evolved to adapt to different latitudes and longitudes. Fruit length showed positive linear regression coefficient associated with fruit width, thickness and number of seeds/fruits. Fruit length displayed a weak correlation with the length, width and thickness of seeds. The width of the fruits was highly significant with fruit thickness and seed width the correlation between morphological characteristics of fruits and seeds and geographic characteristics showed a negative and weak correlation with altitude and latitude. Fruit thickness, however, significantly positively correlated with altitude and longitude and was negatively correlated with latitude. Number of seeds/fruits did not relate to seed length, width, and thickness but showed a significant correlation with longitude and latitude. There is no linear relationship, either increasing or decreasing, of selected morphological characters of fruits and seeds of P. pashia to its distribution along all gradients (Table 4). The differences in the physical characteristics of seeds and fruits may be due to natural limitations in their location. Environmental factors resulting from variations in latitude and altitude, including soil type, human activity, and wind patterns, along with different communities of dispersers separately and together might be responsible for the differences in fruit size in P. Pashia across or between species.

 Table 3. Variation in morphological characteristics (Length, width, thickness and number of seeds/fruits) of seeds of Pyrus pashia (Mean ± SD)

Location code	Altitude (m)	Seed length (mm)	Seed width (mm)	Seed thickness (mm)	Number of seeds/fruits
L1	650	6.03±0.03	3.05±0.06	2.11±0.02	5.20±2.19
L2	1220	6.53±0.05	3.74±0.06	2.39±0.08	5.97±2.12
L3	1260	6.88±0.1	3.78±0.06	2.02±0.01	4.76±2.27
L4	1300	5.96±0.05	3.52±0.02	2.01±0.01	5.57±2.51
L5	1400	5.98±0.04	3.56±0.03	2.19±0.04	5.32±2.56
L6	1420	6.20±0.00	3.64±0.04	1.78±0.03	5.48±2.21
L7	1460	6.25±0.00	3.29±0.02	1.83±0.02	5.69±2.27
L8	1465	6.24±0.00	3.27±0.02	1.99±0.00	5.46±2.32
L9	1630	5.38±0.15	2.92±0.08	1.54±0.07	5.62±1.57
L10	1700	6.30±0.01	3.55±0.02	1.90±0.00	5.71±1.77
L11	1780	6.46±0.03	3.83±0.08	1.97±0.00	5.50±2.50
L12	1790	5.68±0.10	3.25±0.02	2.12±0.03	5.69±2.07
L13	1970	6.45±0.03	3.18±0.03	2.09±0.02	5.76±2.97
L14	2100	6.75±0.09	3.06±0.06	1.68±0.04	5.59±2.25
L15	2190	6.61±0.06	3.27±0.0	1.61±0.06	5.55±2.36
F		43.95*	30.32**	246.47**	219.99**

Statistically significant difference, \*=P<0.01, \*\*=P<0.05

**Principal component analysis (PCA) and hierarchical cluster analysis between morphological traits and locations:** The populations were classified into three main groups, with group 3 being the largest, consisting of 10 populations. The first three principal components (PC1, PC2, and PC3) explained 73.6% of the location changes. PC1's first component accounted for 37.91% of the total variation, followed by the second component which accounted for 19.52%. Through hierarchical cluster analysis, three primary groups of locations were identified based on their similarity. The first group consisted of L1 and L12 (650 m, 1790 m) and the third cluster comprised L3, L7, and L14 (1260 m, 1460 m, 2100 m) and the remaining 10 location formed the second cluster.

The lowest fruit length, weight, and thickness values were recorded at middle elevations around 1440-1600 meters. However, these findings partially contradict earlier observations that fruit size increases with elevation (Dinis et



**Fig. 2.** Dendrogram (Hierarchical clustering) of 15 locations of *Pyrus pashia* based on fruits and seeds morphological characters



Fig. 1. Principal component analysis (PCA) of fruits and seed characteristics at different locations (a) and projection of morphological characters (b) on the space formed by the first three axes

Table 4. Pearson's correlation	coefficient relationship betwe	en locations parametei	rs concerning fruit and	d seed morphological
traits of Pyrus pashia				

Characters	1	2	3	4	5	6	7	8	9
Fruit length 1	1								
Fruit width 2	0.621**	1							
Fruit thickness 3	0.444*	0.521**	1						
Number of seeds/fruits 4	0.837**	0.317	0.151	1					
Seed length 5	0.134	0.207	0.315	0.205	1				
Seed width 6	0.165	0.403*	0.426*	0.232	0.028	1			
Seed thickness 7	0.041	0.079	0.683	0.304	0.149	-0.538	1		
Altitude 8	-0.328	0.373	0.725*	-0.726*	-0.261	0.628*	0.564	1	
Latitude 9	0.514	0.417	0.801**	0.768*	0.703*	-0.508	0.650	0.416	1
Longitude 10	-0.458	0.137	-0.780**	0.805*	0.695	0.255	-0.634	-0.380	-0.840*

Statistically significant difference, \*=P<0.01, \*\*=P<0.05

al 2011, Trad et al 2013, Silvanini et al 2014, Maro et al 2014). This can be ascribed to the different climatic and soil conditions at the three monitored altitudes. Contrasting results in previous studies could be due to differences in the altitude of the studied areas. The results demonstrate considerable morphological diversity in fruit and seed characteristics. This was evident from the PCA and hierarchical clustering trends. However, despite being geographically distant, populations belonging to the same group were morphologically similar (PCA and Cluster dendrogram). This highlights the importance of considering location and geographical origins when analysing such data. These findings indicate that location is crucial in determining the physical features of fruits and seeds. Brenes-Gamoba (2017) observed the need to consider the specific characteristics of each site, such as soil slope, acidity, organic matter content, and the impact of climatic factors, when comparing different altitudes. Furthermore, optimal environmental conditions are necessary to achieve the highest possible yields and qualities from fruit plantations, conditions that are linked to the genetic potential of the species (Perez and Melgarejo 2015).

#### CONCLUSIONS

The altitude of 1220 m exhibited the most considerable fruit length, width, and seed thickness, making the ideal location for collecting germplasm for propagation and germination to achieve the best fruit quality. Correlation coefficient, PCA, and hierarchical clustering analyses confirmed that fruit width, thickness, altitude, and number of seeds/fruits are crucial factors in determining the optimal altitude for germplasm collection, especially when fruit quality is the primary consideration. The study found a negative and weak correlation between the morphological characteristics of fruits and seeds and geographic features (latitude and longitude). Despite the altitude, seed morphology varied widely and irregularly, indicating that specific morphological parameters should be considered when selecting suitable germplasm of *P. pashia* from different location.

### REFERENCES

Arya V, Gupta R and Gupta VK 2011. Pharmacognostic and

Received 06 July, 2024; Accepted 21 September, 2024

phytochemical investigations on *Pyrus pashia* Buch. - Ham. ex D. Don stem bark. *Journal of Chemical Pharmaceutical Research* **3**(3): 447-456.

- Brenes-Gamboa S 2017. Parámetros de producción y calidad de loscultivares de banano FHIA-17, FHIA-25 y Yangambi. *Agronomia Mesoamericana* **28**(3):719-733.
- Dinis LT, Peixoto F, Pinto T, Costa R, Bennett RN and Gomes-Laranjo J 2011. Study of morphological and phenological diversity in chestnut trees ('Judia' variety) as a function of temperature sum. *Environmental and Experimental Botany* **70**: 110-120.
- Kishor Arun, Verma SK, Brijwal Manoj, Kumar Anil, Attri BL, Narayan Raj and Debnath Sovan 2017. Evaluation of Genetic Diversity in Wild Pear (*Pyrus pashia*) under Kumaon Hills of Uttarakhand. *Environment & Ecology* **35**(1B): 524-529.
- Lone AF and Wafal BA2000. Varietal diversity in the germplasm of cherries under cultivation in Kashmir. In: Khan MA, Farooq S. (eds). *Environmental Biodiversity and Conservation* 319-340.
- Maro LAC, Pio R, Guedes MNS, Abreu CMP and Moura PHA 2014. Environmental and genetic variation in the post-harvest quality of raspberries in subtropical areas in Brazil. Acta Scientiarum Agronomy 36(3): 323-328.
- Mkwezalamba Idah, Munthali RV Chimuleke and Missanjo Edward 2015. Phenotypic Variation in Fruit Morphology among Provenances of Sclerocarya birrea (A. Rich.) Hochst. International Journal of Forestry Research, Doi.org/10.1155/ 2015/735418
- Perez LV and Melgarejo LM 2015. Photosynthetic performance and leaf water potential of gulupa (*Passiflora edulis* Sims, Passifloraceae) in the reproductive phase in three locations in the Colombian Andes. *Acta Biológica Colombiana* **20**(1): 183-194.
- Prakash Om, Chauhan AT and Kudachikar VB 2021a. Traditional uses, nutrition, phytochemistry and various pharmacological properties of Indian wild pear (Review), *International Journal of functional nutrition* 2:9.
- Prakash Om, Selvi Karthika Moorthy, Vijayaraj Panneerselvam and Kudachikar VB 2021b. Lipidome, nutraceuticals and nutritional profiling of *Pyrus pashia* Buch-Ham ex D. Don (Kainth) seeds oil and its antioxidant potential. *Food chemistry* 338.
- Shankar Uma and Synrem L Idaiarilin 2012. Variation in morphometric traits of fruits and seeds of *Prunus nepaulensis* Steud. in Meghalaya, India. *Tropical Ecology* **53**(3): 273-286.
- Silvanini Annalisa, Dall'Asta Chiara, Morrone Lucia, Cirlini Martina, Beghe Deborah, Fabbri Andrea, Ganini Tommaso 2014. Altitude effects on fruit morphology and flour composition of two chestnut cultivars. *Scientia Horticulturae* **176**: 311-318.
- Sindelar J 2002. Toward threatened forest tree species preservation on the example of crab apple (*Malus sylvestris* L.) and wild pear (*Pyrus pyraster* L. [Burgsdorf]). *Zprav Lesnic Vyzk* **47**: 199-203.
- Trad M, Gaaliche B, Renard CMGC and Mars M 2013. Inter and intratree variability in quality of figs. Influence of altitude leaf area and fruit position in the canopy. *Scientia Horticulturae* **162**: 49-54.
- Zafar G, Mir MS and Sofi AA 2004. Genetic divergence among apricot (*Prunus armeniaca* L.) genotypes of Kargil, Ladakh. *Indian Journal of Horticulture* **61**: 6-9.



# Wood-rotting Macrofungi of Kikruma Community Forest, Phek, Nagaland

# Kuno Chuzho and M.S. Dkhar<sup>1</sup>

Department of Botany, University of Science & Technology Meghalaya, Baridua-793 101, India <sup>1</sup>Department of Botany, North-Eastern Hill University, Shillong-793 022, India E-mail: kunochuzho@gmail.com

**Abstract:** Survey and documentation of wood-rotting macrofungi of Kikruma community forest, Phek, Nagaland was carried out for period of three years from January 2015 to December 2017. The identified wood-rotting macrofungi are classified under two phyla, sixteen families, thirty one genera and forty three species. Thirty six wood-rotting macrofungi belonged to phylum Basidiomycota and the remaining seven to phylum Ascomycota. Polyporaceae was the most dominating Family with fourteen wood-rotting macrofungi, followed by Stereaceae and Xylariaceae with five wood-rotting macrofungi each. For seasonal variation, Pre monsoon (March to April), Monsoon (may to august), Retreating Monsoon (September to November) and Dry Season (December to January) were considered. Majority of the species were sampled during retreating monsoon season followed by monsoon and pre monsoon seasons respectively. Three species, *Auricularia polytricha, Lopharia cinerascens* and *Schizophyllum commune* were found growing in all the four seasons. The species dominance index, species evenness and Shannon's diversity index were calculated to be 0.064, 0.517 and 3.077 respectively.

Keywords: Basidiomycota, Diversity, Edible wood-rotting macrofungi, Polyporaceae, Seasonal variation

Wood-rotting fungi comprised of a largely unexplored group of magnanimous macrofungi. Their role and mechanisms involved in wood biodegradation is studied in detail however, the full potential of majority of the species still remain unexplored. They are the only known group of living organism actively involved in wood biodegradation. Singh et al (2016) reported the ability of bacteria to degrade wood however their ability to degrade wood is much slower as compared to basidiomycete wood biodegradation. Woodrotting fungi degrade the complex components of the plant cell wall employing various complex enzymatic degradation and reactive oxygen species (Riley et al 2014, Castano et al 2018, Goodell 2020). White rot fungi have the ability degrade a number of persistent environmental pollutants such as chlorinated aromatic compounds, hetero-cyclic aromatic hydrocarbons, dyes and synthetic polymers (Bennett 1994). In addition to biodegradation of wood, this group of macrofungi play important roles in balancing ecological services, improving soil fertility, in bioremediation and also many species are known for their medicinal and nutritive properties. It has been reported that lignicellulose hydrolytic enzyme machinery of Basidiomycota members makes low cost bioremediation projects more attractive (Sanchez 2008). Wood-rotting macrofungi are reported to be the only group of organism able to decompose all woody components and thus, they play an important role in both carbon and nitrogen cycles by degrading non-living organic substrates (Pinar and Rodriguez-Conto 2024). The use of polypore fungi such as *Ganoderma lucidum*, *Phellinus linteus* and *Trametes versicolor* in the treatment a number of diseases including hypertension, prostate cancer, as cancer chemotherapy agents and their antibacterial, antifungal and antiviral effects have been studied by many researchers and reported positive results (Hsieh and Wu 2001, Silva 2003, Liang et al 2010).

Kikruma village is well-known for its rich cultural and patriotic history. It is located under Phek administrative district of Nagaland, Northeast India. Studies on wood-rotting macrofungi in northeast India is still at its dormant phase and majority of the region is still unexplored. The forest canopy of Kikruma community forest is neither open nor dense but it is of moderate type. The present study provides a preliminary documentation of wood-rotting macrofungi from Kikruma village, Phek, Nagaland.

#### MATERIAL AND METHODS

Survey and documentation of wood-rotting macrofungi of Kikruma Community forest, Phek district, Nagaland was carried out for period of three years from January 2015 to December 2017 on seasonal basis. The village is located at an altitudinal range of 1,581 to 1,650 meters above sea level (masl) with the geographical coordinates 2537.38' N and 094° 13.71'E. The village is surrounded by broad-leave deciduous forest with a vegetation hugely dominated by

Quercus serrata distributed all along the altitudinal gradient interfered with few patches of Pinus wallichiana and Prunus cerasoides. Based on the prevailing seasons, samplings were conducted during Pre monsoon (March to April), Monsoon (May to August), Retreating Monsoon (September to November) and Dry Seasons (December to January). Depending on the slope of the areas, both transect (50x5m) and quadrat methods (50x50m) were used for sampling wood-rotting fungi following the protocol recommended by Mueller et al (2004). The transects and quadrats were laid randomly. The collected basidiocarps were bought to the laboratory and dried under light to moderate sunlight condition. The dried basidiocarps were then kept in clean and labelled polythene bags with naphthalene balls wrapped in cotton. For microscopic studies, 4% KOH, lactophenol cotton blue and Melzer's reagent were used for staining, observation and identification. Identification was done through comparative consultations from the available monograph and literatures (Bakshi 1971, Ryvarden and Johansen 1980, Gilbertson and Ryvarden 1987, Nunez and Ryvarden 2000, 2001, fungifromindia.com, mycobank.org and indexfungorum.org). Diversity indices such as Dominance index (D), Evenness (e<sup>+</sup>H/S) and Shannon's diversity index (H') were used for calculating the diversity of wood-rotting fungi using PAST 1.93 software. All the identified wood-rotting fungi were deposited in Department of Botany, North-Eastern Hill University (NEHU), Shillong Campus, Meghalaya.

#### **RESULTS AND DISCUSSION**

Forty three wood-rotting macrofungi were collected and identified. The identified wood-rotting macrofungi are classified under two phyla (Basidiomycota and Ascomycota) with a total of sixteen families, thirty one genera and forty three species. The phylum Basidiomycota comprised of fourteen families, twenty eight genera and thirty six woodrotting macrofungi whereas, the phylum Ascomycota comprised of two families, three genera and seven species (Table 1). Polyporaceae was the most dominating Family with fourteen wood-rotting macrofungi, followed by Stereaceae and Xylariaceae with five wood-rotting macrofungi each (Fig. 1). Majority of the species were sampled during retreating monsoon season followed by monsoon and pre monsoon seasons respectively. Thirty six wood-rotting fungi were found growing during retreating monsoon, thirty two during monsoon, seventeen during premonsoon and only three during dry season.

Three species, Auricularia polytricha, Lopharia cinerascens and Schizophyllum commune were found growing in all the four seasons studied. Five species,

Auricularia auricula-judae, A. polytricha, Lentinussajor-caju, Pleurotus ostreatus, Schizophyllum commune and one species belonging to the genus Ramaria are reportedly consumed by the localities of the village as edible macrofungi. The remaining wood-rotting fungal species are identified as non-edible. The species dominance index, species evenness and Shannon's diversity index were calculated to be 0.0638, 0.5165 and 3.077 respectively. Few selected photographs of wood-rotting macrofungi sampled from Kikruma community forest (reproduced from Chuzho 2021), are shown in Figure 2.

Study of wood-rotting macrofungi was for the first time documented from Kikruma village, Phek, Nagaland through the present study however, studies on wood-rotting macrofungi, particularly, members of Basidiomycota is well documented in India by many workers (Leelavathy and Ganesh 2000, Forutan and Jafary 2007, Swapna et al 2008, Ranadive et al 2011, Ranadive 2013, Lyngdoh 2014, Adarsh et al 2018, Vabeikhokhie et al 2019a,b, Ao and Deb 2019, Chuzho 2021, Chungreiliu 2023). The family Polyporaceae is the most dominant taxon at level as compared to the other families recorded in the present study. The dominance Polyporaceae members have also been reported by many researchers (Sharma 2007, Prasher and Ashok 2013, Tapwal et al 2013, Lyngdoh 2014, Chuzho 2021, Chungreiliu 2023). Most of the polyporaceae members are characterized by having rigid and corky basidiocarps and the rigidity of the basidiocarps have attributed to their persistence thereby increasing their chances of being frequently encountered unlike other ephemeral wood-rotting macrofungi with soft basidiocarps which persist only for one to few days. Lyngdoh and Dkhar (2018) study on the decay potential of four woodrotting fungi on Batula alnoides and Quercus dealbata woodblocks showed that polypore species have higher degradation ability as compared to non-polypore species. The ability of polypores to degrade lignin more effectively might have helped them to be better opportunist on wood degradation and thereby increasing their chance to colonize woody debris more effectively.

Occurrence of wood-rotting fungi varied with different seasons. Majority of wood-rotting fungi were recorded during retreating monsoon season followed by monsoon season, pre monsoon and dry season. Baptista et al (2010) also showed a bimodal pattern in fruiting phenology. They observed that basidiocarp formation occurred in two distinct seasons (retreating monsoon and pre-monsoon) with the higher number of species collected during retreating monsoon than during pre-monsoon. The present result showed that maximum wood-rotting fungi occurred during retreating monsoon followed by monsoon which slightly

Family	Specimen name		Sea	son*	
	-	1	2	3	4
Phylum Ascomycota					
Hypoxylaceae	Daldinia concentrica (Bolton) Ces. & De Not.	+	+	+	-
	U1 01 Pul - Hypoxylaceae	+	-	-	-
Xylariaceae	Kretzschmaria deusta (Hoffm.) P. M. D. Martin	-	+	+	-
	<i>Xylaria grammica</i> (Mont.) Mont.	-	+	+	-
	X. hypoxylon (L.) Grev.	+	+	+	-
	X. longipes Nitschke	+	+	+	-
	X. polymorpha (Pers.) Grev.	+	+	+	-
Phylum Basidiomycota					
Agaricaceae	Cyathus striatus (Huds.) Willd.	+	+	-	-
Auriculariaceae	Auricularia auricula-judae (Bull.) J. Schrot.	-	+	+	-
	A. delicata (Mont.) Henn.	+	+	+	-
	A. polytricha (Mont.) Sacc.	+	+	+	+
Dacrymycetaceae	Dacryopinax spathularia (Schwein.) G.W. Martin	-	+	+	-
Fomitopsidaceae	Daedalea quercina (L.) Pers.	-	-	+	-
	Fomitopsis pinicola (Sw.) P. Karst.	+	+	+	-
Ganodermataceae	Ganoderma lucidum (Curtis) P. Karst.	-	+	+	-
Gloeophyllaceae	Gloeophyllum sepiarium (Wulfen) P. Karst.	-	+	-	-
Gomphaceae	Ramaria sp.	-	+	-	-
Hymenochaetaceae	Hymenochaete tabacina (Sowerby) S.H. He & Jiao Yang	-	-	+	-
	Phellinus gilvus (Schwein.) Pat.	-	+	+	-
	Phellinus sp.	-	-	+	-
Phanerocahetaceae	Bjerkandera adusta (Willd.) P. Karst	+	+	+	-
Pleurotaceae	Pleurotus ostreatus (Jacq.) P. Kumm.	-	+	+	-
Polyporaceae	Coriolopsis polyzona (Pers.) Ryv.	-	-	+	-
	Heterobasidion insulare (Murrill) Ryv.	-	+	-	-
	Hexagonia tenuis (Hook.) Fr.	-	+	+	-
	Lentinus sajor-caju (Fr.) Fr.	-	-	+	-
	Lopharia cinerascens (Schwein.) G. Cunn.	+	+	+	+
	Microporus xanthopus (Fr.) Kuntze	+	+	+	-
	Nigrofomes melanoporus (Mont.) Murrill	-	+	-	-
	N.vinosus (Berk.) Murrill	+	+	+	-
	Pycnoporus cinnabarinus (Jacq.) P. Karst.	-	-	+	-
	P. sanguineus (L.) Murrill	-	-	+	-
	Rigidoporus microporus (Sw.) Overeem	-	-	+	-
	Trametes hirsuta (Wulfen) Pil.	+	+	+	-
	T. versicolor (L.) Lloyd	+	+	+	-
	Trichaptum abietinum (Pers. ex. J.F. Gmel.) Ryv.	-	+	+	-
Psathyrellaceae	Coprinellus micaseus (Bull.) Fr.	+	+	+	-
Schizophyllaceae	Schizophyllum commune Fr.	+	+	+	+
Stereaceae	Aleurodiscus ahmadii (Biodin) Biodin	+	+	-	-
	Stereum complicatum (Fr.) Fr.	-	+	+	-
	S. gausapatum (Fr.) Fr.	-	-	+	-
	S. hirsutum (Willd.) Pers.	-	+	+	-
	S. ostrea (Blume& T. Nees)	-	-	+	-

Table 1.	Wood-rotting	fungi, their	families and	distribution	at different seasons
----------	--------------	--------------	--------------	--------------	----------------------

\*1 - Pre monsoon (March to April); 2 - Monsoon (May to August); 3 - Retreating Monsoon (September to November); 4 - Dry Season (December to January)

differs from the observation made by Baptista et al (2010), however, in both cases, maximum number of wood-rotting macrofungi occurred during retreating monsoon. Ho et al (2002), Pouska et al. (2010) and Adarsh et al. (2015) also reported the maximum occurrence of basidiocarps of agarics and polypores during August to October which supports the present finding. Basidiocarp formation of wood-rotting fungi



Fig. 1. Wood-rotting fungi belonging to different families

required high level of water supply and seasonal drought can limit fruiting of wood-rotting fungi to a great extent (Olou et al 2019). Three wood-rotting fungi viz., A. polytricha, L. cinerascens and S. commune were sampled from all the four seasons studied. This finding indicated that these species have wide range of seasonal adaptation. Nagadesi and Arya (2014a) and Lyngdoh (2014) also reported the wide range of seasonal adaptation of certain wood-rotting fungal species including S. commune. Apart from seasonal variations and precipitation, occurrence of wood-rotting macrofungi depends on other environmental factors such as altitude, type of forest vegetation, presence of host tree species and other micro- and macro-environmental factors (Chuzho and Dkhar 2019a,b). The lower number of wood-rotting macrofungi sampled in three years survey may be because Kikruma village is dominated by Q. serrata and thus it mostly harbours those species that grow on Q. serrata. Northeast India is a major storehouse for biodiversity. The exploring into wood-rotting macrofungal communities from Nagaland and other North-eastern states of India, the actual figure macrofungal bioresources of Northeast India is expected to increase drastically.



Rigidoporus microporus

Schizophyllum commune

Stereum ostrea

Fig. 2. Selected photographs of wood-rotting macrofungi sampled from Kikruma community forest (Photographs reproduced from Chuzho 2021)

### CONCLUSION

The present study provided a preliminary documentation of wood-rotting macrofungi from Kikruma village, Phek, Nagaland. Forty three wood-rotting macrofungi, classified under two phyla, sixteen families, thirty one genera were collected and identified from the present study. Majority of the species were sampled during retreating monsoon season followed by monsoon and pre monsoon seasons respectively. The Shannon's diversity index with a value of 3.077 showed that Kikruma community forest harboured high wood-rotting macrofungal diversity.

#### REFERENCES

- Adarsh CK, Kumar V, Vidyasagaran K and Ganesh PN 2015. Decomposition of wood polypore fungi in the tropics - biological, ecological and environmental factors: A case study. *Research Journal of Agricultural and Forestry Sciences* 3: 15-37.
- Adarsh CK, Vidyasagran K and Ganesh PN 2018. A checklist of polypores of Kerela state, India. *Studies in Fungi* **3**(1): 202-228.
- Ao T and Deb CR 2019. Wild mushrooms of Nagaland: An important bio-resource. *Studies in Fungi* **4**(1): 61-78.
- Bakshi BK 1971. Indian Polyporaceae (on trees and timber). ICAR, New Delhi, pp. 1-246.
- Baptista P, Martins A, Tavares RM and Lino-Neto T 2010. Diversity and fruiting pattern of macrofungi associated with chestnut (*Castinea sativa*) in the Tras-os-Montes region (Northeast Portugal). *Fungal Ecology* **3**: 9-19.
- Bennet JW 1994. Prospectus for fungal bioremediation of TNT munition wastes. International Bio-deterioration and Biodegradation 34: 21-24.
- Castano JD, Zhang J, Anderson CE and Schilling JS 2018. Oxidative damage control during decay of wood by brown rot fungus using oxygen radicals. *Applied & Environmental Microbiology* **84**(22): e10937-18.
- Chungreiliu AK 2023. Studies on Diversity of wood-rotting fungi with special emphasis on seleted edible fungi in different forest stands of Manipur. Ph. D. Thesis. North-Eastern Hill University, Shillong, India.
- Chuzho K and Dkhar MS 2019a. Diversity of Ascomycetous woodrotting fungi along an altitudinal gradient in forests of Nagaland and first report of *Jackrogersella minutela* from India. *Journal of The Indian Academy of Wood Sciences* **16**(1): 36-43.
- Chuzho K and Dkhar MS 2019b. Ecological determinants of woodrotting fungal diversity and first report of *Favolaschia calocera*, an invasive species from India. *Proceedings of The National Academy of Sciences, India Section B – Biological Sciences* **89**(4): 1177-1188.
- Chuzho K 2021. Altitudinal variation in diversity of wood-rotting fungi from forests of Nagaland. Ph. D. Thesis. North-Eastern Hill University, Shillong, India.
- Foroutan A and Jafary N 2007. Diversity of heart and root rot fungi on parks and roadside trees in Maharashtra, India. *Journal of Applied Science and Environment Management* **11**(4): 55-58.
- Gilbertson RL and Ryvarden L 1987. North American Polypores. Fungiflora, Oslo.
- Goodell B 2020. Fungi involved in the biodeterioration and bioconversion of lignocellulose substrates. *The Mycota II* Chap. 15. doi.org/10.1007/978-3-030-49924-2\_15.
- Ho WH, Yanna, Hyde KD and Hodgkiss IJ 2002. Seasonality and sequential occurrence of fungi on woods submerged in Tai Po Kau forest stream, Hong Kong. *Fungal Diversity* **10**: 21-43.
- Hsieh T and Wu JM. 2001. Cell growth and gene modulatory activities of Yunzhi (Windsor Wunxi) from mushroom Trametes

versicolor in androgen-dependent and androgen-insensitive human prostate cancer cells. *International Journal of Oncology* **18**: 81-88.

- Kaarik A 1974. Decomposition of wood. Biology of Plant Litter Decomposition 1: 129-174.
- Leelavathy KM and Ganesh PN 2000. *Polypores of Kerela*. Daya Publishing House, Delhi, pp. 1-165.
- Liang C, Tsai S, Huang S, Liang Z. 2010. Taste quality and antioxidant properties of medicinal mushroom *Phellinus lenteus* and *Sparassis crispa* mycelia. *International Journal of Medicinal Mushrooms* **12**(2): 141-150.
- Lyngdoh A 2014. Diversity of wood-rotting macrofungi of East Khasi Hills and decay potential of some selected species. Ph. D. Thesis. North-Eastern Hill University, Shillong, India.
- Lyngdoh A and Dkhar MS 2018. Decay potential of four wood-rot fungi on *Batula alnoides* and *Quercus dealbata* wood-blocks. *Asian Journal of Microbiology, Biotechnology & Environmental Sciences* **20**(3): 935-942.
- Mueller GM, Schmit JP, Leacock P, Buyck B 2004. Global diversity and distribution of macrofungi. *Biodiversity and Conservation* **16**: 37-48.
- Nagadesi PK and Arya A 2014a. Timber degrading fungi in sawmills of Gujarat, India. *International Letters of Natural Sciences* **2**: 13-22.
- Nunez M and Ryvarden L 2000. *East Asian Polypores* Volume 1 (Ganodermataceae and Hymenochaetaceae). Synopsis fungorum 13. Fungiflora, Oslo, pp. 1-168.
- Nunez M and Ryvarden L 2001. East Asian Polypores Volume 2 (Polyporaceae). Synopsis fungorum 14. Fungiflora, Oslo, pp. 170-522.
- Olou BA, Yorou NS, Striegel M, Bassler C and Krah F 2019. Effects of microclimate and diversity of tropical wood-inhabiting fungi. *Forest Ecology and Management* **437**: 79-87.
- Pinar O, Rodriguez-Conto S 2024. Biologically active secondary metabolites from white rot fungi. *Frontiers in Chemistry* **12**: 1363354.
- Pouska V, Svoboda M and Lepsova A 2010. The diversity of wooddecaying fungi in relation to changing site condition in an old growth mountain spruce forest, Central Europe. *European Journal of Forest Research* **129**(2): 219-231.
- Prasher IB and Ashok D 2013. A checklist of wood rotting fungi (nongilled Agaricomycotina) of Himachal Pradesh. *Journal of New Biological Reports* **2**(2):71-98.
- Ranadive KR 2013. An overview of Aphyllophorales (wood-rotting fungi) from India. *International Journal of Current Microbiology and Applied Sciences* **2**(12): 112-139.
- Ranadive KR, Viadya JG, Jite PK, Ranade VD, Shosale SR, Rabba AS, Hakimi M, Deshpande GS, Rathod MM, Forutan A, Kaur S, Naik-Vaidya CD, Bapat GS, and Lamrood P 2011. Checklist of Aphyllophorales from the Western Ghats of Maharashtra state, India. *Mycosphere* **2**(2): 91-114.
- Riley R, Salamov AA, Brown D, Nagy LG, Floudas D, Held BW, Levasseur A, Lombard V, Morin E, Otilar R, Lindquist E, Sun H, LaButti KM, Schmutz J, Jabbour D, Luo H, Baker SE, Pisabarro AG, Walton JD, Blanchette RA, Henrissat B, Martin F, Cullen D, Hibbett DS and Grigoriev IV 2014. Extensive sampling of basidiomycete genomes demonstrates inadequacy of the whitebrown-rot paradigm of wood decay fungi. *Proceedings of The National Academy of Sciences*, USA **111**(27): 9923-9928.
- Ryvarden L and Johansen I 1980. A Preliminary Polypore Flora of EastAfrica. *Fungiflora*, Oslo.
- Sanchez C 2008. Lignocellulosic residues: Biodegradation and bioconversion by fungi. *Biotechnology Advances* 27(2): 185-194.
- Sharma JR 2007. Wood rotting fungi of temperate Himalaya. In: Mukerji KG and Manoharachary C. (eds), Current concepts in Botany, IK International Publishing House Pvt. Ltd., New Delhi, pp. 101-120.

- Silva D. 2003. *Ganoderma lucidum* (Reishi) in cancer treatment. *Integrative Cancer Therapies* **2:** 358-364.
- Singh AP, Kim YS and Singh T 2016. Bacterial degradation of wood. In: Secondary Xylem Biology. doi.org/10.1016/B978-0-12-802185-9.00009-7.
- Swapna S, Sye A and Krishnappa M 2008. Diversity of macrofungi in semi-evergreen and moist deciduous forest of Shimoga District-Karnataka, India. *Journal of Mycology and Plant Pathology* 38(1): 21-26.
- Tapwal A, Kumar R and Pandey S 2013. Diversity and frequency of macrofungi associated wet evergreen tropical forest in Assam, India. *Biodiversitas* **14**(2): 73-78

Received 01 July, 2024; Accepted 21 September, 2024

- Vabeikhokhie JMC, Zohmangaiha, Zothanzama J and Lalrinawmi 2019a. Taxonomic study of wood inhabiting fungi of Reiek Reserved Forest, Mizoram, India. *Journal of Emerging Technologies and Innovative Research* **6**(4): 698-702.
- Vabeikhokhie JMC, Zohmangaiha, Zothanzama J and Lalrinawmi 2019b. Diversity of wood rotting fungi from two different forests in Mizoram, India. *International Journal of Current Microbiology* and Applied Sciences **8**(4): 2775-2785.
- www.fungifromindia.com/fungifromindia/buildPage.php?page=data bases

www.indexfungorum.org www.mycobank.org



Indian Journal of Ecology (2024) 51(5): 977-984 DOI: https://doi.org/10.55362/IJE/2024/4339 Manuscript Number: 4339 NAAS Rating: 5.38

# Genetic Analysis of Morpho-Physiological Traits in Relation to Heat Tolerance in Barley (*Hordeum vulgare* L.)

Ashok, Yogender Kumar\* and Amit

Wheat and Barley Section, Department of Genetics and Plant Breeding, CCS Haryana Agricultural University, Hisar-125 004, India \*E-mail: yogenderkgulia@gmail.com

Abstract: The subject of the current study was to assess heat susceptibility index with the aim to identify the heat tolerant genotypes, in addition to recognize the selection criterion for heat tolerance. Forty four barley genotypes were tested in two environments as created by different sowing dates *i.e.* timely sown (non-stress) and late sown (heat stress) during 2021-22 at CCS Haryana Agricultural University, Hisar. The genotypes namely BH 1018, IBON-HI-2021-52, BH 1036, DWRUB 64, 7<sup>th</sup> GSBON-2020-101, IBON-HI-2021-102, IBON-HI-2021-85 and DWRB 123 were identified heat tolerant based on heat susceptibility index of grain yield. Further, results based on average of ranks of HSI of all traits illustrated the genotypes, BH1018, IBON-HI-2021-102, IBON-HI-2021-52, DWRB 209 and IBON-HI-2020-55 with high heat tolerance under stress condition. Correlation among HSI of different traits indicated significant positive association of grain yield per plant with days to heading and maturity, spike length, number of grains per spike, biological yield per plant, harvest index and NDVI 2, exhibiting the relevance of these traits for deciding the selection criterion for stress condition. The genotypes of cluster III portrayed better performance under stress condition for grain yield and other traits studied. The genotypes from this group could be utilized as promising breeding material intended to develop new heat tolerant barley varieties.

#### Keywords: Barley, Heat stress, HSI, Tolerance

Barley (Hordeum vulgare L.) is a true diploid (2n=14) species that belongs to the genus Hordeum of family Poaceae and tribe Triticeae. Since ancient times, barley has been used in diverse ways as human food, livestock feed and in malt production (Kumar et al 2020) and is a hardy and versatile crop that allows its cultivated in a variety of agroclimatic zones. It is one of the globally accepted cereal crops due to its low input requirements and cost effective cultivation. Abiotic stresses remain one of the major yield reducing factors of various crop plants. Among them, high temperature, drought and salinity etc. are some of the factors causing undesirable effects on commercial cultivation of crops (Kumar et al 2022). Heat stress is emerging as a great threat influencing growth and development of most crop plants due to climatic disturbances as a result of global warming. The influence of high temperature on crops depends on the intensity and duration of heat, rate of temperature rise, and developmental stage of plants (Wahid et al2007). Increase in temperature particularly at post anthesis stages shorten the grain filling period and also reduces individual grain weight (Dias and Lidon 2009, Kaur and Behl 2010). Furthermore, crop yield can be affected by high temperatures by inducing pollen sterility as well as causes seed abortion during the reproductive growth stage (Barnabas et al 2008). There is challenge for crop researchers and need of hours to develop heat tolerant high yielding barley cultivars with good malt content for heat stress conditions. The most favorable temperature in barley for grain filling is 20°C in sub-tropical regions (Dwivedi et al 2017). It was,however, also reported that during grain filling even an increase of 1° C temperature from the optimum range causes adverse effects on grain yield (Narayanan 2018).

Under heat stress among the physiological processes, photosynthesis is the most sensitive mechanism (Wang et al 2015). The decline in the performance of genotypes under stress for various attributes could be due to inhibition of photosynthesis, as reflected by the loss of leaf chlorophyll content. The physiological responses of tolerance to abiotic stresses need to be studied in details at the genetic level before they can be exploited in the introgressive breeding. Reduction in crop productivity as a consequence of heat stress during the reproductive stage has also been reported in barley (Klink et al 2014). Vaezi et al (2010) observed grain yield loss by 39.59 and 31.39%, respectively in two-and sixrow barley genotypes owing to reduction in number of spikes per square meter, grain number per spike and 1000-grain weight due to delayed sowing. In addition, average grain yield reduction in barley genotypes by 17% was also stated by Modhej et al (2015), when the crop was exposed to heat stress after anthesis. The development of stable barley genotypes with higher economic yield under different environments is a need of breeding program intended to stress tolerance. The investigation of new sources of genetic variability and their utilization is an essential task that breeders would like to achieve for incorporation of heat tolerance in the varietal development process (Verma et al 2021). The crop production will be seriously challenged as a result of rise in temperature by 2.0-4.5 °C till the end of the century (Liu et al 2017), as reported by the Intergovernmental Panel on Climate Change (IPCC). Hence, effective exploitation of heat tolerant genetic resources in breeding programmes is imperative in context to genetic erosion and climate change scenarios (Bahrami et al 2019). Therefore, to cope up with the alarming threat of high temperature stress, detailed understanding of physiological responses and mechanisms of plants to heat tolerance and their further possible strategies for improving crop thermo tolerance is vital (Devi et al 2021). The heat susceptibility index may be used as a measure of heat tolerance in order to display yield stability under heat stress (Kavita et al 2016). Consequently, fourty four genotypes of barley including six and two-row types were evaluated for heat tolerance using HSI based on grain yield of stress and non-stress environments.

## MATERIAL AND METHODS

The field trial was conducted during 2021-22 crop season at CCS Haryana Agricultural University, Hisar which is situated in subtropical region of North Western Plain Zone of India at latitude of 29°10'N, longitude of 75°46'E and altitude of 215.2 m above sea level. The experimental material consisted of 44 barley genotypes representing both 2-row (23) and 6-row (21) types including four check varieties, DWRB 123 and BH 946 (Timely sown); and DWRB 91 and DWRUB 64 (Late sown genotypes) in randomized block design replicated thrice. Each genotype was planted in paired rows of 2.5 m length spaced at 23 cm apart. The experimental material was grown under timely (non-stress) and late sown (heat stress) conditions on 20th November and 20<sup>th</sup> December, 2021, respectively. The package of practices recommended for both environments in terms of inputs were accordingly applied to raise the crop.

Twelve morphological traits *viz.*, days to heading, days to maturity, grain filling duration, number of effective tillers per plant, plant height (cm), peduncle length (cm), spike length (cm), number of grains per spike, 1000-grain weight (g), biological yield per plant (g), grain yield per plant (g), and harvest index (%) alongwith physiological parameters namely canopy temperature depression at anthesis (CTD 1) and 15 days after anthesis (CTD 2), normalized difference

vegetation index at anthesis (NDVI 1) and 15 days after anthesis (NDVI 2), and SPAD chlorophyll content at anthesis (SPAD 1) and 15 days after anthesis (SPAD 2) were studied under both stress and non-stressed environments.

Heat susceptibility index (HSI) based on grain yield and all other traits were calculated using the formula as suggested by Fischer and Maurer (1978). HSI = [1-YD/YP]/DWhere, YP = Mean of genotypes under timely sown, YD = Mean of genotypes under stress and D = 1-[Mean YD of all genotypes/Mean YP of all genotypes].The recorded data was subjected to statistical analysis using Microsoft Excel for calculation of HSI. R studio version 2023.12.1.402 was used for correlation coefficients and cluster analysis.

The weather parameters for 2021-22 crop season were obtained from the Agricultural Meteorology Department, CCS HAU, Hisar and are presented in Figure 1. During crop season, highest values for maximum (41.1°C) and minimum temperature (21.2°C) were recorded in standard week 15<sup>th</sup> and 16<sup>th</sup> (2022), respectively. The data disclosed that the average minimum and maximum temperature for post heading phase of the crop under timely sown condition was 11.37 and 28.50°C, whereas, under late sown condition, it was 14.64 and 34.32°C, respectively (Fig. 2). An average increase of 3.26°C and 5.82°C were observed for minimum and maximum temperature under stress as compared to non stress conditionduring post heading, denoting the appearanceof significant heat stress under the late sown condition.

#### **RESULTS AND DISCUSSION**

The estimates of HSI for grain yield revealed that the genotypes, BH 1018 followed by IBON-HI-2021-52, BH 1036 and DWRUB 64 exhibited minimum values of HSI, revealing the genotypes with low heat susceptibility and high yield stability under heat stress condition (Table 1). In contrast, IBON-HI-2020-155 followed by 7th GSBYT-2020-20 and BH 1029 recorded maximum HSI for grain yield and were identified as highly heat susceptible genotypes. HSI of various traits in barley were also used by Ram and Shekhawat (2017) for selection and utilization of heat tolerant genotypes in future breeding programme. For biological yield per plant, minimum HSI was shown by 7th GSBON-2020-101 followed by IBON-HI-2021-85, BH 1027 and BH 1026. Similarly, minimum HSI for harvest index was recorded for genotype BH 1018 followed by IBON-HI-2020-6, IBYT-HI-2021-17, IBON-HI-2021-81 and BH 1036. All of these genotypes showed their superiority for tolerance to high temperature than other genotypes for concerned traits. The genotypes confirmed with negative values of HSI for trait under study signifies the better performing genotype under

heat stress as compared to non-stress condition, is suitable for climate resilience (Thakur et al 2020).

Three genotypes namely IBON-HI-2021-52, BH 1034 and BH 946 showed their superiority for days to heading and maturity by exhibiting low estimates of HSI for these traits. Four genotypes were promising for grain filling duration viz., IBON-HI-2021-27, IBYT-HI-2021-3, DWRB 209 and 7th GSBON-2020-101. The HSI estimates for number of effective tillers per plant were minimum for BH 1038 followed by BH 946, DWRUB 64 and BH 1018. The heat tolerant genotypes with respect to plant height were IBON-HI-2021-81 followed by IBYT-HI-2020-6, BH 1038, IBYT-HI-2021-3 whereas, for peduncle length, IBYT-HI-2021-3 followed by BH 1018, IBYT-HI-2021-15 and BH 1027 showed tolerance to heat under stress condition. Spike length and number of grain per spike, important yield attributes for which DWRB 209, IBON-HI-2021-38 and IBON-HI-2020-55 showed low estimates of HSI. The genotype i.e. BH 1035 followed by BH 1026, IBYT-HI-2021-15 and IBON-HI-2021-33 exhibited minimum reduction in 1000-grain weight under heat stress among all evaluated genotypes. Bahrami et al (2020) selected heat tolerant genotypes employing selection indices based on grain yield in cultivated (Hordeum vulgare ssp. vulgare L.) and wild (H. vulgaressp. spontaneum L.) barley genotypes. All the physiological traits considered together and it was found that five genotypes *i.e.* BH 1025, IBON-HI-2021-102, DWRB 209, IBON-HI-2020-155 and DWRB 91 had low estimates of HSI simultaneously for four physiological characters, implies physiologically efficient entries under stress condition in context to heat tolerance.Various morpho-physiological traits were also used for heat tolerance screening by Sallam et al (2018) in barley.

All the genotypes including check varieties were first ranked on the basis of HSI of concerned studied trait and then based on average of ranks of HSI of all traits, the genotypes were further ranked, indicated as overall rank (Table 1). The genotypes *viz.*, BH 1018, IBON-HI-2021-102, IBON-HI-2021-52, DWRB 209 and IBON-HI-2020-55 were the most tolerant to terminal heat stress among the experimental material evaluated. Due to detrimental effect of high temperature during reproductive phase, the genotypes performed differently under stress. Few genotypes could combat with the stress, while some of them were adversely affected (Fig. 3). The promising genotypes showing minimum reduction in grain yield were BH 1018 (2%) followed by IBON-HI-2021-52, BH 1036 and DWRUB 64. The reduction in grain yield, spike length, grains per spike and 1000-grain weight in



Fig. 1. Mean meteorological data during crop season 2021-2022



Fig. 2. Post heading maximum and minimum temperature under normal and late sown condition during 2021-22

Table 1. Heat susce	ptibility	index (ŀ	HSI) of I	oarley g	enotyp	es for c	different	traits												
Genotypes	Н	DM	GFD	ЕТР	ΗН	ΡL	SL	GPS	TGW	ВΥ	Н	GΥΡ	SPAD1	SPAD2	NDVI1	NDVI2	CTD1	CTD2	Я	SR
BH 1029	0.88	0.82	0.96	0.99	0.73	1.47	1.41	0.81	0.26	1.79	-0.15	1.71	0.22	0.64	0.36	0.44	3.74	3.49	42	44
BH 1034	0.53	0.69	1.22	0.66	0.80	1.67	06.0	0.24	0.10	0.84	-0.17	0.71	0.32	0.53	0.40	0.24	3.60	3.14	14	15
BH 1035	0.70	0.79	1.38	0.43	0.80	1.50	0.70	0.12	-0.18	1.17	-0.49	0.82	-0.34	0.35	0.05	0.48	3.72	3.44	20	20
BH 1036	0.63	0.72	1.12	0.40	1.02	2.17	0.67	0.58	0.14	1.15	-1.01	0.42	-0.32	0.07	0.27	0.25	3.72	3.48	Э	10
BH 1038	0.68	0.73	1.14	-0.62	0.28	1.92	0.25	0.96	0.30	1.23	0.28	1.42	-0.11	0.41	0.16	0.34	3.64	3.29	34	13
BH 1039	0.74	0.76	1.05	0.54	0.63	1.93	-0.04	0.76	0.11	1.42	-0.66	0.98	-0.20	0.28	0.41	0.78	3.68	3.33	25	27
BH 1018	0.67	0.69	0.93	0.18	0.52	0.71	0.46	0.61	0.75	1.20	-1.57	0.09	-0.12	0.29	0.31	0.12	3.67	3.38	<del>.</del>	<del></del>
BH 1025	0.74	0.77	1.01	0.64	0.86	0.98	0.61	0.21	0.04	1.49	-0.87	0.94	0.21	0.06	0.31	0.19	3.48	2.63	23	9
BH 1026	0.74	0.74	0.94	1.01	0.63	1.46	0.95	0.39	-0.04	0.70	0.88	1.43	-0.50	0.59	0.20	0.61	3.56	2.89	35	16
BH 1027	0.77	0.77	1.06	0.78	0.36	0.83	0.80	0.50	0.05	0.65	0.08	0.72	0.62	0.16	0.21	0.44	3.65	3.22	16	14
IBYT -HI-2021-3	0.83	0.72	0.61	0.78	0.32	0.22	0.34	0.53	0.22	1.05	0.05	1.09	0.33	0.38	0.15	0.38	3.58	2.99	29	7
IBYT -HI-2021-9	0.62	0.78	1.55	0.66	0.95	1.59	0.43	0.17	0.50	1.70	-0.73	1.27	0.07	1.05	0.41	0.13	3.72	3.48	32	36
IBYT -HI-2021-15	0.82	0.84	1.03	0.34	0.83	0.78	0.46	0.84	0.00	1.67	-0.70	1.25	0.22	0.27	0.05	0.67	3.60	3.05	30	18
IBYT -HI-2021-17	0.85	0.87	1.21	0.90	0.95	1.38	0.97	0.86	0.96	2.08	-1.06	1.54	-0.02	0.19	0.05	0.58	3.68	3.34	38	40
IBYT -HI-2021-18	0.87	0.84	1.06	0.49	0.76	1.27	0.18	0.84	0.27	1.48	-0.72	1.02	0.09	0.34	0.27	0.50	3.73	3.52	27	35
IBON-HI-2021-27	0.87	0.75	0.53	0.23	0.55	1.47	0.79	0.30	0.54	1.94	-0.65	1.59	0.08	0.75	0.37	0.21	3.77	3.52	39	34
IBON-HI-2021-31	0.82	0.81	1.01	0.70	0.55	2.19	0.93	0.52	0.21	1.22	0.56	1.62	0.18	0.18	0.31	0.95	3.74	3.41	40	42
IBON-HI-2021-33	0.81	0.80	0.94	0.41	0.62	1.59	0.84	0.38	0.04	1.20	-0.69	0.71	0.11	0.04	0.27	0.57	3.73	3.48	14	21
IBON-HI-2021-38	0.70	0.78	1.22	0.43	0.69	1.19	0.17	0.24	1.19	1.17	-0.21	1.02	0.17	0.14	0.26	0.55	3.79	3.60	27	30
IBON-HI-2021-52	0.53	0.66	1.18	0.35	0.63	1.18	0.48	0.34	0.06	0.85	-0.79	0.23	0.06	0.17	0.26	0.47	3.75	3.51	2	e
IBON-HI-2021-56	0.74	0.81	1.31	0.68	0.64	1.09	0.94	0.48	1.19	1.36	-0.15	1.25	0.32	0.23	0.31	0.49	3.79	3.62	30	43
IBON-HI-2021-81	0.75	0.85	1.30	0.89	0.14	1.33	0.03	0.58	0.82	1.32	-1.02	0.63	0.33	0.53	0.37	0.14	3.63	3.16	12	25
IBON-HI-2021-85	0.51	0.71	1.40	0.86	0.77	1.72	0.59	0.18	1.20	0.38	0.17	0.53	0.32	0.85	0.26	0.36	3.57	3.07	7	24
IBON-HI-2021-99	0.62	0.73	1.28	0.41	1.07	1.58	1.61	0.47	0.80	1.25	-0.36	1.00	0.49	0.04	0.38	0.06	3.69	3.41	26	33
IBON-HI-2021-102	0.75	0.78	0.99	0.64	0.08	0.94	0.38	06.0	0.47	0.76	-0.31	0.51	-0.20	0.19	0.16	0.23	3.59	3.16	9	7
RD 3002	0.68	0.78	1.27	0.67	0.53	1.91	0.76	0.27	0.67	1.12	-0.69	0.61	-0.19	0.28	0.36	0.32	3.68	3.33	11	19
DWRB 197	0.63	0.75	1.41	0.38	0.51	1.12	1.01	1.03	1.12	1.16	-0.49	0.81	0.31	0.62	0.05	0.66	3.69	3.37	19	31
DWRB 209	0.70	0.64	0.63	0.52	0.87	1.72	0.11	0.03	0.62	0.73	0.05	0.77	0.08	0.16	0.26	0.12	3.65	3.32	18	4
BCLA 11-6	0.69	0.78	1.34	0.39	0.64	1.64	0.45	1.50	0.36	1.47	0.00	1.47	0.11	0.22	0.20	0.13	3.70	3.40	37	32
IBON-HI-2020-71	0.72	0.82	1.32	0.52	1.08	1.89	1.25	0.19	0.34	1.38	-0.78	0.86	0.59	0.15	0.32	0.68	3.76	3.53	21	41
7 <sup>th</sup> GSBON-2020-90	0.66	0.75	1.22	0.21	1.03	1.47	0.84	1.18	1.23	1.46	-0.23	1.30	0.39	0.73	0.37	0.26	3.66	3.31	33	39
IBON-HI-2020-6	0.59	0.72	1.34	0.59	0.63	1.97	0.54	0.29	1.34	1.35	-1.13	0.58	0.51	0.62	0.27	0.39	3.69	3.33	0	28
IBON-HI-2020-51	0.67	0.65	0.83	0.59	0.59	2.09	0.62	0.30	0.43	1.05	-0.61	0.60	0.58	0.67	0.21	0.18	3.65	3.30	10	7
7 <sup>th</sup> GSBON-2020-140	0.68	0.65	0.74	0.81	0.39	1.59	0.31	1.23	0.23	1.60	0.03	1.62	0.20	0.65	0.00	0.57	3.64	3.24	40	21
IBON-HI-2020-55	0.67	0.65	0.74	0.61	0.67	1.61	0.37	0.16	0.29	1.12	-0.61	0.67	0.07	0.67	0.21	0.06	3.70	3.40	13	5
7 <sup>th</sup> GSBON-2020-101	0.72	0.67	0.68	0.33	0.41	2.19	0.57	0.65	0.80	-0.02	0.48	0.47	0.23	0.36	0.21	0.06	3.66	3.38	5	12
IBYT-HI-2020-6	0.73	0.72	0.92	0.24	0.28	0.86	0.36	0.91	0.33	0.89	0.05	0.93	0.21	0.17	0.36	0.42	3.67	3.32	22	6
7 <sup>th</sup> GSBYT-2020-20	0.65	0.78	1.33	1.02	0.57	1.53	0.45	0.56	1.32	1.69	0.05	1.72	-0.07	0.39	0.05	0.70	3.69	3.33	43	37
IBYT-HI-2020-11	0.61	0.76	1.37	0.40	0.77	2.22	0.14	0.73	0.66	0.97	-0.27	0.76	0.17	0.26	-0.05	0.55	3.61	3.21	17	17
IBON-HI-2020-155	0.70	0.80	1.19	0.89	0.57	2.01	1.56	0.09	0.70	1.80	0.11	1.86	-0.08	0.59	0.10	0.64	3.64	3.20	44	38
DWRB 123 (c)	0.61	0.76	1.42	0.34	0.99	2.10	1.25	0.45	0.18	1.05	-0.64	0.56	0.19	0.23	-0.27	0.54	3.69	3.41	8	23
BH 946 (c)	0.56	0.67	1.27	-0.08	0.78	2.02	0.71	0.68	0.58	1.40	-0.63	0.97	0.00	0.65	0.16	0.22	3.71	3.48	24	26
DWRB 91 (c)	0.72	0.76	1.05	0.45	0.62	1.66	1.36	0.67	0.70	1.83	-0.72	1.43	-0.12	0.20	0.10	0.12	3.71	3.44	35	29
DWRUB 64 (c)	0.52	0.74	1.35	0.04	0.39	1.07	0.68	0.23	0.50	0.85	-0.52	0.43	0.06	0.34	0.26	0.30	3.76	3.57	4	8
DH: Days to heading, DM: grain weight, BY: Biologic Normalized difference v 3. Canonytemperaturedent	Days to m al yield p∉ egetation	eturity, GI Plant, H index a	FD: Grain II: Harves: it anthes	filling dura t index, G is, NDVI	YP: Grain 2: Nor based of	<ul> <li>Numbe n yield pe malized</li> </ul>	er of effecti er plant, S difference	ve tillers p PAD 1: Cl vegetat	er plant, P Ilorophyll ion index based on	H: Plant h content (5 < at 15	eight, <b>PL</b> : SPAD) at days af	Peduncl anthesis, ter anthe	e length, S SPAD 2: esis, CTD	L: Spike le Chlorophy 1: Car	ength, <b>GP</b> Il content opy tem	S: Numbe (SPAD) ( perature	er of grair at 15day depress	s per spik s after an ion at a	e, <b>TGW</b> : hesis, NI anthesis,	1000- 11: CTD
<ul> <li>Callupylelliperature user</li> </ul>	בססוחומו	100dys an		o, <b>N</b> . Malin	המספת כ	יו או מווו	) ' <b>()</b> 'n	עבו מוו ו מו ייי	המסמת כיו	dvelayer										

barley also cited by Pathak et al (2017) under stress condition.

Grain yield per plant exhibited significant positive association with days to heading and maturity, spike length, number of grains per spike, biological yield per plant, harvest index and NDVI 2, exhibiting the relevance of these traits for deciding the selection criterion for stress conditions (Table 2). HSI based association study for heat tolerance was also conducted by Shehrawat et al (2020). Significant positive correlations were also observed for days to heading with days to heading, number of effective tillers per plant, and biological yield; days to maturity with grain filling duration, number of effective tillers per plant, biological yield and NDVI 2; grain filling duration with plant height and 1000-grain weight; plant height with peduncle length, spike length and biological yield; biological yield with CTD 2; CTD 1 with CTD 2. The positive association enhances the progress under selection and viceversa. Similarly, significant negative correlation was existed for days to heading with grain filling duration and peduncle length; number of effective tillers per plant with CTD 2; plant height with harvest index; biological yield with harvest index; and NDVI 1 with NDVI 2. These results corroborates for one or more traits with the findings of Devi et al (2021).

The clustering pattern recognized cluster I as largest one with 13 genotypes, followed by cluster V (9) and cluster III (8), while the cluster II and IV being smallest contained seven genotypes each (Table 3). The estimates of various stress indices were also used by Lamba et al (2023) for clustering of genotypes evaluated under stress condition. Several other genetic studies for classification of genotypes into different tolerance categories have also been conducted in barley based on HSI (Parashar et al 2019, Yadav et al 2023). The maximum intra-cluster distance for cluster I followed by cluster II and cluster III, implies the genotypes with relatively more diversity compared to genotypes belonging to other clusters. Similarly, the genotypes of cluster V showed more similarity as deciphered by minimum intra-cluster distance. The results in addition also revealed that cluster II placed most distantly from cluster III as exhibited by maximum intercluster distance among all cluster combinations, followed by clusters I and III. However, cluster III is most closely placed to cluster V as observed based on minimum distance among inter cluster distances.

The low estimates of HSI indicates the tolerance of genotypes to terminal heat. Cluster II exhibited superiority for days to maturity, grain filling duration, number of grains



Fig. 3. Per cent reduction in grain yield in barley genotypes

able 2. Co	orrelatio	n among	HSI of di	ifferent tr	aits in b	arley ger	otypes											
raits	Н	DM	GFD	ЕТР	Ηd	ΡL	SL	GPS	TGW	ВΥ	H	GYР	SPAD 1	SPAD 2	NDVI 1	NDVI 2	CTD 1	CTD 2
Н	1.000	0.564**	-0.550**	0.297*	-0.228	-0.358*	0.003	0.236	-0.278	0.305*	0.057	0.436**	0.003	-0.229	0.074	0.230	0.008	-0.038
Mo		1.000	0.307**	0.310*	0.109	-0.199	0.213	0.104	-0.020	0.497**	-0.174	0.407**	-0.033	-0.248	0.043	0.466**	0.242	0.173
SFD			1.000	-0.032	0.340*	0.163	0.183	-0.119	0.325*	0.069	-0.165	-0.094	0.067	0.058	-0.071	0.187	0.206	0.195
Ш				1.000	-0.058	-0.039	0.097	-0.226	0.009	0.106	0.200	0.317	0.080	0.126	0.002	0.268	-0.258	-0.340*
H					1.000	0.252*	0.272*	-0.262	-0.054	0.223*	-0.299*	0.063	0.078	-0.144	0.129	000.0	0.186	0.227
بر ا						1.000	0.100	-0.158	0.086	-0.028	0.106	-0.042	-0.086	0.139	-0.055	-0.023	0.053	0.069
۶Ľ							1.000	-0.149	0.010	0.162	0.011	0.173*	0.125	-0.052	0.019	0.131	0.241	0.227
SPS								1.000	0.058	0.225	0.070	0.256*	000.0	-0.002	-0.213	0.136	-0.137	-0.116
MD-									1.000	0.159	-0.007	0.028	0.180	0.242	0.048	-0.199	0.085	0.109
۶۲										1.000	-0.352**	0.702**	-0.099	0.147	0.066	0.162	0.289*	0.237
=											1.000	0.338**	0.106	0.145	-0.223	0.147	-0.272	-0.296
ЗΥР												1.000	-0.065	0.167	-0.099	0.348*	0.080	-0.005
SPAD 1													1.000	0.053	0.214	-0.042	-0.109	-0.137
SPAD 2														1.000	-0.031	-0.120	-0.167	-0.179
JDVI 1															1.000	-0.283*	0.239	0.222
																1.000	0.097	-0.007
3TD 1																	1.000	0.973**
3TD 2																		1.000
ee Table 1 fo	nr details																	



<sup>(</sup>For detail of genotypes with Sr. No., refer to Table 1)

Fig. 4. Dendrogram portraying clustering pattern of 44 barley genotypes

per spike, biological yield as well as for NDVI 2 (Table 4). The genotypes of cluster III were promising for days to heading, number of effective tillers per plant, harvest index, grain yield per plant, SPAD1 and 2, and NDVI 1. Similarly, cluster IV was positively associated with the traits *viz.*, plant height, peduncle length, spike length, 1000-grain weight, and CTD.Accordingly, the promising genotypes may be sorted from different clusters for their utilization in breeding program intended to develop heat tolerant cultivars. Suresh et al (2018) observations substantiated results for deciding criterion of selection and /or identification of genotypes is presented in Figure 4 in the form of circular dendrogram displaying the serial number of genotypes that may be decoded by conferring Table 1.

### CONCLUSION

HSI used was recognized as vital for identifying cultivars with high tolerance to heat stress. The genotypes, BH 1018, IBON-HI-2021-52, BH 1036, DWRUB 64, 7<sup>th</sup> GSBON-2020-101, IBON-HI-2021-102, IBON-HI-2021-85 and DWRB 123 were promising based on HSI of grain yield. However, results based on average of ranks of HSI of all traits illustrated the genotypes, BH 1018, IBON-HI-2021-102, IBON-HI-2021-52, DWRB 209 and IBON-HI-2020-55 with high heat tolerance under stress condition.Further, association study signifies the importance of days to heading and maturity, spike length,

Cluster members	Number of	Clusters		CI	uster distan	ces	
	genotypes		Cluster I	Cluster II	Cluster III	Cluster IV	Cluster V
BH 1029, BH 1038, BH 1039, IBYT-HI-2021-17, IBYT- HI-2021-18, IBON-HI-2021-27, IBON-HI-2021-31, IBON-HI-2021-33, BCLA 11-6, 7 <sup>th</sup> GSBON-2020-140, 7 <sup>th</sup> GSBYT-2020-20, IBON-HI-2020-155, DWRB 91	13	Cluster I	2.594	3.187	3.271	2.653	2.531
BH 1034, BH 1026, IBON-HI-2021-85, DWRB 209, IBON-HI-2020-51, IBON-HI-2020-55, 7 <sup>th</sup> GSBON-2020-101	7	Cluster II	3.187	2.346	3.312	2.896	3.028
BH 1035, BH 1036, BH 1018, IBON-HI-2021-52, RD 3002, IBYT-HI-2020-11, DWRB 123, DWRUB 64	8	Cluster III	3.271	3.312	2.311	2.692	2.331
BH 1025, BH 1027, IBYT-HI-2021-3, IBYT-HI-2021-15, IBON-HI-2021-81, IBON-HI-2021-102, IBYT-HI-2020-6	7	Cluster IV	2.653	2.896	2.692	1.955	2.667
IBYT-HI-2021-9, IBON-HI-2021-38, IBON-HI-2021-56, IBON-HI-2021-99, DWRB 197, IBON-HI-2020-71, 7 <sup>th</sup> GSBON-2020-90, IBON-HI-2020-6, BH 946	9	Cluster V	2.531	3.028	2.331	2.667	1.830

Table 3. Clustering of barley genotypes based on HSI of different traits and their genetic distances

Table 4. Performance of clusters for HSI of barley genotypes

	DH	DM	GFD	ETP	PH	PL	SL	GPS	TGW	BY	HI	GYP	SPAD 1	SPAD 2	NDVI 1	NDVI 2	CTD 1	CTD 2
СІ	0.77	0.78	1.04	0.55	0.60	1.67	0.73	0.73	0.46	1.60	-0.28	1.44	0.03	0.38	0.20	0.50	3.70	3.38
CII	0.65	0.68	0.92	0.65	0.68	1.78	0.59	0.28	0.49	0.69	0.03	0.74	0.16	0.55	0.25	0.23	3.63	3.21
C III	0.62	0.74	1.25	0.35	0.71	1.61	0.64	0.42	0.35	1.05	-0.75	0.49	-0.06	0.25	0.15	0.38	3.70	3.42
C IV	0.77	0.78	0.99	0.62	0.41	0.85	0.43	0.64	0.28	1.12	-0.39	0.87	0.25	0.25	0.23	0.35	3.60	3.08
CV	0.65	0.76	1.32	0.42	0.82	1.55	0.83	0.53	0.92	1.36	-0.52	1.01	0.32	0.47	0.28	0.38	3.72	3.46

See Table 1 for details

number of grains per spike, biological yield per plant, harvest index and NDVI 2 towards grain yield under stress condition. The genotypes of cluster III portrayed better performance under stress for grain yield and other traits studied, could be incorporated in elite barley cultivars intended to develop new heat tolerant varieties.

#### REFERENCES

- Bahrami F, Arzani A and Rahimmalek M 2019. Photosynthetic and yield performance of wild barley (*Hordeum vulgare* ssp. *spontaneum*) under terminal heat stress. *Photosynthetica* **57**: 9-17.
- Bahrami F, Arzani A and Rahimmalek M 2020. A novel tolerance index to identify heat tolerance in cultivated and wild barley genotypes. doi.org/10.1101/2020.05.31.125971.
- Barnabas B, Jager K and Feher A 2008. The effect of drought and heat stress on reproductive processes in cereals. *Plant, Cell and Environment* **31**: 11-38.
- Devi S, Kumar Y and Shehrawat S 2021. Identification of heat tolerant barley genotypes based on heat susceptibility index. *Journal of Cereal Research* **13**(2): 197-204.
- Dias AS and Lidon F C 2009. Evaluation of grain filling rate and duration in bread and durum wheat under heat stress after anthesis. *Journal of Agronomy and Crop Science* **195**(2): 137-147.
- Dwivedi SK, Basu S, Kumar S, Kumar G, Prakash V, Kumar S and Arora A 2017. Heat stress induced impairment of starch mobilisation regulates pollen viability and grain yield in wheat: Study in Eastern Indo-Gangetic Plains. *Field Crops Research* 206:106-114.

Fischer RA and Maurer R 1978. Drought resistance in spring wheat

cultivars. 1. Grain yield responses. Australian Journal of Agricultural Research **29**(5): 897-912.

- Kaur V and Behl RK 2010. Grain yield in wheat as affected by short periods of high temperature, drought and their interaction during pre-and post-anthesis stages. *Cereal Research Communications* **38**(4): 514-520.
- Kavita, Munjal R, Kumar N and Dhanda SS 2016. Stress response behavior in different wheat species in relation to heat tolerance. *Journal of Wheat Research* **8**(2): 49-53.
- Klink K, Wiersma JJ and Crawford CJ 2014. Impacts of temperature and precipitation variability in the Northern Plains of the United States and Canada on the productivity of spring barley and oat. *International Journal of Climatology* **34**: 2805-2818.
- Kumar A, Verma RPS, Singh A, Sharma HK and Devi G 2020. Barley landraces: Ecological heritage for edaphic stress adaptations and sustainable production. Environmental and Sustainability Indicators 6:10003.
- Kumar D, Lal C, Bishnoi SK, Verma RPS and Singh GP 2022. Biochemical and molecular basis of abiotic stress tolerance in barley. *Journal of Cereal Research* 14 (Spl-1): 83-95.
- Lamba K, Kumar M, Singh V, Chaudhary L, Sharma R, Yashveer S and Dalal MS 2023. Heat stress tolerance indices for identification of the heat tolerant wheat genotypes. *Scientific Reports* **13**(1):10842.
- Liu Q, Yan S, Yang T, Zhang S, Chen Y-Q and Liu B 2017. Small RNAs in regulating temperature stress response in plants. *Journal of Integrative Plant Biology* **59**: 774-791.
- Modhej A, Farhoudi R and Afrous A 2015. Effect of post-anthesis heat stress on grain yield of barley, durum and bread wheat genotypes. In: *Proceedings of the 10<sup>th</sup> International Barley Genetics Symposium*. Alexandria 230-235.
- Narayanan S 2018. Effects of high temperature stress and traits associated with tolerance in wheat. Open Access Journal of Science 2(3): 177-186.

- Parashar N, Gothwal DK and Singh G 2019. Study of heat susceptibility indices for yield and its attributes in barley (Hordeum vulgare L.). Journal of Pharmacognosy and Phytochemistry 8(2):1115-1119.
- Pathak S, Poudyal C, Ojha BR and Marahatta S 2017. Evaluation of the effects of terminal heat stress on grain traits of barley (Hordeum vulgare L.) in Chitwan, Nepal. International Journal of Agriculture and Environmental Research 3(2): 2856-2869.
- Ram M and Shekhawat AS 2017. Genotypic analysis for heat susceptibility index in two environments in barley for grain yield and its associate traits. *Plant Archives* 17(2):1305-1310.
- Sallam A, Amro A, El-Akhdar A, Dawood MFA, Kumamaru T and Baenziger PS 2018. Genetic diversity and genetic variation in morpho-physiological traits to improve heat tolerance in spring barley. *Molecular Biology Reports* **45**: 2441-2453.
- Shehrawat S, Kumar Y and Singh J 2020. Use of multiple stress indices as a measure of heat tolerance in wheat accessions. *Journal of Cereal Research* **12**(3): 297-308.
- Suresh, Bishnoi OP and Behl RK 2018. Use of heat susceptibility index and heat response index as a measure of heat tolerance in wheat and triticale. *Ekin Journal of Crop Breeding and Genetics* **4**(2): 39-44.

Thakur P, Prasad LC, Prasad R, Chandra K and Rashmi K 2020.

Received 28 June, 2024; Accepted 25 September, 2024

Estimation of genetic variability, heat susceptibility index and tolerance efficiency of wheat (*Triticum aestivum* L.) for timely and late sown environments. *Electronic Journal of Plant Breeding* **11**(3): 769-775.

- Vaezi B, Bavei V, Shiran B and Rahmani-Moghadam N 2010. Different contributions of yield components to grain yield in two-and six-row barley genotypes under terminal heat stress. *International Journal* of Applied Agricultural Research **5**(3): 385-400.
- Verma S, Yashveer S, Rehman S, Gyawali S, Kumar Y, Chao S, Sarker A and Verma RPS 2021. Genetic and agro-morphological diversity in global barley (*Hordeum vulgare* L.) collection at ICARDA. *Genetic Resources and Crop Evolution* 68(1): 1315-1330.
- Wahid A, Gelani S and Ashraf M 2007. Heat Tolerance in Plants: An overview. Environmental and Experimental Botany 61: 199-223.
- Wang X, Dinler BS, Vignjevic M, Jacobsen S and Wollenweber B 2015. Physiological and proteome studies of responses to heat stress during grain filling in contrasting wheat cultivars. *Plant Science* 230: 33-50.
- Yadav GL, Gothwal DK and Gupta D 2023. Identification of heat tolerant barley (*Hordeum vulgare* L.) genotypes based on heat susceptibility index. *The Pharma Innovation Journal* **12**(10): 548-552.



# Grain Quality, Soil Fertility Status and Nutrient Uptake Pattern of Emmer Wheat (*Triticum dicoccum* L.) under System of Wheat Intensification

Nagesh Rathod, Kumar D. Lamani, Milid Potdar and Uday G. Reddy<sup>1</sup>

Department of Agronomy, <sup>1</sup>Department of Genetics and Plant Breeding, UAS, Dharwad-580 005, India E-mail: nagesh551999@gmail.com

**Abstract:** Maintaining sustainable soil fertility level and enhancing emmer wheat production on smallholder farms is a great challenge in the Northern Dry Zone of Karnataka. Therefore, the present investigation on the grain quality, soil fertility status and nutrient uptake pattern of emmer wheat (*Triticum dicoccum* L.) under System of Wheat Intensification (SWI) in irrigated ecosystem conditions. Field experiment was conducted during *rabi* season of 2022-23 at Research field of Ugar Khurd, Belagavi, Karnataka. The experiment was laid out in a split plot design with four main plots *viz.*, Gokak local and DDK 1029 with and without seed priming and four planting geometries *viz.*, 30 × 15 cm; 45 × 15 cm; 20 × 20 cm and 20 cm (RPP) in sub plots under three replications. The primed seeds sown at 30 × 15 cm planting geometry recoded significantly higher number of effective tillers (463.83 m<sup>-2</sup>), grain yield (44.13 q ha<sup>-1</sup>), straw yield (70.64 q ha<sup>-1</sup>), biological yield (114.77 q ha<sup>-1</sup>), grain protein content (12.54 %), nitrogen, phosphorous and potassium content (2.65, 0.54 and 1.78 %) and total nitrogen, phosphorous and potassium uptake (133.63, 28.38 and 112.44 kg ha<sup>-1</sup>) as compared to un-primed seeds with other planting geometries.

Keywords: Emmer wheat, Nutrient contents and uptakes, Planting geometries, Seed priming, System of wheat intensification

Wheat is grown across a wide range of environmental conditions around the world. Cultivation of wheat dates back to more than 5000 years, during the period of Indus valley civilization during which the original species Triticum sphaerococcum, traditionally known as Indian wheat has vanished and replaced by modern day cultivars like Triticum aestivum (Bread wheat), Triticum durum (Macaroni wheat or Kathia wheat) and Triticum dicoccum (Emmer wheat or Khapli). Among different species of wheat, Triticum aestivum has 95 per cent production, Triticum durum has 4 per cent and Triticum dicoccum has one per cent production in India. Nearly, 82 to 85 per cent of the wheat grown in India is under irrigated situation, while the rest is grown under rainfed condition. The emmer wheat (Triticum dicoccum L.) is grown on a very restricted scale in Gujarat, Maharashtra and Karnataka. In general emmer wheat varieties are rich source of protein and complex carbohydrate (dietary fibre) as compared to bread wheat and it keeps one full for longer and hence helps in weight loss. The traditional products of emmer wheat varieties have better taste, texture and flavour (Singh, 2015).In India, wheat cultivation covers an extensive 30.47 million hectares and contributes significantly to food grain production, accounting for 36 per cent of the total output (Anonymous 2022). Lower wheat productivity is primarily due to delayed sowing, a short winter season, improper input management and terminal heat stress, particularly in the North Eastern Plain Zone of India, which leads to reduced tillering, inadequate crop establishment and shrivelled grains and yields. Traditionally, wheat cultivation has relied on the broadcasting and continuous sowing of seeds in rows without adhering to specific planting geometry. This approach has served well for extensive agricultural areas. However, it comes with limitations, such as the lack of uniformity in crop stand. This lack of uniformity results in the dilution effect of inputs and impacting on crop yield (Haque et al 2015). The System of Wheat Intensification (SWI) represents an innovative approach to wheat production on manipulating the soil environment with minimal external inputs and utilizing a very low seed rate. This technique aims to create optimal conditions for wheat cultivation, emphasizing the importance of maintaining an ideal plant population, SWI ensures adequate aeration, moisture, sunlight, and nutrient availability, which are crucial for fostering proper root system development during the early stages of crop growth. This holistic approach to wheat farming holds promise for improving yields and resource utilization while reducing external inputs.

The System of Wheat Intensification method was initially introduced in India, Africa, and Nepal by dedicated community workers on the fields of small and marginal farmers in the year 2006. These grassroots efforts yielded encouraging results, which played a pivotal role in inspiring

and guiding systematic research initiatives involving farmers, government agencies and research workers in India. SWI, a synergistic wheat farming technique, involves reduced seed density and wider spacing, coupled with seed treatment using organic formulations like cow dung, cow urine, jaggery and curd, to enhance plant health and productivity. Under the SWI the management practices create more favourable conditions for wheat crop growth. This is achieved through the greater proliferation of root hair and increased root length compared to traditional wheat farming methods, (Hague et al 2015). Despite encouraging results, there is limited information available regarding the overall effect of the SWI technique on wheat growth and productivity. Further research and data collection in this area could provide valuable insights into the full potential and benefits of adopting SWI in wheat farming. Therefore, the present experiment was conducted to investigate the "Grain quality, soil fertility status and nutrient uptake pattern of emmer wheat (Triticum dicoccum L.) under System of Wheat Intensification (SWI).

#### MATERIAL AND METHODS

**Experiment site:** The present investigation was conducted at Research field of Ugar Khurd, Belagavi, Karnataka, during *rabi* 2022-2023 located in the Northern Dry Zone (Zone-3) of Karnataka. Geographically it lies between 16° 38' 30" N latitude, 74° 49' 39" E longitude and at an altitude of 561 m above mean sea level (MSL).

**Soil status:** Experiments were laid out on well drained, deep black calcareous soil taxonomically belonging to vertisol with a depth of 2.5 to 3.5 m and slightly saline in nature with pH of pH ranging from 8.0 with medium electrical conductivity (0.39 dS  $m^{-1}$ ), low in organic carbon content (0.58 %), low in available nitrogen (279.1 kg  $ha^{-1}$ ), high in available phosphorous (32.6 kg  $ha^{-1}$ ) and potassium (371.2 kg  $ha^{-1}$ ).

**Treatment details:** The experiment was laid out in split plot design with four main plots *viz.*, gokak local and DDK 1029 with and without seed priming and four planting geometries in sub plots *viz.*,  $S_1$ - 30 × 15 cm;  $S_2$ - 45 × 15 cm;  $S_3$ - 20 × 20 cm and  $S_4$ - 20 cm (RPP). The seed rate varies with respective planting geometries and crop was sown during 1<sup>st</sup> week of November and harvest during 2<sup>nd</sup> week of March.

**Seed priming procedure:** Seed priming formulation comprising of water, cow dung, cow urine, jaggery and curd in the ratio of 2.0: 0.5: 0.5: 0.1: 0.05 was used to treat the seeds before sowing. In this case for priming 10 kg of emmer wheat seed, 10 litres of water, 2.5 kg of cow dung, 2.5 litre of cow urine, 500 g of jaggery and 250 g curd was taken in a bucket and was kept overnight (Sunaratiya and Banik 2022). The priming material was stirred during preparation period.

Thereafter, the material was sieved using cotton cloth and the resultant solution so obtained was used as priming formulation to treat the seeds. The seeds of wheat to be treated were first immersed in water contained in the tub to remove the chaffy seeds which were found floating on the surface of water. The seeds which settled in the bottom of the tub were collected and immersed in the priming solution for eight to ten hours. Took the seeds out of the priming solution and dried in shade. Thereafter were sown directly in the field.

**Quality parameters analysis:** i) Protein, nitrogen is determined by a method of chemical analysis known as the Kjeldahl procedure. Crude protein content was calculated by following formula: Crude protein of wheat = Per cent nitrogen × 5.7. ii) Grain N content was determined by Kjeldahl method described by (Kjeldahl, 1883) and expressed in percentage. iii) Grain P Content Total phosphorous on dry weight basis at harvest of wheat was estimated by Vanadomolybdo-phosphoric acid yellow colour method and expressed in percentage. iv) Grain K Content Available potassium in grain was extracted with neutral normal ammonium acetate and potassium content in the extract will be determined by using flame photometer (EelicoD-22) as outlined by (Jackson, 1973) and expressed in percentage.

**Nutrient uptake by wheat:** The uptake of nutrients by different parts of wheat was worked out by multiplying the nutrient content in seed and straw yield of the plant using the following formula:

Nutrient uptake	Nutrient concentration in grain (%) × Grain yield (kg ha <sup>-1</sup> )
(kg na ) – –	100
	+
	Nutrient concentration in Straw (%) × Straw yield (kg ha <sup>-1</sup> )
_	100

**Soil sample analysis:** Soil samples were collected before sowing and after harvest from individual plots of experiment by taking slice of soil from the depth of 0-30 cm. Soil samples were shade dried, powdered using wooden pestle and mortar and sieved through 2 mm sieve and chemically analysed to estimate soil reaction (pH), EC (dS m<sup>-1</sup>), organic carbon (OC %), available soil nitrogen (N), phosphorus ( $P_2O_5$ ) and potassium ( $K_2O$ ).

**Statistical analysis and interpretation of data**: The data were analysed in MS EXCEL (var. 2020) by using DMRT.

## **RESULTS AND DISCUSSION**

Yield and yield attributes of emmer wheat as influence by seed priming

Number of effective tillers (m<sup>-2</sup>): Seed priming significantly

influenced the number of effective tillers of emmer wheat. Primed seeds ( $M_4$ ) recorded significantly higher number of effective tillers (422.14 m<sup>-2</sup>) and was on par with  $M_2$  and  $M_3$ . However, significantly lowest number of effective tillers (401.86 m<sup>-2</sup>) was in un-primed seeds ( $M_1$ ) (Table 1). This can be due to fact that cow urine contains physiologically active substances *viz.*, growth regulators and nutrients that promote profusely more number of tiller per plant. Bhoopathi et al (2001) also observed significant increase number of tillers per plant due to treating of seed with cattle urine than untreated seed.

**Grain yield (q ha<sup>-1</sup>):** Grain yield differed significantly with respect to with and without seed priming. Primed seeds ( $M_4$ ) recorded significantly higher grain yield (38. 53 q ha<sup>-1</sup>) and was on par with  $M_2$  and  $M_3$ . The, significantly lower grain yield

(32.78 q ha<sup>-1</sup>) was in un-primed seeds) (Table 1). Seed priming also imparted a positive influence on grain and straw yield of wheat as compared to un-primed seed. The increase in grain yield of wheat under seed priming might be due to its positive effect on growth and yield parameters *viz.*, crop emergence, complete crop emergence, both above and belowground plant growth and yield attributes over unprimed seeds. Khadka and Raut (2011) and Sarlach et al (2013) have also reported that seed priming improves emergence, stand establishment, tillering, grain and straw yields.

**Straw yield (q ha**<sup>-1</sup>): Primed seeds ( $M_4$ ) recorded significantly higher straw yield (64.26 q ha<sup>-1</sup>). Whereas, significantly lower straw yield (58.88 q ha<sup>-1</sup>) was recorded with respect to un-primed seeds ( $M_1$ ) (Table 1). The higher

Table 1. Yield and yield attributes of emmer wheat as influenced by seed priming and different planting geometries

Ireatment	tillers m <sup>-2</sup>	(q ha⁻¹)	Straw yield (q ha⁻¹)	Biological yield (q ha <sup>-1</sup> )
Main plots:- (Two genotypes with	n and without seed priming )			
M₁-Gokak local (WOP)	401.86 <sup>⁵</sup>	32.78 <sup>b</sup>	58.88 <sup>b</sup>	91.65 <sup>⁵</sup>
M <sub>2</sub> - Gokak local (WP)	412.88 <sup>ab</sup>	36.15 <sup>ab</sup>	61.09 <sup>b</sup>	97.24 <sup>ab</sup>
M <sub>3</sub> -DDK 1029 (WOP)	409.51 <sup>ab</sup>	35.01 <sup>ab</sup>	61.42 <sup>b</sup>	96.43 <sup>b</sup>
M₄-DDK 1029 (WP)	422.14ª	38.53°	64.26 <sup>ª</sup>	102.79ª
Sub plots:- (Four planting geome	etries)			
S₁- 30 × 15 cm	452.85°	40.20ª	66.30ª	106.50°
S <sub>2</sub> - 45 × 15 cm	357.61⁴	34.08 <sup>b</sup>	60.06 <sup>bc</sup>	94.14 <sup>bc</sup>
S₃- 20 × 20 cm	433.58⁵	36.74 <sup>ab</sup>	61.78 <sup>b</sup>	98.51 <sup>ab</sup>
S₄- 20 cm (RPP)	402.34°	31.45°	57.52°	88.96°
Interaction (M × S)				
$M_1S_1$	439.73 <sup>bc</sup>	36.31 <sup>a-d</sup>	63.01 <sup>cd</sup>	99.32 <sup>cd</sup>
$M_1S_2$	347.77 <sup>°</sup>	31.52 <sup>cd</sup>	57.93 <sup>r</sup>	89.45 <sup>e-g</sup>
$M_1S_3$	424.83 <sup>cd</sup>	34.06 <sup>b-d</sup>	59.88 <sup>ef</sup>	93.94 <sup>d-f</sup>
$M_1S_4$	395.10°	29.21 <sup>d</sup>	54.68 <sup>°</sup>	83.89 <sup>ª</sup>
$M_2S_1$	460.17ª	42.35 <sup>ab</sup>	65.99 <sup>b</sup>	108.34 <sup>ab</sup>
$M_2S_2$	356.87 <sup>°</sup>	34.13 <sup>b-d</sup>	59.20 <sup>ef</sup>	93.33 <sup>d-g</sup>
$M_2S_3$	434.70 <sup>bc</sup>	37.02 <sup>a-d</sup>	61.66 <sup>de</sup>	98.67 <sup>с-е</sup>
$M_2S_4$	399.77°	31.11 <sup>cd</sup>	57.52 <sup>fg</sup>	88.64 <sup>fg</sup>
$M_3S_1$	447.68 <sup>ab</sup>	38.01 <sup>a-c</sup>	65.55 <sup>bc</sup>	103.56 <sup>bc</sup>
$M_3S_2$	350.77 <sup>g</sup>	35.00 <sup>b-d</sup>	60.16 <sup>ef</sup>	95.16 <sup>c-f</sup>
$M_3S_3$	434.87 <sup>bc</sup>	35.47 <sup>b-d</sup>	61.89 <sup>de</sup>	97.36 <sup>c-f</sup>
$M_3S_4$	404.73°	31.56 <sup>cd</sup>	58.07 <sup>f</sup>	89.62 <sup>e-g</sup>
$M_4S_1$	463.83°	44.13°	70.64ª	114.77ª
$M_4S_2$	375.03 <sup>r</sup>	35.68 <sup>b-d</sup>	62.94 <sup>cd</sup>	98.62 <sup>с-е</sup>
$M_4S_3$	439.93 <sup>bc</sup>	40.40 <sup>ab</sup>	63.68 <sup>b-d</sup>	104.08 <sup>bc</sup>
$M_4S_4$	409.77 <sup>de</sup>	33.90 <sup>b-d</sup>	59.80 <sup>ef</sup>	93.70 <sup>d-f</sup>

Mean followed by the same letter(s) did not differ significantly by DMRT (p= 0.05)

straw yield in primed seeds compared to un-primed seeds can be attributed to increased seedling vigour, stronger vegetative growth, greater dry matter production and improved growth parameters. This collective effect results in enhanced straw biomass, making seed priming with organic formulations a valuable method for boosting wheat straw yield. Similar results were also obtained by Khadka and Raut (2011) and Sarlach et al (2013).

**Biological yield (q ha**<sup>-1</sup>): Primed seeds ( $M_4$ ) recorded significantly higher biological yield (102.79 q ha<sup>-1</sup>) and it was recorded on par with  $M_2$  (97.24 q ha<sup>-1</sup>). The significantly lowest biological yield (91.65 q ha<sup>-1</sup>) was recorded with respect to un-primed seeds ( $M_1$ ) (Table 1). The increase in grain and straw yields in primed seeds as compared to unprimed seeds can be attributed to improved seedling vigour,

early growth and enhanced vegetative development. Seed priming proves effective in boosting overall crop performance, leading to higher yields in both grain and straw in wheat cultivation. Similar results were also obtained by Sharma (2020).

## Quality Parameters of Emmer Wheat as Influenced by Seed Priming

**Grain crude protein content of emmer wheat:** Significant differences in grain crude protein content were observed with respect to with and without seed priming methods. Primed seeds ( $M_4$  and  $M_2$ ) recorded significantly higher grain crude protein content (12.05 and 11.96 %, respectively). The significantly lowest grain crude protein content (11.08 %) was with respect to un-primed seeds ( $M_1$ ) (Table 2). These might be due to higher grain nitrogen content in primed seeds and

 Table 2. Grain crude protein and nitrogen content of emmer wheat as influenced by seed priming and different planting geometries

Treatment	Grain crude protein	Nitrogen content in (%)			
		Grain	Straw	Total	
Main plots:- (Two genotypes with a	and without seed priming )				
M <sub>1</sub> -Gokak local (WOP)	11.08°	1.62 <sup>b</sup>	0.48 <sup>b</sup>	2.09 <sup>b</sup>	
M2- Gokak local (WP)	11.96ª	1.78ª	0.51 <sup>ab</sup>	2.29ª	
M <sub>3</sub> -DDK 1029 (WOP)	<b>11.53</b> <sup>⁵</sup>	1.79ª	0.53 <sup>ab</sup>	2.32ª	
M <sub>4</sub> -DDK 1029 (WP)	12.05°	1.87ª	0.56ª	2.43ª	
Sub plots:- (Four planting geometr	ies)				
S₁- 30 × 15 cm	12.13ª	1.90 <sup>ª</sup>	0.60 <sup>ª</sup>	2.50 <sup>ª</sup>	
S <sub>2</sub> - 45 × 15 cm	11.71 <sup>b</sup>	1.75 <sup>ab</sup>	0.50 <sup>b</sup>	2.24 <sup>bc</sup>	
S <sub>3</sub> - 20 × 20 cm	11.89 <sup>ab</sup>	1.83ªb	0.56ª	2.39 <sup>ab</sup>	
S₄- 20 cm (RPP)	10.91°	1.59 <sup>b</sup>	0.41°	2.00°	
Interaction (M × S)					
$M_1S_1$	11.65 <sup>d-f</sup>	1.78 <sup>a-c</sup>	0.57 <sup>bc</sup>	2.35 <sup>b-e</sup>	
$M_1S_2$	11.05 <sup>gh</sup>	1.58 <sup>cd</sup>	0.42 <sup>ef</sup>	2.00 <sup>gh</sup>	
$M_1S_3$	11.26 <sup>tg</sup>	1.64 <sup>b-d</sup>	0.53 <sup>cd</sup>	2.17 <sup>d-g</sup>	
$M_1S_4$	10.37	1.47 <sup>d</sup>	0.38 <sup>r</sup>	1.85 <sup>h</sup>	
$M_2S_1$	12.32 <sup>ab</sup>	1.87 <sup>ab</sup>	0.58 <sup>bc</sup>	2.45 <sup>°-c</sup>	
$M_2S_2$	12.11 <sup>bc</sup>	1.77 <sup>a-c</sup>	0.51 <sup>d</sup>	2.28 <sup>b-f</sup>	
$M_2S_3$	12.17 <sup>a-c</sup>	1.84 <sup>a-c</sup>	0.54 <sup>cd</sup>	2.38 <sup>a-d</sup>	
$M_2S_4$	11.25 <sup>tg</sup>	1.64 <sup>b-d</sup>	0.40 <sup>ef</sup>	2.04 <sup>f-h</sup>	
$M_3S_1$	12.01 <sup>b-d</sup>	1.92ª	0.61 <sup>ab</sup>	2.53 <sup>a-c</sup>	
$M_3S_2$	11.51 <sup>ef</sup>	1.76 <sup>a-c</sup>	0.51 <sup>d</sup>	2.27 <sup>c-f</sup>	
$M_3S_3$	11.83 <sup>c-e</sup>	1.87 <sup>ab</sup>	0.58 <sup>bc</sup>	2.45 <sup>°-°</sup>	
$M_3S_4$	10.79 <sup>hi</sup>	1.61 <sup>b-d</sup>	0.41 <sup>ef</sup>	2.02 <sup>f-h</sup>	
$M_4S_1$	12.54°	2.01 <sup>ª</sup>	0.64 <sup>ª</sup>	2.65ª	
$M_4S_2$	12.16 <sup>a-c</sup>	1.87 <sup>ab</sup>	0.54 <sup>cd</sup>	2.41 <sup>a-d</sup>	
$M_4S_3$	12.28 <sup>ab</sup>	1.95ª	0.60 <sup>ab</sup>	2.55 <sup>ab</sup>	
$M_4S_4$	11.24 <sup>fg</sup>	1.64 <sup>b-d</sup>	0.45 <sup>e</sup>	2.09 <sup>e-h</sup>	

Mean followed by the same letter(s) did not differ significantly by DMRT (p= 0.05)

seed priming acts as a catalyst for several physiological and biochemical processes, creating a conducive environment for higher grain protein content and improved protein quality in emmer wheat. These results are in conformity with the finding of Muchhadiya et al (2021).

Nitrogen content in grain, straw and total content (%): Primed seeds (M<sub>4</sub>) recorded higher grain, straw and significantly higher total nitrogen content (1.87, 0.56 and 2.43 %, respectively) which was on par with M<sub>2</sub> and M<sub>2</sub>. The lower nitrogen content in grain, straw and significantly lower total nitrogen content (1.62, 0.48 and 2.09 %, respectively) was in un-primed seeds (M<sub>1</sub>) (Table 2). These might be due to seed priming with cow dung and cow urine contributes to higher grain nitrogen content in wheat through a combination of nutrient enrichment, beneficial microbial activity, hormonal stimulation, improved root development, enhanced water retention, reduced nitrogen loss and increased stress tolerance. This holistic approach harnesses the organic properties of these substances to optimize nitrogen availability and utilization for improved wheat crop performance. This was in conformity with the finding of (Sharma 2020).

**Phosphorous content in grain, straw and total content** (%): Primed seeds ( $M_4$ ) recorded significantly higher grain, straw and total phosphorous content (0.34, 0.17 and 0.50 %, respectively). The significantly lower grain, straw and total phosphorous content (0.30, 0.13 and 0.43 %, respectively) was recorded in un-primed seeds ( $M_1$ ) (Table 3). These might be due to organic substances offer a comprehensive approach to enhance phosphorus availability and utilization during key growth stages. Similar type of results was recorded by (Sharma 2020).

**Potassium content in grain, straw and total content (%):** Primed seeds recorded higher grain, straw and total potassium content (0.46, 1.30 and 1.76 %, respectively) as compared to un-primed seeds (0.44, 1.23 and 1.66, respectively) (Table 3). These might be due to organic substances provide a holistic approach to improve potassium availability, offering insights for sustainable and quality-focused agricultural practices. Similar type of results was recorded by (Sharma 2020).

# Nutrient Uptake by Emmer Wheat as Influenced by Seed Priming

**Nitrogen uptake in grain, straw and total uptake (kg ha**<sup>-1</sup>): Primed seeds ( $M_4$ ) recorded significantly higher nitrogen uptake in grain, straw and total uptake. The significantly lower nitrogen uptake in grain, straw and total uptake was in un-primed seeds (Table 4). This may be due to seed treatment helps in increasing the absorbing surface of root system causes increased plant growth and yield attributes and nutrient absorption by roots (N,  $P_2O_5$  and  $K_2O$ ). This was in conformity with the finding of Zheng et al (2013).

**Phosphorous uptake in grain, straw and total uptake (kg**  $ha^{-1}$ ): Primed seeds (M<sub>4</sub>) recorded significantly higher phosphorous uptake in grain, straw and total uptake (as compared to un-primed seeds (Table 4). These might be due to higher grain yield, straw yield, higher phosphorous content in grain, straw and total phosphorous content was in primed seeds as compared to un-primed seeds. This was in conformity with the finding of (Kumar et al 2021).

**Potassium uptake in grain, straw and total uptake (kg ha**<sup>-1</sup>): Primed seeds ( $M_4$ ) recorded significantly higher potassium uptake in grain, straw and total potassium uptake as compared to un-primed seeds (Table 4). This may be due to seed treatment helps in increasing the absorbing surface of root system causes increased plant growth and yield attributes and nutrient absorption by roots (N,  $P_2O_5$  and  $K_2O$ ). This was in conformity with the finding of Zheng et al (2013).

Influence of seed priming on soil chemical properties after harvest of emmer wheat crop: The chemical properties of soil *viz.*, soil reaction (pH), electrical conductivity (dS m<sup>-1</sup>) and organic carbon (%) of the soil after harvest of crop as influenced by seed priming are furnished. The seed priming, did not significantly influence the soil pH, electrical conductivity (dS m<sup>-1</sup>) and organic carbon (%) (Table 5). This lack of significant influence on soil chemical properties through seed priming may be attributed to the limited impact observed in this particular context. Similar results were obtained by Sharma (2022).

# Influence of Seed Priming on Available Nitrogen, Phosphorus and Potassium Status in Soil after Harvest of the Emmer Wheat Crop

**Available nitrogen in soil:** Seed priming significantly influenced the available nitrogen in the soil. Un-primed seeds recorded significantly higher available nitrogen in soil (246.30 kg ha<sup>-1</sup>) and was on par with  $M_1^{-1}$ . The significantly lower available nitrogen in soil was in primed seeds ( $M_2$  and  $M_4$ ) (234.70 and 237.64 kg ha<sup>-1</sup>, respectively) (Table 5). This could be attributed to the increased nitrogen uptake in primed seeds as compared to un-primed seeds. Similar results were obtained by Dhar et al (2016).

**Available phosphorous in soil:** The available phosphorus in the soil was significantly influenced by seed priming. Unprimed seeds recorded significantly higher available phosphorous in soil (30.24 kg ha<sup>-1</sup>) and was on par with  $M_3$ . The, significantly lower available phosphorous in soil was recorded in primed seeds  $M_2$  and  $M_4$  (Table 5). This could be attributed to the enhanced phosphorus uptake in primed seeds as compared to un-primed seeds (Sharma 2022).

Available potassium in soil: The seed priming did not

significantly influence the available potassium in soil. The available potassium was higher in un-primed seeds (292.00 kg ha<sup>-1</sup>) as compared to primed seeds (284.83 kg ha<sup>-1</sup>) (Table 5).

# Yield and Yield Attributes of Emmer Wheat as Influence by different Planting Geometries

Number of effective tillers (m<sup>-2</sup>): Significantly higher number of effective tillers (452.85 m<sup>-2</sup>) were recorded in 30 × 15 cm planting geometry (S<sub>1</sub>) and significantly lower effective tillers were in S<sub>2</sub> (45 × 15 cm planting geometry - $357.61 \text{ m}^{-2}$ ) (Table 1). These might be due to wider spacing facilitates plants for better utilization of nutrient, water, light and space leading to produced maximum number of effective tillers per unit area than conventional practices and under wider row spacing number of plant population should be decreases as compared to narrow row spacing. Similar results were obtained by Muchhadiya et al (2021).

Grain yield (q ha<sup>-1</sup>): Significantly higher grain yield (40.20 q ha<sup>-1</sup>) was recorded by the 30 × 15 cm planting geometry ( $S_1$ ). However, was on par with  $(S_3) 20 \times 20$  cm planting geometry. Significantly lower grain yield was in (S<sub>4</sub>) 20 cm (RPP) planting geometry (31.45 q ha<sup>-1</sup>) (Table 1). These might be due to higher yield attributes viz., number of effective tillers, number of grains per spike, spike weight, grain weight per spike and thousand grain weight was recorded in wider row spacing as compared to conventional practices. Similar results were obtained by Haque et al (2015).

Straw yield (g ha<sup>-1</sup>): The 30  $\times$  15 cm planting geometry (S<sub>1</sub>)

Table 3. Phosphorous and potassium content of emmer wheat as influenced by seed priming and different planting geometries Treatment Phosphorous content in (%) Potassium content in (%)

	Grain	Straw	Total	Grain	Straw	Total	
Main plots:- (Two genotypes with and	d without seed prir	ming)					
M₁-Gokak local (WOP)	0.30°	0.13 <sup>⁵</sup>	0.43 <sup>d</sup>	0.44 <sup>ª</sup>	1.23ª	1.66ª	
M <sub>2</sub> - Gokak local (WP)	0.31 <sup>bc</sup>	0.14 <sup>b</sup>	0.45°	0.44ª	1.27ª	1.71ª	
M <sub>3</sub> -DDK 1029 (WOP)	0.32 <sup>♭</sup>	0.16ª	0.48 <sup>b</sup>	0.45ª	1.26ª	1.71ª	
M₄-DDK 1029 (WP)	0.34ª	0.17ª	0.50ª	0.46ª	1.30ª	1.76ª	
Sub plots:- (Four planting geometries	s)						
S₁- 30 × 15 cm	0.34ª	0.16ª	0.50 <sup>ª</sup>	0.46 <sup>ab</sup>	1.28ª	1.74ª	
S <sub>2</sub> - 45 × 15 cm	0.31ª	0.16ª	0.46	0.49 <sup>a</sup>	1.29ª	1.78ª	
S₃- 20 × 20 cm	0.32ª	0.15 <sup>ab</sup>	0.47 <sup>b</sup>	0.43 <sup>ab</sup>	1.26ª	1.70 <sup>ª</sup>	
S₄- 20 cm (RPP)	0.30ª	0.14°	0.44 <sup>b</sup>	0.40 <sup>b</sup>	1.22ª	1.62ª	
Interaction (M × S)							
M <sub>1</sub> S <sub>1</sub>	0.31 <sup>b-e</sup>	0.14 <sup>de</sup>	0.45 <sup>de</sup>	0.46 <sup>a-d</sup>	1.25 <sup>ab</sup>	1.71 <sup>a-c</sup>	
$M_1S_2$	0.29 <sup>de</sup>	0.14 <sup>de</sup>	0.43 <sup>ef</sup>	0.48 <sup>a-c</sup>	1.27 <sup>ab</sup>	1.75ªb	
$M_1S_3$	0.30 <sup>с-е</sup>	0.13 <sup>ef</sup>	0.43 <sup>ef</sup>	0.43 <sup>a-d</sup>	1.23 <sup>ab</sup>	1.66ª-c	
$M_1S_4$	0.28°	0.12 <sup>f</sup>	0.40 <sup>f</sup>	0.38 <sup>d</sup>	1.15⁵	1.53°	
M <sub>2</sub> S <sub>1</sub>	0.33 <sup>a-d</sup>	0.15 <sup>cd</sup>	0.48 <sup>b-d</sup>	0.44 <sup>a-d</sup>	1.27 <sup>ab</sup>	1.71 <sup>a-c</sup>	
$M_2S_2$	0.30 <sup>с-е</sup>	0.15 <sup>cd</sup>	0.45 <sup>de</sup>	0.49 <sup>ab</sup>	1.29 <sup>ab</sup>	1.78 <sup>ab</sup>	
$M_2S_3$	0.31 <sup>b-e</sup>	0.14 <sup>de</sup>	0.45 <sup>de</sup>	0.42 <sup>a-d</sup>	1.26 <sup>ab</sup>	1.68ª-c	
$M_2S_4$	0.29 <sup>de</sup>	0.13 <sup>ef</sup>	0.42 <sup>ef</sup>	0.40 <sup>cd</sup>	1.25 <sup>ab</sup>	1.65 <sup>a-c</sup>	
M <sub>3</sub> S <sub>1</sub>	0.35 <sup>ab</sup>	0.16 <sup>bc</sup>	0.51 <sup>b</sup>	0.47 <sup>a-c</sup>	1.29 <sup>ab</sup>	1.76 <sup>ab</sup>	
M <sub>3</sub> S <sub>2</sub>	0.31 <sup>b-e</sup>	0.16 <sup>bc</sup>	0.47 <sup>cd</sup>	0.48 <sup>a-c</sup>	1.30°	1.78 <sup>a-c</sup>	
$M_3S_3$	0.33 <sup>a-d</sup>	0.15 <sup>cd</sup>	0.48 <sup>b-d</sup>	0.43 <sup>a-d</sup>	1.27 <sup>ab</sup>	1.70 <sup>a-c</sup>	
M <sub>3</sub> S <sub>4</sub>	0.30 <sup>с-е</sup>	0.15 <sup>cd</sup>	0.45 <sup>de</sup>	0.41 <sup>b-d</sup>	1.19 <sup>ab</sup>	1.60 <sup>bc</sup>	
M <sub>4</sub> S <sub>1</sub>	0.36ª	0.18 <sup>ª</sup>	0.54ª	0.48 <sup>a-c</sup>	1.30°	1.78 <sup>ab</sup>	
M <sub>4</sub> S <sub>2</sub>	0.33 <sup>a-d</sup>	0.17 <sup>ab</sup>	0.50 <sup>bc</sup>	0.50ª	1.31°	1.81ª	
$M_4S_3$	0.34 <sup>a-c</sup>	0.16 <sup>bc</sup>	0.50 <sup>bc</sup>	0.45 <sup>a-d</sup>	1.29 <sup>ab</sup>	1.74 <sup>ab</sup>	
M <sub>4</sub> S <sub>4</sub>	0.32 <sup>a-e</sup>	0.15 <sup>cd</sup>	0.47 <sup>cd</sup>	0.42 <sup>a-d</sup>	1.28 <sup>ab</sup>	1.70 <sup>ª-c</sup>	

recorded significantly higher straw yield (66.30 q ha<sup>-1</sup>) and significantly lower straw yield was in (S<sub>4</sub>) 20 cm planting geometry (57.52 q ha<sup>-1</sup>) (Table 1). These might be due to reduction in straw yield at wider row spacing's were mainly be ascribed to the decrease in overall number of plants per unit area rather than number of tillers per hill at wider row spacing's. Wider row spacing permitted better performance per hill than narrow row spacing due to decreased competition between plant for nutrient, water, space and light but decreased overall grain and straw yields might be due to lesser plant biomass production at wider row spacing's. The results corroborate with the finding of Jayawardena and Abeysekera (2011). (S<sub>1</sub>) recorded significantly higher biological yield (106.50 q ha<sup>-1</sup>) and was shown on par with S<sub>3</sub> (98.51 q ha<sup>-1</sup>). However, significantly lower biological yield was in (S<sub>4</sub>) 20 cm (RPP) planting geometry (88.96 q ha<sup>-1</sup>) (Table 1). These might be due to higher grain and straw yields was recorded in wider row spacing as compared to conventional practices (Dhar et al 2016).

# Quality Parameters of Emmer Wheat as Influenced by Planting Geometries

**Grain crude protein content of emmer wheat:** The 30 × 15 cm planting geometry ( $S_1$ ) recorded significantly higher grain crude protein content (12.13 %) and on par with  $S_3$  (11.89 %). Significantly lower grain crude protein content was in ( $S_4$ ) 20 cm (RPP) planting geometry (10.91 %) (Table 2). This is due

**Biological yield (q ha**<sup>-1</sup>): The 30 × 15 cm planting geometry

 Table 4. Nitrogen, phosphorous and potassium uptake by emmer wheat as influenced by seed priming and different planting geometries

Treatment	Νι	uptake (kg h	a <sup>-1</sup> )	Ρι	ıptake (kg h	a⁻¹)	Kι	uptake (kg h	a⁻¹)	
	Grain	Straw	Total	Grain	Straw	Total	Grain	Straw	Total	
Main plots:- (Two genotypes w	Main plots:- (Two genotypes with and without seed priming)									
M₁-Gokak local (WOP)	53.91°	28.21°	82.12°	9.67 <sup>⁵</sup>	7.82°	17.48°	14.30 <sup>b</sup>	72.25°	86.55°	
M <sub>2</sub> - Gokak local (WP)	64.20 <sup>b</sup>	31.23 <sup>bc</sup>	95.43 <sup>⊳</sup>	11.17 <sup>⊳</sup>	8.76 <sup>bc</sup>	19.93 <sup>bc</sup>	15.77 <sup>⊳</sup>	77.66 <sup>b</sup>	93.43 <sup>⁵</sup>	
M <sub>3</sub> -DDK 1029 (WOP)	63.43 <sup>b</sup>	32.58 <sup>♭</sup>	96.01 <sup>b</sup>	11.38 <sup>ab</sup>	9.52 <sup>ab</sup>	20.90 <sup>b</sup>	15.75 <sup>⊳</sup>	77.59 <sup>⊳</sup>	93.34 <sup>b</sup>	
M <sub>4</sub> -DDK 1029 (WP)	73.38ª	36.01ª	109.39ª	12.98ª	10.56ª	23.54°	17.68ª	82.96ª	100.64ª	
Sub plots:- (Four planting geor	netries)									
S <sub>1</sub> - 30 × 15 cm	76.60ª	39.88ª	116.48ª	13.62ª	10.49ª	24.11ª	18.62ª	84.84ª	103.46ª	
S <sub>2</sub> - 45 × 15 cm	61.19°	29.80°	91.00°	10.44°	9.32 <sup>♭</sup>	19.76°	16.39 <sup>b</sup>	77.72 <sup>⊳</sup>	94.11 <sup>₅</sup>	
S <sub>3</sub> - 20 × 20 cm	66.85 <sup>b</sup>	34.74 <sup>b</sup>	101.59 <sup>⊳</sup>	11.76 <sup>⊳</sup>	8.94°	20.71 <sup>b</sup>	15.86 <sup>♭</sup>	<b>77.87</b> ⁵	93.73 <sup>⊳</sup>	
S <sub>4</sub> - 20 cm (RPP)	50.28 <sup>d</sup>	<b>23.60</b> <sup>d</sup>	73.88 <sup>d</sup>	9.37 <sup>d</sup>	7.90 <sup>d</sup>	17.27 <sup>d</sup>	12.65°	70.02°	82.66°	
Interaction (M × S)										
$M_1S_1$	$66.04^{\text{de}}$	35.94⁴	101.98°	11.12 <sup>ef</sup>	8.83 <sup>f</sup>	19.95°	16.32°	<b>78.83</b> <sup>d</sup>	95.15°	
$M_1S_2$	49.83 <sup>h</sup>	24.39	74.22 <sup>9</sup>	9.22 <sup>hi</sup>	8.13 <sup>g</sup>	17.36 <sup>h</sup>	15.23 <sup>f</sup>	73.88 <sup>ef</sup>	89.12 <sup>°</sup>	
$M_1S_3$	56.49 <sup>9</sup>	31.70 <sup>f</sup>	88.19°	10.18 <sup>g</sup>	7.73 <sup>h</sup>	17.91 <sup>gh</sup>	14.57 <sup>9</sup>	73.34 <sup>f</sup>	87.91°	
$M_1S_4$	43.28 <sup>i</sup>	20.80 <sup>k</sup>	64.08 <sup>h</sup>	8.15 <sup>i</sup>	6.57 <sup>i</sup>	14.72 <sup>i</sup>	11.08 <sup>j</sup>	62.93 <sup>h</sup>	74.02 <sup>g</sup>	
$M_2S_1$	75.54 <sup>₅</sup>	38.36°	113.89 <sup>₅</sup>	14.16 <sup>b</sup>	9.96 <sup>cd</sup>	24.13 <sup>⊳</sup>	18.94 <sup>b</sup>	84.23 <sup>bc</sup>	103.17 <sup>♭</sup>	
$M_2S_2$	62.73 <sup>f</sup>	30.20 <sup>g</sup>	92.93 <sup>d</sup>	10.17 <sup>9</sup>	8.89 <sup>ef</sup>	19.06 <sup>f</sup>	16.52°	76.45 <sup>de</sup>	92.97 <sup>cd</sup>	
$M_2S_3$	68.46 <sup>cd</sup>	33.34°	101.80°	11.36 <sup>de</sup>	8.70 <sup>f</sup>	20.06°	15.29 <sup>f</sup>	77.98 <sup>d</sup>	93.27 <sup>cd</sup>	
$M_2S_4$	50.08 <sup>h</sup>	23.02 <sup>i</sup>	73.10 <sup>g</sup>	8.98 <sup>i</sup>	7.49 <sup>h</sup>	16.47 <sup>i</sup>	12.34 <sup>i</sup>	71.96 <sup>f</sup>	84.30 <sup>f</sup>	
M <sub>3</sub> S <sub>1</sub>	76.34 <sup>₅</sup>	40.07 <sup>b</sup>	116.41 <sup>b</sup>	13.46°	10.55 <sup>⊳</sup>	24.01 <sup>b</sup>	18.28°	84.82 <sup>b</sup>	103.10 <sup>b</sup>	
$M_3S_2$	62.88 <sup>f</sup>	30.69 <sup>fg</sup>	93.57 <sup>d</sup>	10.77 <sup>f</sup>	9.64 <sup>d</sup>	$20.42^{\text{de}}$	16.33°	78.29 <sup>d</sup>	94.62°	
$M_3S_3$	64.33 <sup>ef</sup>	35.80⁴	100.13°	11.76 <sup>ª</sup>	9.23°	20.99 <sup>d</sup>	15.36 <sup>f</sup>	<b>78.35</b> <sup>d</sup>	93.71 <sup>cd</sup>	
$M_3S_4$	50.16 <sup>h</sup>	23.76 <sup>ij</sup>	73.92 <sup>g</sup>	9.55 <sup>h</sup>	8.65 <sup>f</sup>	18.19 <sup>ª</sup>	13.04 <sup>h</sup>	68.90 <sup>9</sup>	81.94 <sup>f</sup>	
$M_4S_1$	88.47ª	45.15°	133.63°	15.75°	12.62ª	28.38°	20.95°	91.49ª	112.44°	
$M_4S_2$	69.34°	33.94°	103.27°	11.61 <sup>ª</sup>	10.61 <sup>b</sup>	22.22°	17.45⁴	82.27 <sup>bc</sup>	99.73 <sup>⊳</sup>	
$M_4S_3$	78.12 <sup>♭</sup>	38.13°	116.25⁵	13.75 <sup>bc</sup>	10.11°	23.86 <sup>b</sup>	18.21°	81.81°	100.02 <sup>b</sup>	
$M_4S_4$	57.58 <sup>9</sup>	26.82 <sup>h</sup>	84.40 <sup>f</sup>	10.80 <sup>f</sup>	8.90 <sup>ef</sup>	19.70 <sup>ef</sup>	14.11 <sup>9</sup>	76.27 <sup>de</sup>	90.38 <sup>de</sup>	

Mean followed by the same letter(s) did not differ significantly by DMRT (p= 0.05)

to the higher uptake of nitrogen in turn resulting in higher grain crude protein content. Higher protein content was also recorded in the system of wheat intensification when compared to normal practices also highlighted by Muchhadiya et al (2021).

Nitrogen content in grain, straw and total content (%): The 30 × 15 cm planting geometry (S<sub>1</sub>) exhibited significantly higher nitrogen content in grain, straw and total content (1.90, 0.60 and 2.50 %, respectively) which was comparable with S<sub>3</sub> (1.83, 0.56 and 2.39 %, respectively). Conversely, significantly lower nitrogen content in grain, straw and total content was observed in the 20 cm planting geometry (S<sub>4</sub>) (Table 2). The right planting geometry promotes efficient nutrient uptake, minimizes nutrient competition, enhances soil conditions for nitrogen availability and reduces nitrogen loss, all of which collectively contribute to higher nitrogen content in wheat grain. Similar results were obtained by Sharma (2020).

**Phosphorous content in grain, straw and total content** (%): Numerically higher phosphorous content in grain, straw and significantly higher total phosphorous content was recorded in 30 × 15 cm planting geometry (0.34, 0.16 and 0.50 %, respectively). The significantly lower phosphorous content in grain, straw and total content was recorded in 20 cm (RPP) planting geometry (0.30, 0.14 and 0.44 %, respectively) (Table 3). The right planting geometry supports efficient phosphorus uptake by promoting optimal root development, minimizing competition, enhancing fertilizer

 Table 5. Soil chemical properties and nutrient available status in soil after harvest of emmer wheat as influenced by seed priming and different planting geometries

Treatment	Soil pH	EC (dS m <sup>-1</sup> )	OC (%)	Avai. N (kg ha⁻¹)	Avai. P (kg ha <sup>-1</sup> )	Avai. K (kg ha <sup>-1</sup> )				
Main plots:- (Two genotypes with and without seed priming)										
M₁-Gokak local (WOP)	7.80 <sup>ª</sup>	0.37ª	0.54°	240.15 <sup>ab</sup>	30.24ª	292.00ª				
M₂- Gokak local (WP)	7.73°	0.37ª	0.55ª	234.70 <sup>b</sup>	27.82 <sup>b</sup>	284.83ª				
M₃-DDK 1029 (WOP)	7.80ª	0.37ª	0.54ª	246.30ª	29.21 <sup>ab</sup>	290.63ª				
M₄-DDK 1029 (WP)	7.75°	0.37ª	0.54ª	237.64 <sup>b</sup>	28.11 <sup>b</sup>	286.42ª				
Sub plots:- (Four planting geometries	s)									
S₁- 30 × 15 cm	7.80 <sup>ª</sup>	0.37 <sup>a</sup>	0.54ª	234.52 <sup>⊳</sup>	26.20 <sup>d</sup>	274.10 <sup>⊳</sup>				
S <sub>2</sub> - 45 × 15 cm	7.75°	0.37ª	0.54°	241.23 <sup>ab</sup>	29.55 <sup>b</sup>	293.18ª				
S₃- 20 × 20 cm	7.78 <sup>ª</sup>	0.37ª	0.54ª	235.93 <sup>₅</sup>	27.89°	287.80ª				
S <sub>4</sub> - 20 cm (RPP)	7.75 <sup>°</sup>	0.38 <sup>a</sup>	0.55ª	247.12ª	31.74ª	298.81ª				
Interaction (M × S)										
M <sub>1</sub> S <sub>1</sub>	7.80 <sup>ab</sup>	0.37 <sup>ab</sup>	0.55 <sup>ab</sup>	234.61 <sup>ef</sup>	27.39 <sup>f-h</sup>	281.90 <sup>de</sup>				
$M_1S_2$	7.70 <sup>ab</sup>	0.38 <sup>a</sup>	0.54 <sup>bc</sup>	241.90 <sup>b-e</sup>	31.58 <sup>ab</sup>	295.40 <sup>a-c</sup>				
$M_1S_3$	7.80 <sup>ab</sup>	0.36 <sup>bc</sup>	0.53°	235.40 <sup>ef</sup>	29.41 <sup>c-e</sup>	290.54 <sup>a-e</sup>				
$M_1S_4$	7.90 <sup>ª</sup>	0.37 <sup>ab</sup>	0.55 <sup>ab</sup>	248.67 <sup>ab</sup>	32.59ª	300.14 <sup>ab</sup>				
$M_2S_1$	7.90 <sup>a</sup>	0.37 <sup>ab</sup>	0.54 <sup>bc</sup>	234.60 <sup>ef</sup>	25.48	269.36 <sup>f</sup>				
$M_2S_2$	7.70 <sup>ab</sup>	0.38ª	0.55 <sup>ab</sup>	234.10 <sup>ef</sup>	28.16 <sup>e-g</sup>	289.90 <sup>a-e</sup>				
$M_2S_3$	7.70 <sup>ab</sup>	0.36 <sup>bc</sup>	0.54 <sup>bc</sup>	228.70 <sup>f</sup>	27.16 <sup>gh</sup>	284.39 <sup>e</sup>				
$M_2S_4$	7.60 <sup>b</sup>	0.38ª	0.56ª	241.39 <sup>b-e</sup>	30.48 <sup>bc</sup>	295.68 <sup>a-c</sup>				
M <sub>3</sub> S <sub>1</sub>	7.90 <sup>ª</sup>	0.36 <sup>bc</sup>	0.53°	239.40 <sup>cde</sup>	26.45 <sup>hi</sup>	278.67 <sup>ef</sup>				
$M_3S_2$	7.80 <sup>ab</sup>	0.35°	0.54 <sup>bc</sup>	248.30 <sup>ab</sup>	29.48 <sup>cd</sup>	293.89 <sup>a-d</sup>				
M <sub>3</sub> S <sub>3</sub>	7.70 <sup>ab</sup>	0.37 <sup>ab</sup>	0.53°	243.90 <sup>b-d</sup>	28.46 <sup>d-f</sup>	288.45 <sup>b-e</sup>				
$M_3S_4$	7.80 <sup>ab</sup>	0.38ª	0.56°	253.61°	32.45ª	301.50°				
$M_4S_1$	7.60 <sup>b</sup>	0.36 <sup>bc</sup>	0.53°	229.45 <sup>f</sup>	25.48 <sup>i</sup>	266.45 <sup>r</sup>				
$M_4S_2$	7.80 <sup>ab</sup>	0.36 <sup>bc</sup>	0.54 <sup>bc</sup>	240.60 <sup>b-e</sup>	28.97 <sup>de</sup>	293.51 <sup>a-d</sup>				
$M_4S_3$	7.90ª	0.37 <sup>ab</sup>	0.54 <sup>bc</sup>	235.70 <sup>d-f</sup>	26.54 <sup>hi</sup>	287.80 <sup>b-e</sup>				
$M_4S_4$	7.70 <sup>ab</sup>	0.38ª	0.53°	244.80 <sup>bc</sup>	31.45 <sup>ab</sup>	297.90 <sup>ab</sup>				

Mean followed by the same letter(s) did not differ significantly by DMRT (p= 0.05)

utilization, reducing soil fixation, encouraging beneficial associations, optimizing plant density and improving water use efficiency all of which collectively contribute to higher phosphorus content in wheat grain. Similar results were obtained by Sharma (2020).

**Potassium content in grain, straw and total content (%):** Planting geometry of 45 × 15 cm recorded significantly higher grain potassium content and numerically higher straw and total potassium content (0.49, 1.29 and 1.78 %, respectively) as compared to 20 cm (RPP) planting geometry (Table 3). The right planting geometry supports efficient potassium uptake by promoting optimal root development, minimizing competition, enhancing fertilizer utilization, reducing soil fixation, facilitating root soil interactions, optimizing plant density and improving water use efficiency all of which contribute to higher potassium content in wheat grain.

# Nutrient Uptake by Emmer Wheat as Influenced by Planting Geometries

**Nitrogen uptake in grain, straw and total N uptake (kg ha<sup>-1</sup>):** The 30 × 15 cm planting geometry ( $S_1$ ) recorded significantly higher nitrogen uptake in grain, straw and total N uptake (76.60, 39.88 and 116.48 kg ha<sup>-1</sup>, respectively). Significantly lower nitrogen uptake in grain, straw and total N uptake was in ( $S_4$ ) 20 cm (RPP) planting geometry (50.28, 23.60 and 73.88 kg ha<sup>-1</sup>, respectively) (Table 4). The right planting geometry supports efficient nitrogen uptake by promoting optimal root development, minimizing competition, enhancing fertilizer utilization, reducing nitrogen loss, facilitating symbiotic nitrogen fixation, optimizing plant density and improving water use efficiency all of which contribute to higher nitrogen uptake by wheat (Chatterjee et al 2016).

**Phosphorous uptake in grain, straw and total P uptake** (kg ha<sup>-1</sup>): Significantly higher phosphorus uptake in grain, straw and total uptake (13.62, 10.49 and 24.11 kg ha<sup>-1</sup>, respectively) was recorded by the 30 × 15 cm planting geometry ( $S_1$ ). The significantly lower phosphorous uptake in grain, straw and total uptake was recorded in ( $S_4$ ) 20 cm

(RPP) planting geometry (9.37, 7.90 and 17.27 kg ha<sup>-1</sup>, respectively) (Table 4).

**Potassium uptake in grain, straw and total K uptake (kg** ha<sup>-1</sup>): The 30 × 15 cm planting geometry ( $S_1$ ) recorded significantly higher potassium uptake in grain, straw and total uptake (18.62, 84.84 and 103.46 kg ha<sup>-1</sup>, respectively) and significantly lower potassium uptake in grain, straw and total uptake was in ( $S_4$ ) 20 cm (RPP) planting geometry (12.65, 70.02 and 82.66 kg ha<sup>-1</sup>, respectively) (Table 4). Similar results were obtained by Chatterjee et al (2016).

# Influence of planting geometry on soil chemical properties after harvest of emmer wheat crop

The planting geometries, did not significantly influence the soil pH, electrical conductivity (dS m<sup>-1</sup>) and organic carbon (%) (Table 5). This lack of significant influence on soil chemical properties through planting geometries may be attributed to the limited impact observed in this particular context. The non-significant impact of planting geometries on soil chemical properties post-harvest may be due to the inherent stability of the soil system under the studied configurations. Similar results were obtained by Sharma (2020).

# Influence of planting geometries on available nitrogen, phosphorus and potassium status in soil after harvest of the emmer wheat crop

The 20 cm (RPP) planting geometry ( $S_4$ ) recorded significantly higher available nitrogen, phosphorous and potassium in soil (247.12, 31.74 and 298.81 kg ha<sup>-1</sup>, respectively) and was found on par with  $S_2$ . In converse, significantly lower available nitrogen, phosphorous and potassium in soil was recorded in 30 × 15 cm planting geometry ( $S_1$ ) (234.52, 26.20 and 274.10 kg ha<sup>-1</sup>, respectively). These might be due to lower nutrients uptake *viz.*, nitrogen, phosphorous and potassium was recorded in 20 cm (RPP) planting geometry as compared to other planting geometries. Similar results were obtained by Dharet al (2015).

 Table 6. Correlation coefficient between grain protein, nitrogen, phosphorous and potassium content and economics of emmer wheat

Parameter	Grain protein content (%)	Total grain N content (%)	Total grain P content (%)	Total grain K content (%)	Gross returns (Rs. ha <sup>-1</sup> )	Net returns (Rs. ha <sup>-1</sup> )	B:C ratio
Grain protein content (%)	1						
Total grain N content (%)	0.937**	1					
Total grain P content (%)	0.781**	0.873**	1				
Total grain K content (%)	0.734**	0.670**	0.690**	1			
Gross returns (Rs. ha <sup>.1</sup> )	0.875**	0.912**	0.833**	0.550**	1		
Net returns (Rs. ha <sup>-1</sup> )	0.900**	0.932**	0.803**	0.644**	0.972**	1	
B:C ratio	0.880**	0.9070**	0.742**	0.688**	0.909**	0.9808**	1

\*\*Significant at 5 %

# Combined influence of seed priming and planting geometries on yield, quality parameters and nutrient uptake by emmer wheat

**Yield and yield attributes:** Primed seeds sown with  $30 \times 15$  cm planting geometry (M<sub>4</sub>S<sub>1</sub>) recorded significantly higher grain (44.13 q ha<sup>-1</sup>) and straw yields (70.64 q ha<sup>-1</sup>) and grain yield was found on par with M<sub>4</sub>S<sub>3</sub>, M<sub>3</sub>S<sub>1</sub>, M<sub>2</sub>S<sub>1</sub>, M<sub>2</sub>S<sub>3</sub> and M<sub>1</sub>S<sub>1</sub> (Table 1). The significantly lower grain yield was in M<sub>1</sub>S<sub>4</sub> (conventional line sown at 20 cm (RPP) planting geometry). This increase in grain yield might be due to the synergistic effect of seed priming with bio-formulation along with optimum planting geometry and increased yield attributes *viz.*, number of effective tillers (463.83 m<sup>-2</sup>), number of grain per spike, spike weight, grain weight per spike, thousand grain weight and spike length and is on par with M<sub>2</sub>S<sub>1</sub>, M<sub>4</sub>S<sub>3</sub> and M<sub>4</sub>S<sub>2</sub>. These findings are consistent with the outcomes reported by Haqueet al (2015) and Chatterjee et al (2016).

**Grain crude protein content:** The primed seeds sown with 30 × 15 cm planting geometry (M<sub>4</sub>S<sub>1</sub>) recorded significantly higher grain crude protein content (12.54 %) and was on par with M<sub>4</sub>S<sub>2</sub>, M<sub>4</sub>S<sub>3</sub>, M<sub>2</sub>S<sub>1</sub> and M<sub>2</sub>S<sub>3</sub> (Table 2). The significantly lower grain crude protein content was recorded in M<sub>1</sub>S<sub>4</sub> (unprimed seed with 20 cm (RPP) planting geometry). This was due to higher uptake of nitrogen in grain increasing the grain protein content.

Nitrogen, phosphorous and potassium content in grain, straw and total content of emmer wheat: The primed seeds sown with 30 × 15 cm planting geometry ( $M_4S_1$ ) recorded numerically higher nitrogen and phosphorous content in grain, straw and total content. However, higher potassium content in grain, straw and total content was recorded in primed seeds sown with 45 × 15 cm planting geometry ( $M_4S_1$ ) as compared to un-primed seeds sown with 20 cm (RPP) planting geometries (Table 2, 3). These might be due to the combination of seed priming and strategic planting geometries positively influences nutrient content in wheat grain by improving germination efficiency, optimizing root development, enhancing nutrient uptake systems, minimizing competition, facilitating soil-root interactions, promoting efficient fertilizer utilization and reducing nutrient loss. This is in accordance with the findings of Sharma (2020)

Nutrient uptake patterns of emmer wheat: The primed seeds sown with 30  $\times$  15 cm planting geometry (M<sub>4</sub>S<sub>4</sub>) recorded significantly highest uptake of N, P and K by grain, straw and total uptake and lowest uptake of N, P and K was recorded in M<sub>1</sub>S<sub>4</sub> (un-primed seed with 20 cm (RPP- planting geometry) (Table 4). The system of wheat intensification recorded higher uptake of N, P and K as compared to conventional method of cultivation which could be ascribed to better vegetative and reproductive growth under SWI method of cultivation. Wider spacing reduced the above and below ground competition, well developed and healthy root system causes increase N, P and K uptakes. Further primed seed recorded higher nutrient uptake than un-primed seed. This may be due to the fact that seed treatment helps in increasing the absorbing surface of root system causes increased plant growth and yield attributes and nutrient absorption by roots (N,  $P_2O_5$  and  $K_2O$ ). This is in accordance with the findings of Chandrapala et al (2010) and Zheng et al (2013).

Soil fertility status as influenced by varied planting geometries and seed priming: Combination of un-primed seeds sown with 20 cm (RPP) planting geometry ( $M_3S_4$ ) recorded significantly higher soil available N , P , and K , after harvest of emmer wheat and it was on par with  $M_3S_2$  ,  $M_1S_4$  ,  $M_1S_2$  and  $M_4S_4$ . The significantly lower soil available N, P and K after harvest of wheat was in  $M_4S_1$  and  $M_2S_1$  (primed seed with 30 × 15 cm planting geometry) (Table 5). This might be due to higher uptake of N, P and K in primed seed sown with 30 × 15 cm planting geometry. Similar results were obtained by Sharma (2022).

Influence of seed priming and planting geometries on economics: Among the main plots, primed seeds ( $M_4$ ) recorded significantly higher gross return (₹1,41,915 ha<sup>-1</sup>), net returns (₹92,033 ha<sup>-1</sup>), and B:C ratio (2.88) and was on par with  $M_2$  and  $M_3$ . The significantly lower gross returns, net

 
 Table 7. Correlation coefficient between Macro nutrient uptake by emmer wheat and available macro nutrient in soil after harvest of emmer wheat

Parameter	Total N uptake (kg ha⁻¹)	Total P uptake (kg ha <sup>-1</sup> )	Total K uptake (kg ha <sup>-1</sup> )	Available N in soil (kg ha⁻¹)	Available P in soil (kg ha⁻¹)	Available K in soil (kg ha⁻¹)
Total N uptake (kg ha <sup>-1</sup> )	1					
Total K uptake (kg ha <sup>-1</sup> )	0.961**	1				
Total K uptake (kg ha <sup>-1</sup> )	0.958**	0.959**	1			
Available N in soil (kg ha <sup>-1</sup> )	-0.669**	-0.512**	-0.625**	1		
Available P in soil (kg ha <sup>-1</sup> )	-0.937**	-0.830**	-0.874**	0.795**	1	
Available K in soil (kg ha <sup>-1</sup> )	-0.870**	-0.817**	-0.838**	0.745**	0.911**	1

\*\*Significant at 5 %

Table 8.	Economics of	emmer wheat	as influenced b	v seed i	priming an	d different	planting	aeometries
				, ,				

Treatment	Cost of cultivation (Rs ha <sup>-1</sup> )	Gross returns (Rs ha <sup>-1</sup> )	Net returns (Rs ha <sup>-1</sup> )	B:C ratio
Main plots:- (Two genotypes with and w	ithout seed priming )			
M₁-Gokak local (WOP)	48400°	121192 <sup>⊾</sup>	72792 <sup>b</sup>	2.53⁵
M₂- Gokak local (WP)	49022ª	133253ªb	84231 <sup>ab</sup>	2.76 <sup>ab</sup>
M₃-DDK 1029 (WOP)	48511°	129284 <sup>ab</sup>	80773 <sup>ab</sup>	2.70 <sup>ab</sup>
M <sub>4</sub> -DDK 1029 (WP)	49883ª	141915°	92033°	2.88ª
Sub plots:- (Four planting geometries)				
S₁- 30 × 15 cm	<b>46247</b> <sup>⁵</sup>	147989°	101742 <sup>ª</sup>	3.20ª
S₂- 45 × 15 cm	45834 <sup>b</sup>	125890°	80056°	2.75°
S₃- 20 × 20 cm	47162 <sup>⊳</sup>	135377⁵	88215 <sup>⊳</sup>	2.87 <sup>⁵</sup>
S₄- 20 cm (RPP)	56572°	116388⁴	59816 <sup>⁴</sup>	2.06 <sup>d</sup>
Interaction (M × S)				
M <sub>1</sub> S <sub>1</sub>	46082 <sup>d</sup>	134009 <sup>ef</sup>	87927 <sup>ef</sup>	2.91 <sup>⁵</sup>
M <sub>1</sub> S <sub>2</sub>	<b>45702</b> <sup>d</sup>	116698 <sup>h</sup>	70996	2.55 <sup>h</sup>
$M_1S_3$	46234 <sup>d</sup>	125807°	79573 <sup>h</sup>	2.72 <sup>g</sup>
M <sub>1</sub> S <sub>4</sub>	55582 <sup>b</sup>	108254	52672	1.95 <sup>k</sup>
$M_2S_1$	<b>46362</b> <sup>d</sup>	155474⁵	109112 <sup>b</sup>	3.35⁵
$M_2S_2$	45926⁴	125963°	80037 <sup>h</sup>	2.74 <sup>fg</sup>
$M_2S_3$	<b>46536</b> <sup>⁴</sup>	136344 <sup>de</sup>	89808°	2.93 <sup>de</sup>
$M_2S_4$	57262 <sup>ab</sup>	115229 <sup>h</sup>	57967 <sup>k</sup>	2.01 <sup>jk</sup>
M <sub>3</sub> S <sub>1</sub>	46132 <sup>d</sup>	140234 <sup>ª</sup>	94102 <sup>d</sup>	3.04°
$M_3S_2$	<b>45742</b> <sup>d</sup>	129102 <sup>fg</sup>	83360 <sup>°</sup>	2.82 <sup>ef</sup>
$M_3S_3$	<b>46288</b> <sup>d</sup>	130965 <sup>r</sup>	84677 <sup>fg</sup>	2.83 <sup>ef</sup>
$M_3S_4$	55882ªb	116836 <sup>h</sup>	60954 <sup>ĸ</sup>	2.09 <sup>ij</sup>
$M_4S_1$	46412 <sup>d</sup>	162240°	115828°	3.50ª
$M_4S_2$	45966 <sup>d</sup>	131798 <sup>ef</sup>	85832 <sup>fg</sup>	2.87°
$M_4S_3$	49590°	148392°	98802°	2.99 <sup>cd</sup>
$M_4S_4$	57562°	125232 <sup>°</sup>	67670j	2.18 <sup>i</sup>

Mean followed by the same letter(s) did not differ significantly by DMRT (p= 0.05)

returns and B:C ratio was recorded in un-primed seeds (M<sub>1</sub>). Within the sub plot, 30 × 15 cm planting geometry (S<sub>1</sub>) recorded significantly higher gross return (₹1,47,989 ha<sup>-1</sup>), net returns (₹1,01,742 ha<sup>-1</sup>), and B:C ratio (3.20) when compared to (S<sub>4</sub>) conventional line sown at 20 cm (RPP) planting geometry. Among the interaction effect, primed seeds sown with 30 × 15 cm planting geometry (M<sub>4</sub>S<sub>1</sub>) recorded significantly higher gross return (1,62,240 ha<sup>-1</sup>), net returns (1,15,828 ha<sup>-1</sup>), and B:C ratio (3.50) (Table 8) when compared to un-primed seeds with conventional line sown at 20 cm (RPP) planting geometry (M<sub>1</sub>S<sub>4</sub>). This might be due to lower usage of external input *viz.*, very low seed rate, higher grain yield, straw yield and less labour requirement *etc.* Similar type of results was also obtained by Kumar et al (2015) and Bhargava et al (2016).

### CONCLUSION

Among the different treatment combinations, primed seeds sown with 30 × 15 cm planting geometry recorded significantly increased number of effective tillers, grain yield, straw yield, biological yield, grain crude protein content, macro nutrients content with greater uptake and higher economics. Thus, system of wheat intensification is best suitable option for resource poor farmers for getting higher yield and profit in irrigated ecosystem of Northern Dry Zone of Karnataka (Zone-3).

## REFERENCES

Anonymous 2021. Area, production and productivity of wheat in Karnataka. www. indiastat. com. Available at: https://www.indiastat.com/data/agriculture/wheat/data-year/allyears all-years and Accessed on: 13 July 2022.

- Anonymous 2022. Agricultural statistics at a glance Directorate of economics and statistics, Ministry of Agriculture and Farmers Welfare. Govt. of India. Available at: https://desagri.gov.in Agricultural-Statistics and Accessed on: 12 may 2023.
- Anonymous 2022. Fourth advance estimates of production of food grains for 2021-22 as on 17. 08. 2022, Directorate of economics and statistics, Ministry of Agriculture and Farmers Welfare. Govt. of India. Available at: 17 Aug 2022.
- Anonymous 2009. SWI: An Innovation, Dehradun, India. Available at: https://peoplesscienceinstitute.org/ and Accessed on: 22 may 2021.
- Abraham B, Araya H, Berhe T, Edwards S, Gijja B and Khadka R B 2014. The system of crop intensification: Reports from the field on improving agricultural production, food security and resilience to climate change for multiple crops. *Agriculture and Food Security* **3**(4): 1-12.
- Adhikari D 2013. Short communication; system of wheat intensification in farmer's field of Sindhuli, Nepal. Agronomy Journal of Nepal 3: 169-171.
- Adhikari D 2013. System of wheat intensification in farmer's field of Sindhuli, Nepal. Agronomy Journal of Nepal 3: 36-39.
- Anonymous 2021. Area, production and productivity of wheat in Karnataka. www.indiastat.com.
- Anonymous 2022. Agricultural statistics at a glance, Directorate of economics and statistics, Ministry of Agriculture and Farmers Welfare. Govt. of India.
- Anonymous 2022. Fourth advance estimates of production of food grains for 2021-22 as on 17. 08. 2022, Directorate of economics and statistics, Ministry of Agriculture and Farmers Welfare. Govt. of India.
- Bhargava C, Deshmukh G, Sawarkar SD, Alawa SL and Ahirwar J 2016. The system of wheat intensification in comparison with convention method of wheat line sowing to increase wheat yield with low input cost. *Plant Archives* 16(2): 801-804.
- Bhoopathi PK, Kulandaivel S, Surendra T and Ravi S 2001. Effect of sett treatment on germination and productivity of seed cone. *Annuals of Agricultural Research* **22**(2): 271-272.
- Black CA 1965. Soil Plant relationship. First edition Wiley Eastern, Private Limited, New Delhi.
- Blackman VH 1919. The compound interest law and plant growth. Annals of Botany **33**(131): 353-360.
- Chandrapala AG, Yakadri M, Kumar RM and Raj GB 2010. Productivity and economics of rice (*Oryza sativa* L.) - maize (*Zea mays* L.) as influenced by methods of crop establishment, Zn and S application in rice. *Indian Journal of Agronomy* **55**: 171-176.
- Chatterjee K, Singh CS, Singh AK, Singh AK and Singh SK 2016. Performance of wheat cultivars at varying fertility levels under system of wheat intensification and conventional method of wheat production system. *Journal of Applied and Natural Science* 8(3): 1427-1433.
- Chawla OP 1986. Advances in biogas technology publication and information division. Indian Council of Agricultural Research. New Delhi, India 75-95.
- Dhar S, Barah BC, Vyas AK and T Uphoff N 2016. Comparing system of wheat intensification (SWI) with standard recommended practices in the northwestern plain zone of India. *Archives of Agronomy and Soil Science* **62**(7): 994-1006.
- Donald CM 1962. Competition among crop and pasture plants. *Advances in Agronomy* **15**: 1-118.
- FAO 2020. FAOSTAT, *Statistics Division* Food and Agriculture Organization of the United Nation, Rome. http:// faostat. Fao. org/default.Aspx.
- Gomez KA 1972. Techniques for Experimentation with Rice. International Rice Research Institute, Manila, Philippines.

Received 01 July, 2024; Accepted 21 September, 2024

- Gomez KA and Gomez AA 1984. *Statistical Procedure for Agricultural Research*, 2nd Ed., A Wiley-International Science Publication, New York (USA), **p**.680.
- Haque M, Chowdhury AR, Acharya SS, Ghosh M, Kumar S and Kumar N 2015. Effect of planting geometry and seed treatment on growth and yield of wheat (*Triticum aestivum* L.) under system of wheat intensification. *International Journal of Bio-resource and Stress Management* **6**(3): 335-340.
- Jackson ML 1967. Soil Chemical Analysis, Prentice, Hall India, Pvt. Ltd. New Delhi, pp. 67-70.
- Jackson ML 1973. Soil chemical analysis. Prentice Hall of India Pvt. Ltd., New Delhi. p. 498.
- Kamalam J and Rajappan N 1989. Effect of seed hardening on germination and seedling vigour in paddy. Seed Research 17(2): 188-190.
- Khadka RB and Raut P 2011, System of wheat intensification (SWI). A new concept on low input technology for increasing wheat yield in marginal land. Mercy Crops, Nepal. Available from http://sri.ciifad.cornell. edu/countries/Nepal/ Nepal\_ SWI\_ Khadka11.pdf.
- Kjeldahl C 1883. A new method for the determination of nitrogen in organic matter. *Journal of Analytical Chemistry* **22**:366.
- Kumar A, Raj R, Dhar S and Pandey UC 2015. Performance of system of wheat intensification (SWI) and conventional wheat sowing under North Eastern Plain Zone of India. *Annual of Agriculture Resources*, New Series **36**(3): 258-262.
- Kumar D, Pathak SK, Kumari S, Shri A, Kumari S, Kumari P, Chandini RK, Kumari V and Ranjan P 2021. Effect of seed priming and seed rate on LAI and uptake of nutrients in wheat (*Triticum* aestivum L.). The Pharma Innovation Journal **10**(10): 779-782,
- Muchhadiya RM, Thanki RB, Sakarvadia HL and Muchhadiya PM 2021. Effect of NPK levels and city compost on soil chemical and biological properties, yield and quality of wheat (*Triticum aestivum* L.) under system of wheat intensification (SWI). *The Pharma Innovation Journal* **10**(11): 1230-35.
- Piper CS 2002. Soil and plant analysis, Academic press, New York, pp. 47-77.
- Sarlach RS, Sharma A and Bains NS 2013. Seed priming in wheat effect on seed germination, yield parameters and grain yield. *Progress Research* 8: 109-112.
- Sharma J, Sharma BC, Puniya R and Jamwal S 2020. Effect of seed priming and plant geometry on yield and economics of wheat in modified system of wheat intensification under sub-tropical conditions of Jammu. *Indian Journal of Ecology* **49**(5): 1696-1699.
- Singh SK 2015. Cultivation of dicoccum wheats in India. Krishisewa p. 1-4
- Thakur AK, Uphoff N and Antony E 2010. An assessment of physiological effects of system of rice intensification (SRI) practices compared with recommended rice cultivation practices in India. *Experimental Agriculture* **46**(1): 77-98.
- Thapa T, Chaudhary P and Ghimire S 2011. Increasing household food security through system of wheat intensification (SWI) techniques. Mercy Crops Nepal.
- Toklu F, Baloch FS, Karakoy T and Ozkan H 2015. Effects of different priming applications on seed germination and some agro morphological characteristics of bread wheat (*Triticum aestivum* L.). *Turkish Journal of Agriculture and Forestry* **39**(6): 1005-1013.
- Uphoff NT, Marguerite JD, Bahera D, Verma AK and Pandian BJ 2011. National colloquium on system of crop intensification (SCI). Field immersion of system of crop intensification (SCI), Patna.
- USAID 2009. The System of wheat intensification (SWI) in Mali. Available at: http://sri.cals.cornell.edu/countries/nepal /Nepal\_SWI\_Khadka11.pdf and Accessed on: 2 sept 2017.



# Nutrient Release Rate of *Crotolaria juncea* Degradation with Incorporation of Ligno-cellulolytic Bioinoculant under Dry and Flooded Conditions

Jaspreet Kaur, S.K. Gosal, S.S. Walia<sup>1</sup>, Jupinder Kaur and Neha Khipla

Department of Microbiology, <sup>1</sup>School of Organic Farming Punjab Agricultural University, Ludhiana-141 004, India E-mail: jaspreetkaursian@gmail.com

**Abstract**: The application of green manure in the field needs fallow period of 20-30 days so as to allow decomposition, the application of bacterial culture that can enhance the degradation rate can decrease the fallow period and allow early sowing of next crop thereby saving time. The present study was conducted to evaluate the nutrient release during decomposition of *Crotolaria juncea* with application of efficient lignincellulose degrading bacterial consortium in laboratory under dry and flooded conditions. The study reported 15 times higher decomposition during first eight days of incubation under aerobic conditions with application of bacterial consortium. Application of consortium culture was more effective in degradation as compared to control and single culture application. Significant increase in  $NO_3$ -N content was observed during incubation period whereas,  $NH_4^+$ -N content showed peak increase at 45 days of incubation. Aerobic conditions supported rapid release of both  $NH_4^+$ -N and  $NO_3^-$ -N content during initial days of incubation. In flooded water regimes nitrogen release ( $NH_4^+$ -N +  $NO_3^-$ -N) was low owing to more denitrification activity in water logged conditions. The phosphorus (P) and potassium (K) content showed positive relation with decomposition of green manure. The application of bacterial consortium has further enhanced the release of P and K as compared to control treatment. This might be due to enhanced degradation that led to release of these nutrients. There was significant impact of water regimes on phosphorous release, as remarkable increase was in P value under flooded conditions. The study showed that application of ligno-cellulolytic consortium bioinoculant enhanced the decomposition as well as nutrient release form *Crotolaria juncea*.

Keywords: Bacterial consortium, Decomposition, Nutrient release, NH4+-N, Phosphorous

Environmental contamination has the potential to be a major threat to the survival of living organisms. The misuse of chemical fertilizers and pesticides can contribute to the deterioration of the environment. This can affect soil fertility resulting in a loss of productivity, and this realization has led to the adoption of sustainable farming practices (Vogel et al 2019). Green manure crops have an increasingly important role in sustainable crop production. The incorporation of green manure in soil plays a key role in the nutrient cycling in the soil ecosystems. Their incorporation in soil causes recycling of nutrients bound in plant biomass through microbial decomposition (Kalita et al 2022), thus making them available for crop uptake (Lei et al 2022). The overall nutritional impact of green manure on crop primarily depend on its efficient decomposition and nutrient release. Organic material decomposition is a complex process determined by three main interacting groups of factors: biotic (microorganisms and invertebrates that take part in litter decomposition), chemical (composition of the litter) and physical such as climate and environment surrounding the litter (Berg and Laskowski 2006). There are several naturally occurring microorganisms that are able to decompose organic waste leading to nutrient mineralisation. These microorganisms are also important to maintain nutrient flows from one system to another and to minimize ecological imbalance (Umsakul et al 2010). Microbes utilize organic waste for their growth and nutrition, when their requirements are met or microbial cells are lysed, excess ions got released into the soil pore water, which is accessible to plant roots. Thus, playing vital role in organic matter decomposition and nutrient mineralization (Duan et al 2021).

Decomposition of organic inputs is depended on survival, colonization and growth of microorganisms associated with it. Although, number of soil microorganisms are able to degrade organic inputs, but; scarce number or limited biodegradation capacity of the autochthonous microbes may result in inefficient and delayed decomposition rates (Xu et al 2019). Therefore, the in-situ degradation of organic amendments can be enhanced by augmentation of potential lignocellulolytic microorganisms into the system (Bhattacharjya et al 2021). The discovery of novel bacterial cultures for biodegradation of lignocellulosic materials is gaining much attention because of their inherent metabolic diversity, higher growth rate, genome variability, biotechnological significance and immense environment stability (Zhao et al 2019). This paper will focus on studying the role of bacterial inoculants in boosting the decomposition and nutrient mineralization from Crotolaria juncea in lab conditions.

#### MATERIAL AND METHODS

The experiments were carried out in the Department of Microbiology, Punjab Agricultural University, Punjab, India. The soil used in the experiment had a pH of 8.46, 5.29 mg/kg available N, 1.32 mg kg<sup>-1</sup> Olsen P, 1.60 mg/kg exchangeable potassium and an electrical conductivity of 0.20dS/m. The bulk soil sample was collected fresh from the experimental field and was air-dried to sieve through 2 mm pore size. The experiment was conducted in two types of soil *i.e.*, sterilized and unsterilized soil. The fresh soil was autoclaved at 15psi (121°C) for 30 minutes, cooled and again autoclaved for three consecutive days. The experiments was carried out in polypropylene bags (18"×12") in triplicate.

**Organic amendments:** The green manure crop (*Crotolaria juncea*) was grown to the age of 45 days in the experimental field of School of Organic Farming, PAU, Ludhiana. The freshly collected green-manure plants were chopped into pieces (3.5-5.5 cm long) for use in this study. Green manure and soil were mixed in 1:2 ratio and contents were thoroughly hand-mixed into the soil.

**Bacterial cultures:** Three bacterial culture were isolated from different soil samples, screened and selected for their cellulose degrading ability namely BC1, BC2 and BC3 which were applied singly and in mixture of three known as consortium bioinoculants. The control treatment devoid of any bacterial culture was also kept during the study. The bacterial culture was identified molecularly as BC1-*Lysinibacillus sp.* VKM B-751 (MT539731.1); BC-2-*Glutamicibacter mishrai* strain S5 (CP032549.1); and BC3-*Paenibacillus sp.* A3 (HG003584.2).

**Moisture regime:** The nutrient transformations in the amended soil were studied at 37±5°C (room temperature) at two moisture regimes using a completely randomized design with two replications. The two moisture regimes were: (i) Aerobic condition (field capacity): The soil was maintained at around 14.0% moisture (w/w). The soil moisture was maintained by adding water after every week. (ii) Flooding conditions: A 1.0 cm layer of water was maintained at the surface of the soil throughout the study period by adding water every 2 days. Total of ten treatments were designed to determine decomposition and nutrient mineralization pattern of *Crotolaria juncea* in two different water regimes (aerobic and flooding) as follows-

**Soil sampling and analysis:** Decomposition of green manure was monitored by the loss of green manure weight at 2 days interval upto  $14^{\text{th}}$  day. Soil samples (10 g) were taken at 2, 4, 6, 8, 10, 12, 14 day and analysed for available phosphorous and exchangeable potassium whereas mineral N (NH<sup>+</sup>-N + NO<sub>3</sub><sup>-</sup>-N) was recorded at 16, 30, 45 and 60<sup>th</sup> day from each treatment (Table 1). The soil phosphorous,

potassium content and Mineral nitrogen were determined using methods given by Olsen et al. (1954), Jackson (1967) and Page et al. (1982), respectively.

**Statistical analysis**: The mean values recorded for different observation and results were subjected to analysis of variance and difference between treatments were analysed by Tukey' b test using SPSS 16.0 software.

#### **RESULTS AND DISCUSSION**

Green manure decomposition: Decomposition was recorded in terms of green manure weight loss during the studies. Weight of green manure was significantly affected by the application of bacterial culture in the aerobic conditions than flooding conditions. However, the significant amount of weight loss occurred during the first eight day of green manure incubation with unsterilized soil + consortium culture (Fig. 1a). In sterilized soil, green manure weight loss rate was 4.49 % day<sup>-1</sup> with the application of the consortium culture which was significantly higher over its uninoculated control i.e., 3.56% day<sup>-1</sup>. Irrespective of the inoculation, there was a higher weight-loss rate in unsterilized soil treatments as compared to sterilized soil. The application of consortium in unsterilized soil increased weight loss rate by 12 times than that of sterilized conditions. Application of consortium culture increased the green manure weight loss during first week by 5.7 % day<sup>-1</sup> which was significantly higher than its uninoculated control (4.14% day<sup>-1</sup>) in sterilized soil. In unsterilized soil this value further increased to 6.38% day <sup>1</sup>with consortium as compared to 5.37% day<sup>-1</sup> in its uninoculated control. The present study revealed that application of consortium to green manure accelerated the decomposition in terms of weight loss rate during first week of incubation. Silva et al (2008) observed the decomposition process associates with the compounds that are degraded in each phase, as a result decomposition occurs faster in the early stages when easily assimilable substrates are available to microbes and slows down when only complex material was available to microbes.

Decomposition of green manure significantly affected by the moisture regimes. Weight loss in flooding conditions

	Table	1. Details	of ex	perimenta	l treatment
--	-------	------------	-------	-----------	-------------

Treatment	Details	Treatment	Details
T1	SS+ GM	T6	US+ GM
T2	SS+ GM+BC1	T7	US+ GM+BC1
Т3	SS+ GM+BC2	Т8	US+ GM+BC2
T4	SS+GM+BC3	Т9	US+GM+BC3
T5	SS+GM+C	T10	US+GM+C

\*SS-Sterilized soil; GM-Green manure; US-Unsterilized soil; BC-Bacterial culture; C- Consortium

indicated that the application of bacterial culture (single /consortium) positively affected the decomposition but not to the extent in aerobic condition (Fig. 1b). Application of single culture in sterilized soil enhanced the weight loss by 1.40-1.46 % day<sup>-1</sup>, whereas application of consortium enhanced the weight loss by 2.22% day-1. Use of unsterilized soil significantly enhanced the weight loss of organic matter in each case, like in its uninoculated control treatment weight loss enhanced by 0.08% day<sup>-1</sup>as compared to uninoculated control in sterilized soil. Application of consortium culture significantly enhanced the percent weight loss day<sup>-1</sup> (2.71) as compared to the control treatment (2.31) during first eight days of incubation. Awasthi et al (2018) reported that bacterial assisted composting of food waste has attained early maturation as compared to uninoculated control; thereby improving C/N ratio, extractable nitrogen (ammonium) content, pH and EC. Manu et al (2019) concluded that microbial inoculation caused early waste stabilization and statistically impacted the degradation rate. Similarly, Greff et al (2022) also mentioned an improved biodegradation of organic inputs using ligninolytic or cellulolytic microbial inoculants. The inoculation with efficient



Fig. 1. Weight reduction of green manure using cellulose degrading bacteria in sterilized and unsterilized soil at different time intervals in a) aerobic conditions and b) in flooding conditions

microbes may help in promoting decomposition by reducing the initial lag phase, increasing the concentration of degrading enzymes, promoting the degree of humification and various nutrient transformations (Xu et al 2019, Chi et al 2020).

Effect of bacterial inoculants on mineral nitrogen content of soil: In present investigations, an increase in the concentration of mineral nitrogen was brought by decomposition of organic matter bound nitrogen which was mediated by inoculated and indigenous microorganisms present under different soil and moisture regimes.

Nitrogen release under aerobic conditions: The NH<sup>+</sup>-N content was significantly increased after the incorporation of green manure as compared to the 0 day content (5.29 mg/kg. In all treatments, during the first two weeks, NH<sup>+</sup>-N content increased significantly over the control afterwards it increased slowly in all treatment. The rapid rise in NH<sup>+</sup><sub>4</sub>-N during early period might be due to decomposition of the easily decomposable nitrogenous substances present in Crotolaria juncea. In sterilized soil, the highest mineralization rate of NH4+-N was observed with the inoculation of BC1 as compared to sole application of other bacterial culture, this might be attributed to the inherent capacity of ammonia released by bacteria (Table 2). However, application of culture as consortium showed further increase in the rate of NH4+N release which indicated the combined efficiency of culture in decomposition of Crotolaria juncea. The release of NH4+N was higher in unsterilized soil as compared to sterilized soil in all treatments showing that the inoculated bacterial culture had worked coordinatively with the natural microflora of soil and positively affected the release of nitrogen. As compared to ammoniacal nitrogen, the NO3-N content of the soil increased gradually throughout the incubation period (Table 2). The maximum increase in the nitrification was recorded during the 8th to 12th day of incubation in all treatments. The rapid increase in soil NO<sub>3</sub>-N content was observed after the 30<sup>th</sup> day indicating that nitrification dominated over the process of ammonification and may be due to stimulation of nitrifying bacteria in response to higher availability of ammoniacal substrate and aerobic soil condition during decomposition of Crotolaria in soils.

The uninoculated control soil also showed a considerable increase in the mineral N content during incubation. This might be due to decomposition of added organic materials that had a positive priming action in increasing mineral N. Present results corroborated with Trinsoutrot et al (2000) that residues having C/N ratio ≤24 induced a surplus of mineral N following addition to soil. Mohanty et al (2011) also reported an immediate nitrogen mineralization on application of

Gliricidia (green manure crop) to soil as rate of nutrient mineralization also dependent on chemical composition of the residue and green manures having low C/N ratio and polyphenols were expected to be easily degraded by soil microbes; releasing nutrients (N) into the soil. In present study an increased N mineralization was recorded under consortium inoculation than uninoculated controls. This could be related to the inherent ability of the inoculated bacterial strains to produce and excrete ammonia or their synergistic effect on the improved degradation of nitrogenous substrates. Resende et al (2000) observed that N contained in stems and leaves of Crotalaria juncea was linked to more easily decomposable compounds. Awasthi et al (2020) reported enhanced the extractable ammonium content from food waste with bacterial inoculation as compared to uninoculated control, by supporting degradative and maturation processes. Wu et al (2019) reported that mixed culture of Trichoderma viride and T. harzianum improved total N, P and K content of corn straw – pig manure and rice straw - pig manure compost over the uninoculated control.

Nitrogen release under flooding condition: Ammoniacal nitrogen content of soil-organic matter mixtures increased during incubation under flooding conditions but, to a lesser extent as compared to the aerobic conditions and can be due to decomposition of organic materials takes place under flooding conditions at slower rate as compared to aerobic conditions. The application of consortium bioinoculant significantly increased NH<sup>+</sup><sub>4</sub>-N content at all-time intervals as compared to uninoculated control treatment. Highest NH<sup>+</sup><sub>4</sub>-N was observed at 45<sup>th</sup> day of incubation and then declined progressively. The NO<sub>3</sub>-N content too has risen slightly only during the 1st week under flooded conditions. After the 8<sup>th</sup> day of inoculation, an immediate decrease was observed in NO<sub>3</sub>-N content in all treatments (Table 2). The results could be supported in terms that nitrification is an oxidation process occurring under aerobic conditions whereas, slight increase could possibly due to the aerobic conditions prevailed at some void areas making the process feasible for a short period. Additionally, increase in NH<sup>+</sup>-N up to 45 days of incubation might have favour denitrification under flooding conditions. Higher availability of nitrate N was for BC1 culture and might be due to higher NH4-N produced with application of this culture that acted as precursor for the nitrate production. Degradation of green manure in the presence of microbial inoculants could significantly improve mineral nitrogen content. The flooded soil is a dynamic heterogeneous soil-water system that has three distinct soil layers established mainly by the prevailing oxidationreduction or redox potential (Eh or pE) of the system. soil  $NO_3-N$  concentrations decreased and soil  $NH_4-N$ concentrations increased with flooding as expected under the anaerobic soil conditions reached under flooding. Under anaerobic conditions, three main N transformations occur: (i) ammonification (the conversion of organic N into NH<sub>4</sub>-N), (ii) nitrate reduction (conversion of NO<sub>3</sub>–N into  $NH_4$ –N), or (iii) denitrification (conversion of NO<sub>3</sub>-N into N<sub>2</sub>). Therefore, under flooded conditions, NH<sub>4</sub>-N is being produced (ammonification and nitrate reduction) while NO<sub>2</sub>-N is being lost (nitrate reduction and denitrification). Nitrate is a mobile form of N and thus it can also be leached from the system under flooded conditions. Unger et al. (2009) also observed that decrease in NO<sub>3</sub>-N and an increase in NH<sub>4</sub>-N was observed in the 5-week flood treatments, however no differences were observed between the flowing and stagnant treatments. Hefting et al. (2004) demonstrated that the height of the water table can affect the outcome of N mineralization. In their study, when the water table was within 10 cm of the soil surface, ammonification was the primary N mineralization reaction taking place while net nitrification was insignificant.

# Effect of bacterial inoculants on soil phosphorus availability

under aerobic conditions: Application of green manure significantly increased the phosphorous content in soil as



Fig. 2. Effect of different bacterial cultures on phosphorous mineralization from green manure in a) in aerobic conditions, b) in flooding conditions

compared to the 0 day (1.60 mg/kg). The phosphorous content was increased up to the 14<sup>th</sup> days of incubation however, application of bacterial culture positively influenced phosphorous levels in soil. Among the sole inoculants, BC3 culture significantly enhanced the phosphorous level as compared to uninoculated control treatments. The highest phosphorous content was in treatment having application of consortium culture. In sterilized soil conditions, there was about 6.0% increase in the phosphorous level with consortium inoculation over its uninoculated control (Fig. 2). This could be directly related to the enhanced decomposition of green manure due to combined activity of cultures present in consortium in sterile soil. However, the phosphorous content in treatments with unsterilized soil was found to be higher than sterilized soil, which indicated the cooperation derived from indigenous microbial flora in degradation biomass and phosphorous mineralization. Decomposition of green manure causes release of high chelation capacity products which lowers the activity of polyvalent that form insoluble salt with phosphorous thus liberates P from the basic phosphates of these elements. In present study, the phosphorous availability positively correlated with the nitrogen content. This might be due to the fact that soils with high ammonification activities usually favour the processes resulting in rapid release of phosphorous in soil.

Soil phosphorous availability under flooding conditions: The different water regimes significantly affected the soil available phosphorous content during green manure decomposition. The available phosphorous increased markedly in the soil maintained under flooding conditions as compared with aerobic conditions. However, in accordance to aerobic conditions, the highest available phosphorous was in treatment having bacterial consortium. The bacterial inoculation was more effective in improving phosphorous availability from green manure decomposition, indicating that bacterial inoculant effectively coordinated with the indigenous microbial flora (Fig. 2).

Chimouriya et al. (2018 and) Kumar et al (2020) also documented that green manures can affect soil P availability through several mechanisms, such as (i) mineralization of biomass P; (ii) increased soil microbial activity due to release of phytases or phosphatases may transform the unavailable P to available P; (ii) Decomposition released  $CO_2$  that react with water to form carbonic acid in soil solution thereby facilitating dissolution of P (iv) Released organic compounds may mask various P adsorption sites in soil or may release P from these sites by anion exchange method. Zhou et al (2021) observed rapid P release from green manure (milk vetch) during first five days, followed by a gradual increase up to 125 days. This rapid release of unstructured P (40-60 % of the total plant P) may be attributed to nucleic acid or other ionic forms of P in plant residues (Mubarak et al 2002). Sun et al (2021) reported that the P release rate from animal manure-maize straw mixture increased gradually; showing a rapid release initially and slow in the latter stages of decomposition. Moreover, the increasing soil P availability with increase in incubation indicated that supply of P from residues remained higher than microbial demand throughout 60 days (Chimouriya et al 2018).

Effect of bacterial inoculants on exchangeable potassium content: The exchangeable potassium content of soil showed positive correlation with the weight loss of green manure. Potassium content increased up to the 14<sup>th</sup> day after inoculation (Fig. 3). Application of consortium culture significantly increased the potassium as compared to the control treatment. Potassium contained in green manure was also thought be the nutrient that evenly releases as the decomposition process proceeds and become available to plants. The extent and rate of K release from crop residues was usually considered greater than residue dry matter decomposition and N or P release (Lupwayi et al 2006). Talgre (2014) observed negative relation of potassium with the C/N ratio of organic matter, i.e., lower the C/N ratio higher release of potassium. Potassium being not a structural constituent of cell, so got easily solubilized by water in early stages of decomposition of organic residues, thus have



Fig. 3. Effect of different bacterial cultures on exchangeable potassium mineralization from green manure in a) in aerobic conditions, b) in flooding conditions

highest release rate among major nutrients (Mubarak et al 2002). Green manure (such as milk vetch) had been suggested as quick acting K fertilizer as it released 70-84 % of K within first 5 days of application (Zhou et al 2021). Kaur (2018) reported that increase in soil K content following green manuring might be due to the release of  $Fe^{2+}$  and  $Mn^{2+}$  under highly reduced conditions created during decomposition. Present results of higher soil available K in consortium inoculated treatments indicated that addition of effective microorganism can influence soil macronutrient content by improving the processes of nutrient mineralization (Wu et al. 2019).

In flooding conditions, the potassium content was low as compared to the aerobic conditions indicating the greater contribution to oxidative degradation reaction towards potassium release and availability. The maximum potassium content was with consortium application in unsterilized soil. Increase in soil solution K concentration might be due to release of Fe<sup>2+</sup> and Mn<sup>2+</sup> under highly reduced conditions created by green manure (Fig. 3). K release rate could also be highly conditioned by the precipitation during the decomposition period (Rodrı'quez-Lizana et al 2010). Lupwayi et al (2004) observed that intensive root decomposition and K release occurred from June to August with much rain received during these months and this effect was less prominent in above ground mass decomposition. Periera et al (2016) also observed that the rate of release of K could be correlated with the rate of decomposition of dry matter. Likewise, Gao et al (2015) used complex biological agent (fungi + bacteria) capable of increasing the total K content of the final corn stalk-swine waste compost compared to control. Wan et al (2020) reported that inoculation of 25

 Table 2. Effect of cellulose degrading bacteria on nitrate and ammoniacal nitrogen in sterilized and unsterilized soil at different time intervals during green manure decomposition

				Nitra	ate nitroge	n (mg/kg) a	t different	days						
	Under aerobic conditions							Under flooding conditions						
	2	8	16	30	45	60	2	8	16	30	45	60		
T1	0.01	0.38 <sup>J</sup>	6.52 <sup>J</sup>	9.57 <sup>J</sup>	11.43 <sup>J</sup>	16.88 <sup>J</sup>	0.01 <sup>⊬</sup>	0.98	0.90 <sup>H</sup>	0.89 <sup>G</sup>	0.91 <sup>⊬</sup>	0.93 <sup>H</sup>		
T2	0.22 <sup>F</sup>	4.44 <sup>∈</sup>	10.86 <sup>F</sup>	15.29 <sup>⊧</sup>	17.98 <sup>⊬</sup>	22.73 <sup>G</sup>	0.12 <sup>E</sup>	1.12 <sup>₌</sup>	1.08 <sup>E</sup>	1.07 <sup>E</sup>	1.11 <sup>⊧</sup>	1.13 <sup>′</sup>		
Т3	0.03 <sup>H</sup>	2.12 <sup>G</sup>	10.60 <sup>H</sup>	14.66 <sup>⊬</sup>	17.76	21.18 <sup>,</sup>	0.07 <sup>G</sup>	1.01 <sup>⊬</sup>	0.95 <sup>G</sup>	1.00 <sup>F</sup>	1.02 <sup>G</sup>	1.08 ⊧		
T4	0.04 <sup>G</sup>	1.54	10.62 <sup>G</sup>	15.19 <sup>°</sup>	18.00 <sup>G</sup>	22.60 <sup>H</sup>	0.09 <sup>F</sup>	1.07 <sup>G</sup>	1.02 <sup>⊧</sup>	1.01 <sup>F</sup>	1.06 ⊧	1.09 <sup>F</sup>		
T5	0.34 <sup>c</sup>	3.31 <sup>⊧</sup>	11.33 <sup>₌</sup>	15.51 <sup>₌</sup>	19.22 <sup>⊧</sup>	22.81 <sup>⊧</sup>	0.22 <sup>c</sup>	1.16 <sup>▷</sup>	1.10 <sup>D</sup>	1.09 <sup>□</sup>	1.13 <sup>□</sup>	1.15 <sup>⊳</sup>		
Т6	0.22 <sup>F</sup>	1.54 <sup>⊬</sup>	9.33'	14.02	19.35 <sup>⊧</sup>	24.62 <sup>⊧</sup>	0.13 <sup>⊧</sup>	1.10 <sup>⊧</sup>	1.02 ⊧	1.01 <sup>F</sup>	1.03 <sup>G</sup>	1.05 <sup>G</sup>		
T7	0.38 <sup>B</sup>	6.42 <sup>B</sup>	14.71 <sup>₿</sup>	19.45 <sup>₿</sup>	25.28 <sup>₿</sup>	28.69 <sup>B</sup>	0.24 <sup>B</sup>	1.24 <sup>₿</sup>	1.20 <sup>в</sup>	1.19 <sup>₿</sup>	1.23 <sup>₿</sup>	1.25 <sup>₿</sup>		
Т8	0.27 <sup>E</sup>	5.38 <sup>▷</sup>	13.80 <sup>c</sup>	18.70 <sup>c</sup>	23.63 <sup>c</sup>	<b>26</b> .51 <sup>□</sup>	0.19 <sup>⊳</sup>	1.13⁵	1.07 <sup>в</sup>	1.12 <sup>c</sup>	1.14 <sup>D</sup>	1.22 <sup>c</sup>		
Т9	0.29 <sup>D</sup>	5.82 <sup>c</sup>	13.15 <sup>⊳</sup>	18.54 <sup>▷</sup>	23.28 <sup>D</sup>	26.53 <sup>c</sup>	0.21 <sup>c</sup>	1.19 <sup>c</sup>	1.14 <sup>c</sup>	1.13 <sup>c</sup>	1.18 <sup>c</sup>	1.21 <sup>c</sup>		
T10	1.54 <sup>A</sup>	6.93 <sup>A</sup>	15.52 <sup>^</sup>	19.88 <sup>^</sup>	26.53 <sup>^</sup>	29.00 <sup>A</sup>	0.34	1.28 <sup>^</sup>	1.22 <sup>A</sup>	1.21 <sup>^</sup>	1.25 <sup>^</sup>	1.27 <sup>▲</sup>		
				Ammo	niacal nitro	aen (ma/ko	<ol> <li>at differe</li> </ol>	ent davs						

Ammoniacal nitrogen (mg/kg) at different days

	Under aerobic conditions							Under flooding conditions						
	2	8	16	30	45	60	2	8	16	30	45	60		
T1	5.8'	7.4 <sup>∺</sup>	11.4 <sup>J</sup>	16.4 <sup>J</sup>	13.5 <sup>J</sup>	13.2 <sup>J</sup>	5.3 ⊧	6.9 <sup>F</sup>	9.8 <sup>1</sup>	11.7 <sup>⊬</sup>	11.1 <sup>H</sup>	10.9 <sup>G</sup>		
T2	6.5 <sup>⊧</sup>	8.8 <sup>F</sup>	13.6 <sup>⊧</sup>	18.4 <sup>'</sup>	16.3	16.1 <sup>1</sup>	5.6 <sup>EF</sup>	7.3 <sup>BCD</sup>	11.7 <sup>⊧</sup>	14.0 <sup>⊧</sup>	13.1 <sup>₌</sup>	12.8 <sup>₌</sup>		
Т3	6.1 <sup>⊬</sup>	8.0 <sup>G</sup>	12.8 <sup>G</sup>	19.2 <sup>⊧</sup>	16.8 <sup>G</sup>	16.5 <sup>°</sup>	5.3 <sup>⊧</sup>	7.2 <sup>CDF</sup>	11.4 <sup>EF</sup>	14.5⁵	12.5 <sup>⊧</sup>	12.5 <sup>⊧</sup>		
T4	6.1 <sup>⊬</sup>	7.8 <sup>G</sup>	12.3 <sup>'</sup>	18.8 <sup>⊬</sup>	16.5 <sup>∺</sup>	16.1 <sup>⊬</sup>	5.4⁵	7.0 <sup>DF</sup>	11.1 <sup>FG</sup>	13.4 <sup>G</sup>	12.1 <sup>G</sup>	11.7 <sup>⊧</sup>		
T5	6.7 <sup>E</sup>	9.1 <sup>⊧</sup>	14.1 <sup>⊧</sup>	21.2 <sup>D</sup>	17.6 <sup>⊧</sup>	16.9 <sup>⊧</sup>	5.8 <sup>de</sup>	7.5 <sup>BC</sup>	12.6 <sup>⊳</sup>	17.5 <sup>c</sup>	14.0 <sup>D</sup>	13.8 <sup>⊳</sup>		
Т6	6.4 <sup>G</sup>	7.8 <sup>⊬</sup>	12.7 <sup>⊦</sup>	18.9 <sup>G</sup>	17.4 <sup>⊧</sup>	17.4 <sup>⊧</sup>	6.0 <sup>CD</sup>	7.5 <sup>BC</sup>	11.0 <sup>G</sup>	17.1 <sup>⊳</sup>	15.4 <sup>c</sup>	15.8 <sup>c</sup>		
Т7	7.4 <sup>₿</sup>	10.0 <sup>D</sup>	18.7 <sup>D</sup>	21.0 <sup>⊧</sup>	19.6 <sup>D</sup>	19.2 <sup>⊳</sup>	6.5 <sup>AB</sup>	9.1 <sup>^</sup>	13.8 <sup>₿</sup>	18.6 <sup>AB</sup>	17.3 <sup>AB</sup>	17.8 <sup>^</sup>		
Т8	7.0 <sup>D</sup>	10.4 <sup>c</sup>	20.7 <sup>c</sup>	23.0 <sup>c</sup>	20.6 <sup>c</sup>	20.3 <sup>c</sup>	6.2 <sup>BC</sup>	8.6 <sup>B</sup>	13.1 <sup>c</sup>	18.3 <sup>₿</sup>	17.1 <sup>₿</sup>	16.8 <sup>₿</sup>		
Т9	7.3 <sup>c</sup>	11.0 <sup>₿</sup>	21.3 <sup>₿</sup>	23.2 <sup>B</sup>	21.0 <sup>B</sup>	21.1 <sup>₿</sup>	6.3 <sup>BC</sup>	9.1 <sup>^</sup>	13.6 <sup>в</sup>	18.8 <sup>A</sup>	17.4 <sup>AB</sup>	17.7 <sup>^</sup>		
T10	7.8 <sup>₿</sup>	11.2 <sup>A</sup>	23.7 <sup>A</sup>	24.6 <sup>A</sup>	22.4 <sup>A</sup>	22.0 <sup>A</sup>	6.7 <sup>A</sup>	8.8 <sup>A</sup>	15.3 <sup>A</sup>	20.8 <sup>A</sup>	17.6 <sup>^</sup>	18.0 <sup>A</sup>		

\*\*Different letter shows significant differences at p=0.05 using post hoc text Tukey's B within treatments
different cellulosic isolates improved co-decomposition of maize straw and chicken manure, resulting in higher nutrient content (NPK) and GI index compared to control.

#### CONCLUSION

Decomposition and nutrient mineralization from green manure can be improved with the application of microbial inoculants. The application of consortium significantly improved the decomposition and nutrient release in aerobic condition. Decomposed the green manure 7 days earlier than uninoculated control, thus have potential to decrease the fallow period between green manure incorporation and crop sowing (particularly maize). In this aspect, microbial consortium is more effective than individual culture and can be integrated with green manure practices to enhance soil nutrient availability as well as it can minimize ecological imbalance by maintaining nutrient flows from one system to another.

## REFERENCES

- Awasthi MK, Wang Q, Wang MJ, Chen HY, Ren XN, Zhao JC and Zhang ZQ 2018. In-vessel co-composting of food waste employing enriched bacterial consortium. *Food Technology and Biotechnology* **56**: 83-89.
- Awasthi MK, Duan YM, Awasthi SK, Liu T and Zhang ZQ 2020. Effect of biochar and bacterial inoculum additions on cow dung composting. *Bioresource Technology* **297**: 122407.
- Berg B and R Laskowski 2006. Litter decomposition: A guide to carbon and nutrient turnover. Advances in Ecological Research 38: 1-421.
- Bhattacharjya S, Sahu A, Phalke DH, Manna MC, Thakur JK, Mandal A, Tripathi AK, Sheoran P, Choudhary M, Bhowmick A, Rahman MM, Naidu R and Patra AK 2021. *In situ* decomposition of crop residues using lignocellulolytic microbial consortia: A viable alternative to residue burning. *Environment Science and Pollution Research* 28: 32416-32433.
- Chi CP, Chu S, Wang B, Zhang D, Zhi Y, Yang X and Zhou P 2020. Dynamic bacterial assembly driven by *Streptomyces griseorubens* JSD-1 inoculants correspond to composting performance in swine manure and rice straw co-composting. *Bioresource Technology* **313**: 123692.
- Chimouriya S, Lamichhane J, Gauchan DP and Dhulikhel K 2018. Green manure for restoring and improving the soil nutrients quality. *International Journal of Research* **5**: 1064-1074.
- Duan R, Lin Y, Zhang J, Huang M, Du Y, Yang L, Bai J, Xiang G, Wang Z and Zhang Y 2021. Changes in diversity and composition of rhizosphere bacterial community during natural restoration stages in antimony mine. *PubMedline* **14**(9): e12302.
- Gao H, Zhou C, Wang R and Li X 2015. Comparison and evaluation of co-composting corn stalk or rice husk with swine waste in China. *Waste Biomass Valor* **6**: 699-710.
- Greff B, Szigeti J, Nagy A, Lakatos E and ´o´ Varga L 2022. Influence of microbial inoculants on co-composting of lignocellulosic crop residues with farm animal manure: A review. *Journal of Environment Management* **302**: 114088.
- Hefting M, Clement JC, Dowrick D, Cosandey AC, Bernal S, Cimpian C, Tatur A, Burt TP and G 2004. Water table elevation controls on soil nitrogen cycling in riparian wetlands along a European climatic gradient. *Biogeochemistry* **67**(1): 113-134.
- Jackson ML 1967. Soil chemical analysis. Precentice Hall of India Pvt. New Delhi.
- Kalita J, Bhattacharyya JHC, Thakuria RK, Bhattacharyya D and

DAS K 2022. Evaluation of suitable rice (*Oryza sativa*) based cropping systems under rainfed medium land situation of Assam for enhancing farmers income and soil health. *Crop Research* **57**: 1-7.

- Kaur J 2018. Impact of long term organic and inorganic fertilization on soil microbial activities and screening of nitrogen fixing methanotrophic bacteria. Ph.D. dissertation. Punjab Agricultural University, Ludhiana, India.
- Kumar S, Samiksha and Sukul P 2020. Green manuring and its role in soil health management. *Soil Health* **2020**: 219-241.
- Lei B, Wang J and Yao H 2022. Ecological and environmental benefits of planting green manure in paddy fields. *Agriculture* **12**: 223.
- Lupwayi NZ, Clayton GW, O'Donovan JT, Harker KN, Turkington TK and Rice WA 2004. Decomposition of crop residues under conventional and zero tillage. *Canadian Journal of Soil Science* **84**: 403-410.
- Lupwayi NZ, Clayton GW, O'Donovan JT, Harker KN, Turkington TK and Soon Y 2006. Potassium release during decomposition of crop residues under conventional and zero tillage. *Canadian Journal of Soil Science* **86**: 473-481.
- Manu MK, Kumar R and Garg A 2019. Decentralized composting of household wet biodegradable waste in plastic drums: Effect of waste turning, microbial inoculum and bulking agent on product quality. *Journal of Cleaner Production* **226**: 233-241.
- Mohanty M, Reddy KS, Probert ME, Dalal RC, Rao AS and Menzies NW 2011. Modelling N mineralization from green manure and farmyard manure from a laboratory incubation study. *Ecological Modelling* 222: 719-726.
- Mubarak AR, Rosenani AB, Anuar AR and Zauyah S 2002. Decomposition and nutrient release of maize stover and groundnut haulm under tropical field conditions of Malaysia. *Communication in Soil Science and Plant* **33**: 609-622.
- Olsen SR, CV Cole, FS Watanabe and LA Dean 1954. Estimation of available phosphorous by extraction with sodium bicarbonate. U.S.D.A. Circular 939-19. Peterson (eds.), Soil Biochemistry, Vol. I. Marcel & Dekker, New York.
- Page AL, Miller RH and Koeney DR 1982. Method of analysis, part 2, Chemical and Microbiological properties. Agronomy monograph No.9 American Society of Agronomy Inc: Soil Science Society of America; Inc; Madison Wisconsin, USA.
- Pereira NS, Soares I and Miranda F Rodrigues De 2016. Decomposition and nutrient release of leguminous green manure species in the Jaguaribe-Apodi region, Ceará, Brazil. Cienc. Rural, Santa Maria, **46**(6): 970-975.
- Resende AS, Xavier RP, Quesada DM, Coelho CHM, Boddey RM, Alves BJR, Guerra JGM and Urquiaga S 2000. *Incorporação de leguminosas para fins de adubação verde em pré plantio de cana-de-açúcar*. Seropédica, Embrapa Agrobiologia, 2000. 18p. (Documentos, 124).
- Rodrı'guez-Lizana A, Carbonell P, Gonza'lez R and Ordo'nez R 2010. N, P and K released by the field decomposition of residues of a pea–wheat–sunflower rotation. *Nutrient Cycling* and Agroecosystem 87: 199-208.
- Silva GTA, Matos LV, Nóbrega PO, Campello EFC and Resende AS 2008. Chemical composition and decomposition rate of plants used as green manure. *Science Agriculture* (Piracicaba, Braz.), 65(3): 298-305.
- Sun L, Sun Z, Hu J, Yaa OK and Wu J 2021. Decomposition characteristics, nutrient release, and structural changes of maize straw in dryland farming under combined application of animal manure. *Sustainability* **13**: 7609.
- Talgre L, Lauringson E, Roostalu H and Makke A 2014. Phosphorus and potassium release during decomposition of roots and shoots of green manure crops. *Biological Agriculture and Horticulture* **30**(4): 264-271.
- Trinsoutrot I, Recous S, Bentz B, Lineres M, Cheneby D and Nicolardot B 2000. Biochemicals qualities of crop residue and carbon and nitrogen minerlization kinetic under nonlimiting

nitrogen conditions. *Soil Science Society of America Journal* **64**(3): 918-926

- Umsakul K, Dissara Y and Srimuang N 2010. Chemical physical and microbiological changes during composting of the water hyacinth. *Pakistan Journal of Biological Sciences* **13**: 985-992
- Unger IM, Motavalli PP and Muzika RM 2009. Changes in soil chemical properties with flooding: A field laboratory approach. *Agriculture, Ecosystems and Environment* **131**(1–2):105-110.
- Vogel HJ, Eberhardt E, Franko U, Lang B, Lie BM, Weller U, Wiesmeier M and Wollschläger U 2019. Quantitative evaluation of soil functions: Potential and state. *Frontiers in Environmental Science* **7**: 164.
- Wan B, Wang XT, Cong C, Li JB, Xu YP, Li XY, Hou FQ, Wu YY and Wang LL 2020. Effect of inoculating microorganisms in chicken manure composting with maize straw. *Bioresource Technology* 301: 122730.

Received 22 June, 2024; Accepted 01 September, 2024

- Wu YP, Chen YX, Shaaban M, Zhu DW, Hu CX, Chen ZB and Wang Y 2019. Evaluation of microbial inoculants pretreatment in straw and manure co-composting process enhancement. *Journal of Cleaner Production* **239**: 118078.
- Xu Z, Lei P, Rui Zhai R, Wen Z and Jin M 2019. Recent advances in lignin valorization with bacterial cultures: Microorganisms, metabolic pathways, and bio-products. *Biotechnology and Biofuels* **12**: 32.
- Zhao X, Yuan G, Wang H, Lu D, Chen X and Zhou J 2019. Effects of full straw incorporation on soil fertility and crop yield in rice-wheat rotation for silty clay loamy cropland. *Agronomy* **9**: 133.
- Zhou G, Chang D, Gao S, Liang T, Liu R and Cao W 2021. Coincorporating leguminous green manure and rice straw drives the synergistic release of carbon and nitrogen, increases hydrolase activities, and changes the composition of main microbial groups. *Biology of Fertile Soils* **57**: 547-561.



# Productivity and Biological Efficiency Indices of Sesamum + Cowpea Intercropping System in Response to Row Ratio and Nutrient Management

Sabitha B., Shalini Pillai P.<sup>1</sup>, Usha C. Thomas<sup>2</sup>, Sheeja K. Raj<sup>3</sup> and Chitra N.<sup>4</sup>

Department of Agronomy, <sup>1</sup>Department of Agronomy, <sup>2</sup>Instructional Farm, <sup>3</sup>Department of Organic Agriculture, <sup>4</sup>Department of Agricultural Microbiology, College of Agriculture, Vellayani, Thiruvananthapuram-695 522, India E-mail: bsabitha1998@gmail.com

**Abstract:** Field experiment on sesamum + cowpea intercropping system was carried out to assess the yield advantage of the system using biological indices, *viz.*, land equivalent ratio (LER), relative crowding coefficient (RCC), aggressivity (A), competition index (CI), competition ratio (CR), sesamum equivalent yield (SEY) and percentage yield difference (PYD). The study was conducted at College of Agriculture, Vellayani, Thiruvananthapuram during the summer season of 2023. The experiment treatments comprised of combinations of three factors, *viz.*, three levels of nitrogen (N) (n<sub>1</sub>- 100 % RDN; n<sub>2</sub>- 75 % RDN; n<sub>3</sub>- 50 % RDN), two row ratios (R) (r<sub>1</sub>- 4:2; r<sub>2</sub>- 6:3) and application of AMF (A) (a<sub>1</sub>- without AMF; a<sub>2</sub>- with AMF). The seed yield of sesamum and cowpea were significantly influenced by individual and interaction effects of treatments. Irrespective of the treatments applied all the intercropping systems showed advantage in terms of LER, CI and PYD. The component crop cowpea was observed with positive (+) aggressivity and higher (>1) CR indicates the dominance of cowpea over sesamum. The intercropping system with the treatment combination 100 % RDN at 6:3 row ratio without AMF (n<sub>1</sub>,r<sub>2</sub>a<sub>1</sub>) resulted in higher LER (2.40), RCC (66.18), SEY (1801), and PYD (139.87%).

Keywords: Sesamum intercropping, Biological indices, Productivity, Row ratio, AMF, levels of nitrogen

Sesamum stands as a traditionally significant and culturally influential crop with widespread impact on agriculture and nutrition. However, sesamum has been recognized as an exhaustive crop based on its ability to utilize and deplete soil nutrients at an accelerated rate, if not managed properly can lead to decline in soil fertility. Sesamum, being a crop highly susceptible to waterlogging, unprecedented rainfall is always a risk for sole cropping. Intercropping involving compatible crops is regarded as a risk alleviator with sustained yield advantage compared to sole cropping (Aulakh et al 2019). In general, pulses are perfect candidates for intercropping due to their short duration and relatively lower sensitivity to light and temperature. Further, they fix atmospheric nitrogen and also share a portion of the fixed nitrogen to the intercropped oilseeds (Jiwan et al 2021). Cowpea (Vigna unguiculata (L.) Walp) is one of pulses with high adaptability and consumer preference in Kerala. Several studies have shown cowpea to be a suitable crop for intercropping. However, cowpea is an aggressive legume and its performance in combination with sesamum has yielded contrasting results. Guedes et al (2010) reporting yield reduction of cowpea and Bhatti et al (2005) reporting reduction in yield of sesamum. Crop density and row configuration are important agronomic considerations for attaining the objectives of intercropping (Afe 2017). Both additive series and replacement series have been tested in several intercropping systems. With increasing demand for more sustainable crop production practices, supplying more nitrogen through nitrogen-fixation than by using synthetic chemical fertilizers has become a lucrative choice (Fustec et al 2010). Interplant nitrogen transfer, though a bi-directional process, nitrogen tends to get transferred from nitrogenfixers to non-nitrogen fixers (Carlsson and Huss-Danell 2014). Arbuscular mycorrhizal fungi (AMF) are obligate symbionts that transport considerable amount of nutrients and water through the hyphal network to the associating crop (Chen et al 2018). This study therefore concentrates on the individual and interactive effects of various factors, *viz.,* nitrogen, row ratios, and AMF, on the productivity and biological efficiency of sesamum + cowpea intercropping system.

#### MATERIAL AND METHODS

The experiment conducted at, College of Agriculture, Vellayani, Thiruvananthapuram, Kerala, India located at 8°25'41.3 N latitude and 76°59'16.7 E longitude, at an altitude of 29 m above mean sea level, during February to May 2023. The soil of the experimental field was sandy clay loam with a moderate acidic pH (5.8), moderate level of organic carbon (0.81%), low status of nitrogen (250.88 kg/ha), high availability of phosphorus (49.68 kg/ha), and medium in potassium (253.57 kg/ha) status. The sesamum variety

Thilak (ACV-3) released from College of Agriculture, Vellayani, Thiruvananthapuram and cowpea variety PGCP -6 (Pant lobia - 3) released from G. B. pant University of Agriculture and Technology, Pantnagar were used for the study. The experiment was laid out in randomised block design with 3 x 2 x 2 treatments replicated thrice. The three factors were, levels of nitrogen (n<sub>1</sub>- 100 % recommended dose of nitrogen (RDN); n<sub>2</sub>- 75 % RDN; n<sub>3</sub>- 50 % RDN), row ratios (r<sub>1</sub>- 4:2; r<sub>2</sub>- 6:3) and application of arbuscular mycorrhizal fungi (AMF) (a<sub>1</sub>- without AMF; a<sub>2</sub>- with AMF). All cultural practices followed as per the KAU package of practices recommendations (KAU 2016). Sole crops of sesamum and cowpea were raised for computing intercropping indices. The sesamum was harvested when the seeds of tenth capsule turned black and cowpea was harvested when 70 to 80 % of the pods turned brown with hard seeds inside. The sesamum and cowpea were harvested separately from the net plot area and the seeds were sun-dried and weight was expressed as seed yield in kg/ha. Biological indices were computed using yield data obtained from the intercropping system and sole crops.

**Seed yield:** Net plot area was harvested and the seeds were sun-dried and weight was expressed as seed yield in kg/ha.

Land equivalent ratio (LER): Land equivalent ratio was calculated as per the formula suggested by Willey (1979).

$$LER = L_s + L_c = \frac{Y_{sc}}{Y_{ss}} + \frac{Y_{cc}}{Y_{cs}}$$

where,  $L_{\rm s},~L_{\rm c}\text{-}$  LER for sesamum and cowpea,  $Y_{\rm sc}\text{-}$  intercrop yield of sesamum

 $Y_{\mbox{\tiny cs}\mbox{-}}$  - intercrop yield of cowpea,  $Y_{\mbox{\tiny ss}}$  - sole crop yield of sesamum

#### Y<sub>cc</sub>- sole crop yield of cowpea

**Relative crowding coefficient (RCC):** Relative crowding coefficient denoted as 'K' was computed (de Wit 1960).

$$K_{sc} = \frac{Y_{sc} \times Z_{cs}}{(Y_{ss} - Y_{sc})Z_{sc}}$$
$$K_{cs} = \frac{Y_{cs} \times Z_{sc}}{(Y_{cc} - Y_{cs})Z_{cs}}$$
$$K = K_{sc} \times K_{cs}$$

where,  $K_{sc}$  and  $K_{cs}$ - relative crowding coefficient of sesamum intercropped with cowpea and sesamum,  $Y_{ss}$ - sole crop yield of sesamum,  $Y_{sc}$ - intercrop yield of sesamum,  $Y_{cc}$ - sole crop yield of cowpea,  $Y_{cs}$ - intercrop yield of cowpea

 $Z_{\rm sc}\mathchar`-$  sown proportion of sesamum,  $Z_{\rm cs}\mathchar`-$  sown proportion of cowpea

**Aggressivity (A):** Aggressivity was computed by using the formula proposed by McGilchrist (1965).

$$A_{sc} = \frac{Y_{sc}}{Y_{ss} \times Z_{sc}} - \frac{Y_{cs}}{Y_{cc} \times Z_{cs}} \qquad A_{cs} = \frac{Y_{cs}}{Y_{cc} \times Z_{cs}} - \frac{Y_{sc}}{Y_{ss} \times Z_{sc}}$$

where,  $A_{sc}$ - aggressivity of sesamum,  $A_{cs}$ - aggressivity of cowpea,  $Y_{ss}$ - sole crop yield of sesamum,  $Y_{cs}$ - intercrop yield of cowpea,  $Y_{sc}$ - sole crop yield of cowpea,  $Y_{sc}$ - intercrop yield of sesamum,  $Z_{sc}$ - sown proportion of sesamum,  $Z_{cs}$ - sown proportion of cowpea

**Competition index (CI):** Competition index was computed as suggested by Donald (1963).

$$CI = \frac{(Y_{ss} - Y_{sc}) \times (Y_{cc} - Y_{cs})}{Y_{ss} \times Y_{cc}}$$

where,  $Y_{\rm ss}\text{-}\,$  sole crop yield of sesamum,  $Y_{\rm sc}\text{-}\,$  intercrop yield of sesamum

 $Y_{\infty}$ - sole crop yield of cowpea,  $Y_{\infty}$ - intercrop yield of cowpea **Competition ratio (CR):** Competition ratio was calculated (Willey et al 1980).

$$\begin{aligned} CR_s &= \frac{Y_{sc}}{Y_{ss} \times Z_{sc}} \div \frac{Y_{cs}}{Y_{cc} \times Z_{cs}} \\ CR_c &= \frac{Y_{cs}}{Y_{cc} \times Z_{cs}} \div \frac{Y_{sc}}{Y_{ss} \times Z_{sc}} \end{aligned}$$

where,

CR<sub>s</sub>CR<sub>c</sub> - competition ratio of sesamum and cowpea

 $Y_{\mbox{\tiny ss}\mbox{-}}$  sole crop yield of sesamum,  $Y_{\mbox{\tiny sc}\mbox{-}}$  intercrop yield of sesamum

 $Y_{\rm cc}\text{-}$  sole crop yield of cowpea,  $Y_{\rm cs}\text{-}$  intercrop yield of cowpea  $Z_{\rm sc}\text{-}$  sown proportion of sesamum,  $Z_{\rm cs}\text{-}$  sown proportion of cowpea

**Sesamum equivalent yield (SEY):** Sesamum equivalent yield was computed based on the seed yield of the intercropped cowpea and prevailing market price of sesamum and cowpea, based on the crop equivalent yield (Lal and Ray 1976. Verma and Modgal 1983).

$$SEY = Y_{sc} + Y_{cs} \times \left\{\frac{P_c}{P_s}\right\}$$

where,

 $Y_{\rm sc}\text{-}$  intercrop yield of sesamum,  $Y_{\rm cs}\text{-}$  intercrop yield of cowpea

P<sub>c</sub>- market price of cowpea, P<sub>s</sub>- market price of sesamum

**Percentage yield difference (PYD):** Percentage yield difference was computed (Afe and Atanda 2015).

$$PYD = 100 - \left[ \left\{ \frac{Y_{SS} - Y_{SC}}{Y_{SS}} + \frac{Y_{CC} - Y_{CS}}{Y_{CC}} \right\} \right] \times \frac{100}{1}$$

where,

 $Y_{\rm ss}\text{-}$  sole crop of sesamum,  $Y_{\rm cc}\text{-}$  sole crop of cowpea  $Y_{\rm sc}\text{-}$  intercrop yield of sesamum,  $Y_{\rm cs}$  - intercrop yield of cowpea

#### **RESULTS AND DISCUSSION**

**Seed yield:** Seed yield of both sesamum and cowpea were significantly influenced by individual and interaction effects (Tables 1, 2 3 ). The highest seed yield (1193 kg/ha) of sesamum was observed with 75 % RDN ( $n_2$ ), whereas, the

**Table 1.** Effect of levels of nitrogen, row ratios and AMF on
 seed yield of sesamum and cowpea

Table 3. Effect of N x R x A interactions on seed yield of sesamum and cowpea

Treatment	Seed yield (kg/		
	Sesamum	Cowpea	
Levels of nitrogen (N)			
$n_{_1}\!-100$ % recommended dose of nitrogen	947 <sup>b</sup>	1647ª	
$n_{_2}\!-75$ % recommended dose of nitrogen	1193°	1292⁵	
$\rm n_{_3}-50$ % recommended dose of nitrogen	802°	1170 <sup>⊳</sup>	
CD (p=0.05)	97.6	257.3	
Row ratios (R)			
r <sub>1</sub> - 4:2	953	1355	
$r_2 - 6:3$	1009	1384	
CD (p=0.05)	NS	NS	
AMF (A)			
a <sub>1</sub> – without AMF	994	1292	
a <sub>2</sub> – with AMF	967	1447	
CD (p=0.05)	NS	NS	

AMF- Arbuscular mycorrhizal fungi; NS - Not significant

Table 2. Effect of N x R, R x A and N x A interactions on se	ed
vield of sesamum and cowpea	

Treatment	Seed yield (kg/ha)	
	Sesamum	Cowpea
Levels of nitrogen (N) x Row ratios (R)		
n,r,	814 <sup>⊳</sup>	1442 <sup>⊳</sup>
n <sub>1</sub> r <sub>2</sub>	1081°	1852°
n <sub>2</sub> r <sub>1</sub>	1216ª	1415⁵
n <sub>2</sub> r <sub>2</sub>	1171 <sup>ª</sup>	1168⁵
n <sub>3</sub> r <sub>1</sub>	829 <sup>b</sup>	1207 <sup>⊳</sup>
n <sub>3</sub> r <sub>2</sub>	775⁵	1132 <sup>⊳</sup>
CD (p=0.05)	138.1	363.9
Row ratios (R) x AMF (A)		
r <sub>1</sub> a1	892°	1220
r <sub>1</sub> a <sub>2</sub>	1014 <sup>ab</sup>	1489
r <sub>2</sub> a <sub>1</sub>	1097ª	1364
r <sub>2</sub> a <sub>2</sub>	921 <sup>bc</sup>	1404
CD (p=0.05)	112.7	NS
Levels of nitrogen (N) x AMF (A)		
<b>n</b> <sub>1</sub> <b>a</b> <sub>1</sub>	970 <sup>bc</sup>	1602
n <sub>1</sub> a <sub>2</sub>	924°	1692
$n_2 a_1$	1311°	1145
$n_2 a_2$	1076 <sup>⊳</sup>	1439
<b>n</b> <sub>3</sub> <b>a</b> <sub>1</sub>	701 <sup>d</sup>	1129
<b>n</b> <sub>3</sub> <b>a</b> <sub>2</sub>	902°	1210
CD (p=0.05)	137.968	NS

See Table 1 for treatment details

Treatment	Seed yield (kg/ha)	Seed yield (kg/ha)					
	Sesamum	Cowpea					
Levels of nitrogen (N) x Row ratios (R) x AMF (A)							
n <sub>1</sub> r <sub>1</sub> a <sub>1</sub>	729	1378					
n <sub>1</sub> r <sub>1</sub> a <sub>2</sub>	898	1507					
$n_1r_2a_1$	1211	1827					
$n_1r_2a_2$	950	1877					
n <sub>2</sub> r <sub>1</sub> a <sub>1</sub>	1240	1173					
n <sub>2</sub> r <sub>1</sub> a <sub>2</sub>	1192	1658					
$n_2 r_2 a_1$	1382	1116					
n <sub>2</sub> r <sub>2</sub> a <sub>2</sub>	959	1221					
n <sub>3</sub> r <sub>1</sub> a <sub>1</sub>	706	1111					
n <sub>3</sub> r <sub>1</sub> a <sub>2</sub>	952	1304					
n <sub>3</sub> r <sub>2</sub> a <sub>1</sub>	696	1148					
n <sub>3</sub> r <sub>2</sub> a <sub>2</sub>	853	1116					
CD (p=0.05)	NS	NS					
Sole crop of sesamum without AMF	1138	1369					
Sole crop of sesamum with AMF	1218	1649					

treatment n<sub>1</sub> (100 % RDN) resulted in highest seed yield (1647 kg/ha) of cowpea. Considering the interaction effects,  $n_2r_1$ ,  $r_2a_1$ , and  $n_2a_1$  observed to have significantly higher seed yield (1216 kg/ha, 1097 kg/ha, and 1076 kg/ha respectively) of sesamum. The highest seed yield (1852 kg/ha) in cowpea was observed with the treatment n<sub>1</sub>r<sub>2</sub>(100 % RDN at 6:3 row ratio). The 75 % RDN improved the yield of sesamum instead of growth. The combination of 75 % RDN with 6:3 row ratio, and without AMF application was favourable for increased yield in sesamum. Cowpea, as a high nitrogen requiring crop, it might have also used part of nitrogen that supplied for sesamum under full dose recommendation. The treatment, n<sub>1</sub>r<sub>2</sub> (100 % RDN at 6:3 row ratio) proved its significance on seed yield of cowpea.

Land equivalent ratio: Irrespective of the treatments, all the sesamum + cowpea system resulted in LER of >1 indicates the yield advantage of the system, hence cowpea can be regarded as better component crop for sesamum. Among the treatment combinations, 100 % RDN at 6:3 row ratio without AMF  $(n_1r_2a_1)$  resulted in higher LER (2.40) followed by  $n_2r_2a_1$ (2.03) (Table 4). Kotadiya et al (2023) reported the yield advantage of sesamum - pulse intercropping system in terms of LER.

Relative crowding coefficient: The 'K' value for sesamum, cowpea and the intercropping system varied among treatments (Table 4). The higher K value for sesamum (2.92) and cowpea (21.14) observed with  $n_2r_1a_2$  and  $n_1r_1a_2$  respectively, whereas RCC of the sesamum + cowpea intercropping system was observed to be higher (66.18) with 100 % RDN at 6:3 row ratio without AMF ( $n_1r_2a_1$ ). When comparing to sesamum, cowpea was higher K value. The similar results were reported by Bhatti et al (2006). Aggressivity, RCC and CR revealed superior competitiveness of cowpea over sesamum when compared to other leguminous intercrops tested. This showed that when planted with sesamum, cowpea outperformed all other intercrops in terms of competitiveness by effectively utilizing the available resources.

the aggressivity value (Table 4) of cowpea was positive (+) and that of sesamum negative (-) indicating the dominant behavior of cowpea over sesamum. The treatment,  $n_1r_2a_2$ recorded higher (2.40) value of aggressivity for cowpea followed by  $n_3r_2a_1$  (2.25) .The findings on aggressivity of cowpea are akin to the results of Bindhu et al (2014) and Bhatti et al (2006). Bhatti et al (2005) reported yield reduction in sesamum due to the luxuriant vegetative development of cowpea compared to other pulse crops.

**Competition index:** The competition index for all the treatment combinations were observed to lesser than unity (Table 5), indicating yield advantage of the sesamum + cowpea intercropping system. This indicates that, even if

Aggressivity: Irrespective of the treatment combinations,

Table 4. Effect of intercropping on land equivalent ratio (LER), relative crowding coefficient (K) and aggressivity (A)

neaunent	LER	Relative crowding coemclent			Aggressivity	
	_	K <sub>sc</sub>	K <sub>cs</sub>	К	A <sub>sc</sub>	A <sub>cs</sub>
n <sub>1</sub> r <sub>1</sub> a <sub>1</sub>	1.65	0.89	-324.12	-288.85	-1.64	+1.64
$n_1r_1a_2$	1.65	1.40	21.14	29.67	-1.52	+1.52
$n_1r_2a_1$	2.40	-8.29	-7.98	66.18	-2.21	+2.21
$n_1r_2a_2$	1.92	1.77	-16.50	-29.24	-2.40	+2.40
$n_2 r_1 a_1$	1.95	-6.08	11.97	-72.76	-1.19	+1.19
$n_{2}r_{1}a_{2}$	1.98	2.92	-190.00	-554.80	-1.54	+1.54
$n_2 r_2 a_1$	2.03	-2.83	-7.74	21.91	-0.34	+0.34
$n_2 r_2 a_2$	1.53	1.85	5.70	10.55	-0.57	+0.57
n <sub>3</sub> r <sub>1</sub> a <sub>1</sub>	1.43	0.82	8.59	7.02	-0.99	+0.99
$n_3 r_1 a_2$	1.57	1.79	7.55	13.50	-0.80	+0.80
n <sub>3</sub> r <sub>2</sub> a <sub>1</sub>	1.45	0.79	10.39	8.19	-2.25	+2.25
$n_3r_2a_2$	1.38	1.17	4.19	4.88	-0.83	+0.83

See Table 1 for treatment details; K<sub>sc</sub>, A<sub>sc</sub> and K<sub>ss</sub>, A<sub>sc</sub> – relative crowding coefficient and aggressivity of sesamum in combination with cowpea respectively and cowpea in combination with sesamum respectively

Table 5. Effect of intercropping	on competition index (CI) و	, competition ratio (CR)	), sesamum equivalent yield	(SEY), percentage
yield difference (PYD	)			

Treatment	tment CI Competition ratio		tion ratio	SEY (kg/ha)	PYD (%)
		CR <sub>s</sub>	CR <sub>c</sub>	_	
n₁r₁a₁	0.05	0.32	3.12	1133	64.68
$n_1r_1a_2$	0.03	0.37	2.71	1389	70.27
$n_1 r_2 a_1$	0.02	0.53	1.88	1801	139.87
$n_1r_2a_2$	-0.04	0.39	2.56	1617	97.28
$n_2 r_1 a_1$	-0.01	0.54	1.84	1679	94.65
$n_2 r_1 a_2$	0.00	0.49	2.04	1753	105.26
$n_2 r_2 a_1$	-0.06	0.61	1.65	1717	102.96
$n_2 r_2 a_2$	0.09	0.39	2.54	1286	58.29
n <sub>3</sub> r <sub>1</sub> a <sub>1</sub>	0.14	0.31	3.22	1004	43.16
$n_3r_1a_2$	0.07	0.39	2.56	1321	62.70
$n_3r_2a_1$	-0.02	0.31	3.27	1188	45.06
$n_3r_2a_2$	0.11	0.35	2.86	1203	42.59

See Table 1 for treatment details

CR<sub>s</sub> – competition ratio of sesamum; CR<sub>c</sub> – competition ratio of cowpea

there exists dominance of cowpea over sesamum, over all it provides a favorable environment for higher productivity of the intercropping system. Kumar et al (2021) concluded that, short duration grain legumes with sesamum in an intercropping system could be advantageous, resulting in a synergistic relationship where both crops benefit from shared resources, further enhanced the overall productivity of the system.

**Competition ratio:** Cowpea recorded higher CR value than sesamum in all treatment combinations (Table 5). The  $n_3r_2a_1$  (50 % at 6:3 row ratio without AMF) recorded higher CR for cowpea (3.27) followed by  $n_3r_1a_1$  (3.22). The higher CR value indicates more competition between sesamum and cowpea. Superiority of cowpea in the intercropping systems was reported by Bhatti et al (2006).

**Sesamum equivalent yield:** The  $n_1r_2a_1$  (100% RDN at 6:3 row ratio without AMF) resulted in significantly higher (1801 kg/ha) SEY (Table 5) followed by  $n_1r_1a_2$  and  $n_2r_2a_1$ . This could be solely attributed to the higher overall seed yield (Table 3) observed in these treatment combinations. The combined individual and interactive effects of the treatments played a significant role in enhancing the yield attributes of both sesamum and cowpea ultimately leading to a comparatively higher total seed yield for the sesamum + cowpea intercropping system, hence the higher SEY.

**Percentage yield difference:** Percentage increase in yield was higher (139.87 %) for  $n_1r_2a_1$  (100 % RDN at 6:3 row ratio without AMF) followed by  $n_2r_1a_2$  and  $n_2r_2a_1$ . The percentage yield difference was the least (42.59 %) in  $n_3r_2a_2$  (50 % RDN at 6:3 row ratio with AMF). The yield reduction of sesamum was compensated by increase in the yield of cowpea. Percentage yield difference can be taken as a parameter to interpret the efficiency of intercropping system as it follows the trend of LER.

#### CONCLUSION

This study reveals the potential of growing sesamum and cowpea together as an intercropping system, showing improved productivity and biological efficiency. The intercropping system outperforms individual crops in terms of overall productivity and resource use efficiency, regardless of the specific treatment conditions. Among the various treatment combinations, the sesamum + cowpea intercropping system planted at 6:3 row ratio, provided with 100% recommended dose of fertilizers and without AMF application observed to be better.

#### REFERENCES

Afe AI 2017. Growth and yield of sesame (Sesamum indicum L.) as influenced by population density of component cowpea in

Received 01 July, 2024; Accepted 11 September, 2024

sesame – cowpea mixture. *Journal of Experimental Agriculture International* **17**(1): 1-6.

- Afe Al and Atanda S 2015. Percentage yield difference, an index for evaluating intercropping efficiency. *American Journal of Experimental Agriculture* **5**(5): 459-465.
- Aulakh GS, Singh G and Singh A 2019. Studies on intercropping of maize (Zea mays L.) with pea (Pisum sativum L.) genotype. Indian Journal of Ecology 46(2): 354-357.
- Bhatti IH, Ahmad R and Nazir MS 2005. Agronomic traits of sesame as affected by grain legumes intercropping and planting patterns. *Pakistan Journal of Agricultural Sciences* **42**(1-2): 56-60.
- Bhatti I H, Ahmad R, Jabbar A, Nazir MS and Mahmood T 2006. Competitive behaviour of component crops in different sesamelegume intercropping systems. *International Journal of Agriculture and Biology* **8**(2): 165-167.
- Bindhu JS, Raj SK and Girijadevi L 2014. Sustainable system intensification of sesamum (*Sesamum indicum*) through legume intercropping in sandy loam tract of Kerala. *Journal of Crop and Weed* **10**(2): 38-42.
- Carlsson G and Huss-Danell K 2014. Does nitrogen transfer between plants confound 15N-based quantifications of N2 fixation. *Plant and Soil* **374**: 345-358.
- Chen M, Arato M, Borghi L, Nouri E and Reinhardt D 2018. Beneficial services of Arbuscular Mycorrhizal Fungi from ecology to application. *Frontiers in plant science* **9**: 1270.
- de Wit CT 1960. On competition. Verslagen Landbouwkundige Onderzoekigen 66(8): 1-82.
- Donald CM 1963. Competition among crop and pasture plants. *Advances in Agronomy* **15:** 1-118.
- Fustec J, Lesuffleur F, Mahieu S and Cliquet JB 2010. Nitrogen rhizodeposition of legumes: A review. Agronomy for Sustainable Development **30**: 57-66.
- Guedes RE, Rumjanek NG, Xavier GR, Guerra JGM and Rebeiro RLD 2010. Cowpea and corn intercrops in organic farming system aiming at production of immature beans and spikes. *HorticulturaBrasileira* **28**(2): 174-177.
- Jiwan, Sharma R and Walia SS 2021. Weed composition and nutrient uptake by weeds in sole and intercropping during *Rabi* season. *Indian Journal of Ecology* 48(1): 300-303.
- KAU [Kerala Agricultural University]. 2016. Package of Practices Recommendation: Crops 2016. (15<sup>th</sup> Ed.). Kerala Agricultural University, Kerala, Thrissur, p 393.
- Kotadiya PB, Leva RL, Usadadiya BJ, Chaudhary B and Shiyal S 2023. Effect of sesame based intercropping system with different levels of nitrogen on nutrient content, uptake and post-harvest status. *Pharma Innovation Journal* **12**(7): 1101-1108.
- Kumar D, Ardeshna R, Singh M, Makarana G, Kumar R, Kumar S, Meena R and Prajapat B 2021. Sustainable production of sesamum through legume intercropping: A review. *Indian Journal of Ecology* 48(1): 1403-1413.
- Lal RB and Ray S 1976. Economics of crop production of different intensities. *Indian Journal of Agricultural Sciences* **46**: 93-96.
- McGilchrist IA 1965. Analysis of competition experiments. *Biometrics* **21**: 975-985.
- Verma SP and Modgal SC 1983. Use of equivalent yields in cropping systems. *Himachal Journal of Agricultural Research* **9**(2): 89-92.
- Willey RW 1979. Intercropping-its importance and research needs. Part-1. Competition and Yield advantages. Agronomy Journal 71(2): 115-119.
- Willey RW, Matarajan M, Reddy MS, Rao MR, Nambiar PTC, Kammainan J and Bhatanagar VS 1980 Intercropping studies with Annual Crops, pp 83-97. In: Nugent, J. and O'Connor, M. (eds), *Better Crops for Food.* Novartis Foundation Symposia, John Wiley and Sons, New Jersey, USA.



# Influence of Potassium and Phosphate Solubilizing Bacteria on Growth and Development of Davana (*Artemisia Pallens* Bees)

# Charul Khatri, Priyanka Mehra, Arul Prakash T., K.M. Prakhyath, N.D. Yogendra, B.S. Dattesh and V.S. Pragadheesh

CSIR – Central Institute of Medicinal and Aromatic Plants Research Centre, Bengaluru-560 065, India \*E-mail: yogendra.nd@cimap.res.in

**Abstract:** The study assessed the impact of KSB and PSB application on *Artemisia pallens* growth, yield, and its impact on endophytic and rhizosphere bacterial populations. The plant height and flower diameter of davana were increased due to the application of  $T_{7:}100\%$  RDF+PSB+KSB over control and Recommended Dose of Fertilizer. Significantly higher plant height and flower diameter was 100% RDF+PSB+KSB compared to RDF alone. Treatments 75% RDF+PSB+KSB exhibited higher fresh plant weight than RDF alone. Soil samples from  $T_7$  showed the highest bacterial population (49×104 Cfu/ml), enhancing plant yield and essential oil quality. Endophytic bacteria were predominantly Gram-negative with rod and cocci shapes. RDF with PSB and KSB increased respiration rate, indicating faster nutrient mineralization. Overall, 75% RDF with PSB and KSB benefits davana performance, bacterial population, and soil nutrient mineralization, reducing reliance on synthetic fertilizers and environmental risks.

#### Keywords: Davana, Essential oil, PSB, KSB, Endophytes

Artemisia pallens Bees (Davana) is an aromatic annual herb, belonging to the Compositae family, indigenous to India and cultivated in Karnataka, Tamil Nadu, and Andhra Pradesh. The genus Artemisia contains more than 480 species around the world, out of which about 45 species are found in India (Singh et al 2021). It is a short-season crop from November to March, prefers red-loamy and well-drained soil performs better during moderate winter conditions; however, it cannot withstand heavy rain. Davana has high economic value due to its aromatic essential oil, commercially used to replace artificial flavoring agents with natural flavoring in food products like cakes, candy, chewing gum, and ice creams (Trendafilova et al 2020). The essential oils are also used widely in the aroma industry for making perfumes on the other side davana extract is used in the beverage industry. Monoterpenes, sesquiterpenes, and derivatives of fatty acids make up the essential oil constituents. More than ninety components were identified, with cis-davanone, bicyclo germacrene, trans-ethyl cinnamate, davana ether isomer, and spathulenol being the main ones. These constituents have also been shown to be effective against certain pathogenic microbes (Singh et al 2021).

Chemical fertilizers are widely employed in large-scale commercial davana production to boost output and meet customer demand, which lowers the amount of nutrients that are readily available in the soil and microflora (Kumar et al 2019). Every nutrient that the plant absorbs from various sources plays a specific role that none of the others can provide. The three main nutrients-nitrogen (N), phosphorus (P), and potassium (K) are needed in high concentrations for the majority of crops and are essential to plant metabolism. The majority of primary minerals are insoluble due to their complex structure, which renders them unavailable for plant uptake. Potassium is designated as a macronutrient because plants absorb substantial amounts of it. Little amount of potash is primarily obtained in India from bedded marine evaporite deposits, glauconite, polyhalite, and sylvite, as well as surface and subsurface potash-rich brines. In addition to minerals, many other microorganisms coexist in symbiotic relationships with plants, including endophytic and rhizosphere microbes. Because of the reported advantages of these microbes, scientists have been interested in them. These microbes have an innate connection to their host plant and are capable of developing endogenously. Within the rhizosphere, bacteria identified as plant growth-promoting bacteria (PGPB) have the capacity to directly enhance plant growth by increasing the absorption of macronutrients and minerals, along with elevating essential hormone concentrations. Additionally, these bacteria can indirectly support plant growth by mitigating the harmful impacts of pathogens. Numerous plant tissues, such as leaves, stems, roots, and flowers, have been found to have endophytic bacteria (Kobayashi and Palumbo 2000). These tissues are a potential but not utilized source of phyto-constituents. Bacterial strains found in association with roots can contribute to the improvement of plant health and growth through mechanisms such as phyto-stimulation, biofertilization, and biocontrol. Pseudomans, Pantoea, and

*Bacillus* (Andreolli et al 2016), are advantageous for plant growth. This pot experiment uncovered a significant gap in research regarding the effects of applying potassiumsolubilizing bacteria (KSB) and phosphate-solubilizing bacteria (PSB) on the economically important davana crop.

#### MATERIAL AND METHODS

Pot experiments followed by analytical lab procedures were conducted to evaluate the effect of phosphate and potassium solubilizing bacteria on the growth, development, and essential oil yield of *Artemisia pallens* followed by the identification and characterization of endophytes and various microorganisms in a soil sample from the rhizosphere of davana.

**Study area:** The pot experiment was conducted at CSIR-Central Institute of Medicinal and Aromatic Plants, Research Centre Bengaluru, Karnataka during 2022-23. Geologically the experiment site was located at 77°33'30" East longitude and 13°05'06" North latitude with a mean sea level of about 920 meters under the southern plateau and hilly zone.

**Experimental design:** The pot experiment consisted of eight treatments including T<sub>1</sub> (control), T<sub>2</sub> (RDF), T<sub>3</sub> (100% RDF+KSB), T<sub>4</sub> (75% RDF+KSB), T<sub>5</sub> (100% RDF+PSB), T<sub>6</sub> (75% RDF+PSB), T<sub>7</sub> (100% RDF+PSB+KSB) and T<sub>8</sub> (75% RDF+PSB+KSB) which were replicated thrice in randomized block design. The weather parameter did not affect the growth due to the pots were kept under a glass house to maintain a stable environment.

**Crop husbandry:** The experiment was performed in pots capable of filling 5 kg of soil using davana as a test crop. The 35 days old seedlings of davana raised in the nursery have been transplanted into pots containing sand, soil, and manure in equal proportions (1:1:1). The required quantity of microbial inoculation was applied at the rate of 10 kg/ha on hectare soil weight basis to respective pots along with RDF. Regular watering was done to maintain the soil moisture as and when required. No signs of pest or disease incidence were noted throughout the crop growth period; however, the precautionary spray of pesticide was taken accordingly. The crop was harvested at physiological maturity followed by drying and extraction of essential oil.

**Extraction of essential oil:** The herbage from each treatment was extracted individually, and using a Clevenger-type apparatus (Soxhlet extraction unit manufactured by Super Scientific Company Bengaluru), the herbage was hydro-distilled for six hours in a 2000 ml flask (Kumara et al 2023). The Triplicate of each treatment sample was obtained and the mean essential oil yield values were taken for the computation. The extracted essential oil was collected over glass vial contains anhydrous sodium sulphate and kept in

cold and dark place until further analysis. To identify various components, gas chromatography analysis was also performed on essential oils.

#### Microbial Analysis of Plant and Soil Samples

Soil and plant sample collection: The soil samples for determination of the microbial population were collected at a depth of 10-15 cm below the surface from each treatment. The isolation of PSB was carried out from the rhizosphere soil. The well-grown plants are identified and plucked by gentle pulling. It was possible to isolate endophytic bacteria from the davana plant's leaves, stems, buds, and roots. Therefore, the mentioned samples were all taken straight to the lab and stored in labelled sterile polythene bags. We ran an hour-long analysis on these samples to get better findings. Bacterial population count and isolation of bacteria: The bacterial population present in the rhizosphere soil was counted at the beginning, and after harvest. Phosphatesolubilizing bacteria were isolated and the bacterial count was estimated in each sample by using the serial dilution technique in nutrient agar media and incubated at 30 °C for 24 hours. The microbial growth was recorded by the colony count parameter. Each colony that appeared on the plate was considered one colony-forming unit (CFU) (Creach et al 2003). The different colonies according to color, shape, and size were further subjected to the identification of genera using staining techniques. As per morphological and biochemical features, the isolates were characterized and classified up to the genus level on Pikovskaya's and Aleksandrov's agar media respectively.

**Isolation of phosphate solubilizing bacteria:** 1g of davana crop rhizospheres soil sample was collected and serially diluted and poured on pikovskaya medium agar (Himedia M520-100G), and incubated at 30°C until halazone appeared. Halazone appearance indicates the presence of phosphate solubilization, and it was recorded as a phosphate solubilization index (SI) (Pande et al 2017).

**Isolation of endophytic bacteria:** Davana plant sample was dissected into roots, stems, and leaves and then thoroughly washed. Surface-sterilized tissues were inoculated on nutrient media (NA) and incubated for 24 hours at 30°C. Plates were sub cultured to gain pure culture isolate and to revive bacteria. Each issue sample's colonies were selected (Based on morphological appearance) for further genera identification by biochemical analysis (Duhan et al 2020).

**Microbial respiration:** The 250 g of soil mixed with the respective solubilizers with and without RDF were prepared. A 50 ml centrifugal tube containing 25 ml of 0.5N NaOH suspended inside the conical flask was incubated at 32°C for 24 hours and carbon dioxide emitted due to the respiration of bacteria was trapped in NaOH which converted to sodium carbonate. Unused NaOH was back titrated against standard sulfuric acid ( $H_2SO_4$  0.5M) in the presence of phenolphthalein indicator. After every 24-hour cycle, fresh NaOH was filled and repeated the above procedure for 10 days. The respiration rate was calculated (Dehsheikh et al (2020).

$$MR = \frac{(N \times (V1 - V2) \times 22)}{(T \times DW)}$$

Where, MR represents the microbial respiration of the soil, expressed in terms of mg of emitted  $CO_2$  per gram per day. N denotes the normality of the acid, V1 is the volume of  $H_2SO_4$ used for titrating the remaining NaOH in the standard control solution (without inoculants), V2 is the volume of  $H_2SO_4$  used for titrating the remaining NaOH in the sample, T represents time in days, and DW stands for the weight of the inoculants.

**Statistical analysis:** Statistical analyses were performed using SPSS, version 16.0 for the Duncan's Multiple Range Test (DMRT), and data visualization was conducted using GraphPad Prism, version 8.0.2.

#### **RESULTS AND DISCUSSION**

#### Effect on Growth, Development, and Yield of Davana

**Growth attributes:** The RDF along with biofertilizers alone or in combination have a significant effect on the growth and yield of davana as compared to control the statistical analysis of different growth attributes such as plant height and numbers of branches (Table 1). Application of 100% RDF+PSB+KSB increase significant plant height (47.40 cm) over control treatment (35.08 cm) and RDF (39.38cm) which was on par with treatment of 100% RDF+PSB+KSB. The sole application of 75% RDF+KSB, 100% RDF+KSB, 100% RDF+PSB, and 75% RDF+KSB was significant in increasing plant height over RDF and control. Biofertilizers like VAM (Vesicular Arbuscular Mycorrhizae) and Azospirillum significantly enhanced the growth and development of Kadam seedlings. These biofertilizers notably increased as shoot length, collar diameter, number of leaves per plant, total leaf area per plant, root length, as well as fresh and dry biomass (Chauhan 2023). The number of branches of davana did not vary significantly due to the application of different solubilizers with recommended doses of fertilizers. The better availability and assimilation of nutrients from the soil by plants enhanced the better photosynthetic activity, build-up of cells, and their elongation where the better availability was aided by the nutrient solubilizing action of PSB and KSB application to the soil (Olaniyan 2022) probably caused the better growth of davana plant.

Yield attributes: An increment of 22.8 and 21.40% enlargement in flower diameter in treatment 75% RDF+PSB+KSB and 100% RDF+PSB+KSB respectively over the untreated plants (Table 1), which was statistically significant over untreated (2.76 mm) and RDF (3.24 mm). Where the different proportions (75 and 100%) of chemical fertilizer with the sole application of PSB and KSB significantly influenced the flower diameter of davana over RDF alone treatment. The least yield was recorded in the control treatment. Kumari et al (2019) also reported a significantly higher flower diameter of marigolds on application of PSB with RDF. The significant rise in davana single plant weight was observed due to the application of RDF, increasing from 33.27 g (control) to 39.43 g. This further increased to 48.14 g and 46.78 g with 100% and 75% RDF+PSB+KSB, respectively. Comparatively, 75% RDF with PSB (42.38 g) or KSB (43.53 g) was similar to 100% RDF with

Treatment	Plant height (cm)	No. of branches	Flower diameter (mm)	Single plant fresh weight (g)	Oil recovery (%)
<b>T</b> <sub>1</sub>	35.08 e	18.67 a	2.76 d	33.27 d	0.21 c
<b>T</b> <sub>2</sub>	39.38 d	21.00 ab	3.24 c	39.43 c	0.22 bc
T <sub>3</sub>	43.78 bc	22.67 b	3.63 b	43.93 b	0.23 a
<b>T</b> <sub>4</sub>	42.49 c	22.33 b	3.56 b	43.53 b	0.24 a
T <sub>5</sub>	44.90 abc	21.47 ab	3.62 b	43.65 b	0.22 a
<b>T</b> <sub>6</sub>	45.02 abc	22.17 b	3.47 bc	42.38 b	0.22 ab
<b>T</b> <sub>7</sub>	47.40 a	21.33 ab	4.14 a	48.14 a	0.23 a
T <sub>8</sub>	46.21 ab	21.00 ab	4.20 a	46.78 a	0.23 a
CD (1%)	2.85	NS	0.28	2.47	NS

The common letter(s) in a column shows no difference at the level of 5% probability as per DMRT; T<sub>1</sub>: Control, T<sub>2</sub>: RDF, T<sub>3</sub>: 100% RDF+KSB, T<sub>4</sub>: 75% RDF+KSB, T<sub>5</sub>: 100% RDF+PSB, T<sub>5</sub>: 75% RDF+PSB+KSB, T<sub>5</sub>: 75% RDF+PSB+KSB, T<sub>6</sub>: 75% RDF+PSB+KSB, T<sub>75</sub> RDF+PSB+KSB, T<sub>75</sub> RDF+PSB+KSB, T<sub>75</sub> RDF+PSB+KSB, T<sub>75</sub> RDF+PSB+KSB, T<sub>75</sub> RDF+PSB+KSB

PSB (43.65 g) or KSB (43.93 g), all significantly higher than control and RDF. The improved nutrient uptake, growth, photosynthesis, and biomass accumulation in davana were due to the mobilization of sparingly soluble P and K minerals or chelating ions, aligning with findings in geranium (Prasad et al 2012) and Sabja (Cheena et al 2021).

Essential oil recovery: The chemical composition of davana essential oil showed little variation among different treatments (Table 2). The major constituent, *cis*-davanone, ranged from 48.82% to 51.25%, with the highest content in the RDF treatment, followed by 75% RDF+PSB. The combined application of 100 and 75% RDF+PSB+KSB recorded cis-davanone levels of 50.88% and 49.78%, respectively. Davana ether (Isomer-1) was the second major constituent, ranging from 11.15% to 13.25%, with the highest levels in 75% followed by 100% RDF+KSB. Bicyclogermacrene content was highest in 75% RDF+PSB+KSB (5.32%) and lowest in the control treatment (4.05%). Other constituents, such as trans-davanone + trans-nerolidol, humulene epoxide-II, epi-alpha-cadinol, cismethyl jasmonate, davanol acetate, and cis-hydroxy davanone, showed minimal variation among treatments.

# Effect Microbiological Parameters of Soil and Plant Sample

Isolation of phosphate and potassium solubilizing bacteria from rhizosphere soil: A single phosphatesolubilizing bacterial colony was isolated, characterized, and identified. The organism was Gram-negative, whitecoloured, and irregular in the shape of the colony; whereas the cell was in rod shape and rough surface with flat elevation. The colony exhibited clear halo zones around the bacterial growth; this isolate had a high phosphate solubilization index (PSI) of 2.5 mm. The isolates might be attributed to rhizosphere colonization, and better nutrient availability reported by Rafique et al (2012). The ability of PSB to solubilize insoluble phosphorus in Pikovaskya's agar medium by observing the halo zone thus concluding that bacteria tend to solubilize the insoluble phosphorous (Yaghoubi et al 2018).

Isolation of endophytic bacteria from leaf, bud, stem, and root of davana: Bacterial species were isolated based on the distinctive features of the colonies. Based on the findings, a variety of bacteria were identified in the leaf, stem, and root. In terms of morphological characterization, the endophytic bacterial isolates showcased a range of colony attributes, including diverse shapes (circular and irregular), colours (orange, white, and yellowish), margins (regular and wavy), and textures (Fig. 1). Concerning gram staining, 6 isolates demonstrated Gram-negative characteristics, while 2 exhibited Gram-positive features. In the biochemical and physiological characterization, all isolates underwent various tests including catalase, IMViC, H2S production, methyl red, and oxidase tests. Positive results were observed in certain isolates, (Table 3). Majority of bacterial endophytes might belong to the genus Bacillus and Streptococcus. Davana possesses a diverse taxonomic range of endophytes, with numerous studies (Suryanarayanan et al 2009) primarily concentrating on fungal endophytes, and there has been limited exploration of endophytic bacteria in this context.

**Impact on bacterial population:** The initial soil sample of the pot experiment count of bacteria was  $25 \times 10^4$  Cfu/ml and varied significantly in soil sampled before harvest of the crop in each treatment. The highest bacterial count was in the combined application of 100% RDF+ PSB+KSB treatment T<sub>7</sub> (49×10<sup>4</sup> Cfu/ml), which was followed by 75% RDF+ PSB+KSB T<sub>8</sub> (49×10<sup>4</sup> Cfu/ml). The lowest bacterial count was in RDF alone T<sub>2</sub> (7×10<sup>4</sup> Cfu/ml) and control (7×10<sup>4</sup> Cfu/ml). Similarly, the impact of the sole application of KSB and PSB along with a recommended dose of inorganic fertilizer (75% and 100% RDF) on bacterial count was obtained after the application of T<sub>4</sub> (34 ×10<sup>4</sup> Cfu/ml) was similar to T<sub>6</sub>, T<sub>3</sub> and T<sub>5</sub> (Fig. 2). The synergic effect of combined application of both the combination and sole application of PSB and KSB

Table 2. Effect of PSB and KSB on essential oil constituents of Artemisia pallens

Treatment	Bicyclo germacne	Davana ether (Isomer -1)	Cis-davanone	Davanol acetate	<i>Cis</i> -hydroxy davanone	Davana ether (Isomer -2)	<i>Trans</i> -davanone + <i>trans</i> -nerolidol	<i>Cis</i> -methyl jasmonate
Τ,	4.05	11.15	48.86	2.6	2.54	2.98	1.78	2.5
<b>T</b> <sub>2</sub>	4.1	11.25	52.56	2.86	2.85	2.99	1.83	3
Τ <sub>3</sub>	4.65	12.65	48.82	2.57	2.6	4.85	1.78	2.28
T <sub>4</sub>	4.85	13.25	49.85	2.78	2.47	3.97	1.86	2.57
T <sub>5</sub>	4.25	12.45	50.65	2.72	2.38	4.23	1.09	2.74
<b>T</b> <sub>6</sub>	4.98	12.83	51.25	3.43	2.43	3.02	1.09	2.3
Τ,	5.1	11.35	50.88	3.94	2.44	3.15	1.86	2.76
T <sub>8</sub>	5.32	11.35	49.48	2.63	2	3.92	1.76	2.57

See Table 1 for details

along with inorganic fertilizer, positive synergistic effects with soil bacteria were observed, which promoted each other's growth. However, the combined application exhibited an overall greater bacterial population as compared to the initial RDF alone and controlled the bacterial population. Aswathy et al (2017) observed that the rapid growth in rhizosphere microbial population, which may be attributed to the multiplication of the strains in the rhizosphere, uses root exudates produced by the plants and synergistic interactions



Fig. 1. Morphological characterization of endophytic bacteria

between introduced microbial inoculants and also the microorganisms in the root zone of the crop. In their study on rose-scented geranium, Negi et al (2022) noted that the presence of plant growth-promoting bacteria contributes to maintaining soil ecological balance, simultaneously improving both yield and essential oil quality. The use of microbial inoculants has the potential to mitigate the negative effects of chemical fertilizers and as a result, promote both the quantity and the quality of plant yield. Microbial inoculants offer a more sustainable way of delivering plant nutrients, and their utilization is environmentally beneficial too.

**Microbial respiration:** Among all the treatments RDF+PSB+KSB was the first to attain the highest peak of respiration i.e., on 3rd day whereas the sole application of RDF+PSB and RDF+KSB was on the 4th day. Among sole applications of PSB and KSB without RDF, inflated emission was recorded in KSB than in PSB with peak respiration on the 6th and 5th day respectively; whereas with RDF it was 4th day in both RDF+PSB and RDF+KSB have a lower amount of carbon evolution than all other treatments indicating quick recession of respiration and more mineralization of substrate. The high



Fig. 2. Effect of PSB and KSB on Bacterial enumeration

Table 3. Biochemica	I characterization of	<sup>r</sup> endophytes and	∄ PSB in soil
---------------------	-----------------------	-----------------------------	---------------

Biochemical test	Endo	phytes	Soil	
	G-Positive	G-Negative	G-Positive	G-Negative
Catalase	100.00	0.00	46.67	53.33
Soxidase	50.00	50.00	40.00	60.00
Starch hydrolysis	62.50	37.50	53.33	46.67
Phenol red	62.50	37.50	46.67	53.33
Urease test	50.00	50.00	46.67	53.33
Indole test	37.50	62.50	66.67	33.33
Methyl red test	50.00	50.00	53.33	46.67



Fig. 3. Respiration study of different solubilizers effect

rate of respiration represents the more activity of microbes intern the rapid mineralization of nutrients (Jacoby et al 2017). The application of RDF showed more rapid respiration in both organisms indicating the significance of bio-fertilizers instead of using chemical fertilizers for the cultivation of crops. The study describes KSB as having a quicker mineralization capacity than PSB.

## CONCLUSION

Application of 75% of recommended dose of fertilizer (RDF) with KSB and PSB had saved 25% chemical fertilizer compared to 100% RDF which may increase the annual income by improving the growth parameters, yield, and essential oil content of davana by the symbiotic bacterial effect. The symbiotic bacterial effect increases the crop productivity by the solubilization ability which reduce the fertilizer utility up to 25% might be valuable to economic and environmental viewpoint while using biofertilizers along with chemical fertilizer.

#### ACKNOWLEDGEMENTS

The authors are thankful to the Director, CSIR-Central Institute of Medicinal and Aromatic Plants, Lucknow, India, and the Council of Scientific and Industrial Research, New Delhi, India.

#### REFERENCES

- Andreolli M, Lampis S, Zapparoli G, Angelini E and Vallini G 2016. Diversity of bacterial endophytes in 3- and 15-year-old grapevines of *Vitis vinifera* cv. Corvina and their potential for plant growth promotion and phytopathogen control. *Microbiological Research* 183: 42-52.
- Aswathy TS, Jintu J, Dhanya MK, Sathyan T, Preethy TT and Murugan M 2017. Effect of biofertilizers and organic supplements on general and beneficial microbial population in the rhizosphere of black pepper cuttings (*Piper nigrum* L.). *International Journal of Chemical Studies* **5**(6): 1260-1264.
- Chauhan P, Behera LK, Tandel MB, Thakur NS, Chauhan RS and Dholariya CA 2023. Influence of Biofertilizers on Early-stage seedling Growth, Biomass and vigour of *Anthocephalus cadamba* (Roxb.)Miq. *Indian Journal of Ecology* **50**(4): 969-974.

Received 30 June, 2024; Accepted 11 September, 2024

- Cheena J, Veni VK, Padma M and Sreenivas M 2021. Studies on growth and yield of Sabja (*Ocimum basilicum*) influenced by different organic manures and biofertilizers. *Growth* **9**: 1.
- Creach V, Baudoux AC, Bertru G and Le Rouzic B 2003. Direct estimate of active bacteria: CTC use and limitations. *Journal of microbiological methods* **52**(1): 19-28.
- Dehsheikh AB, Sourestani MM, Zolfaghari M and Enayatizamir N 2020. Changes in soil microbial activity, essential oil quantity, and quality of Thai basil as response to biofertilizers and humic acid. *Journal of Cleaner Production* **256**: 120439.
- Duhan P, Bansal P and Rani S 2020. Isolation, identification and characterization of endophytic bacteria from medicinal plant *Tinospora cordifolia. South African Journal of Botany* **134**: 43-49.
- Jacoby R, Peukert M, Succurro A, Koprivova A and Kopriva S 2017. The role of soil microorganisms in plant mineral nutrition-current knowledge and future directions. *Frontiers in Plant Science* 8: 1617.
- Kobayashi DY and Palumbo JD 2000. Bacterial Endophytes and There. *Microbial Endophytes* **200**: 99-233.
- Kumar M, Abraham T, Kumari N, Dawson J and Patel JJ 2019. Effect of tillage, nitrogen and zinc management on yield and yield components of wheat (*Triticum aestivum*) in North Eastern Plains Zone of India. *Indian Journal of Agronomy* 64(1): 67-74.
- Negi S, Mazeed A, Maurya P, Kumar D and Suryavanshi P 2022. Effect of microbial bio-elicitors on yield and chemical composition of essential oil in *Pelargonium graveolens*. *Indian Journal of Ecology* 49(3): 1208-1213.
- Olaniyan FT, Alori ET, Adekiya AO, Ayorinde BB, Daramola FY, Osemwegie OO and Babalola OO 2022. The use of soil microbial potassium solubilizers in potassium nutrient availability in soil and its dynamics. *Annals of Microbiology* **72**(1):45.
- Pande A, Pandey P, Mehra S, Singh M and Kaushik S 2017. Phenotypic and genotypic characterization of phosphate solubilizing bacteria and their efficiency on the growth of maize. *Journal of Genetic Engineering and Biotechnology* **15**(2): 379-391.
- Prasad A, Kumar S, Pandey A and Chand S 2012. Microbial and chemical sources of phosphorus supply modulate the yield and chemical composition of essential oil of rose-scented geranium (*Pelargonium* species) in sodic soils. *Biology and Fertility of Soils* 48: 117-122.
- Rafique M, Riaz A, Anjum A, Qureshi MA and Mujeeb F 2018. Role of bioinoculants for improving growth and yield of okra (Abelmoshus esculentum). Universal Journal of Agricultural Research 6(3): 105-112.
- Kumara, RR, Keerthi, PE and Yogendra, ND 2023. Floral biology insights into essential oil yield and chemical composition in davana (Artemisia pallens Bess), a high-value aromatic plant of India. Journal of Spices and Aromatic Crops 32(1): 80-89.
- Singh S, Bhatt D, Singh MK, Sundaresan V, Tandon S, Padalia RC and Verma RS 2021. New insights into the chemical composition, pro-inflammatory Cytokine inhibition profile of Davana (*Artemisia pallens* Wall. ex DC.) essential oil and cis-Davanone in primary macrophage cells. *Chemistry & Biodiversity* 18(11): e2100531.
- Suryanarayanan TS, Thirunavukkarasu N, Govindarajulu MB, Sasse F, Jansen R and Murali TS 2009. Fungal endophytes and bioprospecting. *Fungal Biology Reviews* **23**(1-2): 9-19.
- Trendafilova A, Moujir LM, Sousa PM and Seca AM 2020. Research advances on health effects of edible *Artemisia* species and some sesquiterpene lactones constituents. *Foods* **10**(1): 65.
- Yaghoubi Khanghahi M, Pirdashti H, Rahimian H, Nematzadeh G and Ghajar Sepanlou M 2018. Potassium solubilising bacteria (KSB) isolated from rice paddy soil: From isolation, identification to K use efficiency. *Symbiosis* **76**: 13-23.



Manuscript Number: 4344 NAAS Rating: 5.38

# Productivity and Economic Assessment of Diverse Rice (*Oryza sativa* L.) Varieties with Varied Nitrogen Levels in Eastern Indo-Gangetic Plains

Mohammad Hashim, Man Mohan Deo<sup>1</sup> and Sanjeev Kumar<sup>2</sup>

ICAR- Indian Agricultural Research Institute, Regional Station Pusa Samastipur -848 125, India <sup>1</sup>ICAR- Indian Institute of Pulses Research, Kanpur-208 024, India <sup>2</sup>ICAR- National Dairy Research Institute, Karnal -132 001, India E-mail: hashimagronomy@gmail.com

Abstract: During the Kharif seasons of 2020 and 2021, a field experiment was performed at the Indian Agricultural Research Institute (ICAR), Regional Station, Pusa, Bihar, to investigate the impact of different rice varieties and nitrogen levels on growth, yield, and economic aspects. Five varieties of rice, namely PNR 381, Rajendra Sweta, Sugandha-5, Pusa Sambha-1850, and Pusa-44, along with five nitrogen levels (0, 60, 120, 180, and 240 kg N/ha), were assessed. A progressive improvement in growth parameters and yield as nitrogen levels increased from the control to 200% recommended dose of nitrogen (RDN) equivalent to 240 kg/ha. The application of nitrogen at 240 kg N/ha resulted in significantly elevated plant height (102.1 cm), panicle length (27.1 cm), 1,000 grain weight (21.3 g), grains per panicle (200.9), grain yield (4.94 t/ha), net returns (₹ 61104/ha), and B:C ratio (2.27), as well as increased gross energy output, net energy return, energy use efficiency, and energy productivity. The Pusa-44 variety demonstrated superior performance compared to the other varieties tested. In conclusion, for the calcareous soil of the Indo-Gangetic Plains, cultivating the Pusa-44 rice variety along with the application of 150% RDN (180 kg N/ha) emerges as the optimal strategy to improve productivity, profitability and energetic.

#### Keywords: Economics, Energetics, Nitrogen levels, Rice, Varieties, Yield

Rice (Oryza sativa L.) constitutes a fundamental dietary staple for over half of the South Asian population and serves as a principal energy source in the North-Eastern Plain Zone of India (Hashim et al 2021). Despite being a water-intensive crop, it is cultivated across diverse ecosystems in the Indo-Gangetic Plains of India. The demand for rice continues to escalate, with projections estimating a requirement of approximately 140 million tons by 2025 (Singh et al 2019). In order to endure current food production and meet future demands, India must enhance rice productivity by approximately 3% annually (Kumar et al 2017). Rice is highly responsive to nutrient management, and the judicious application of fertilizers is critical for achieving sustainable and increased rice production. Improper nutrient use, particularly the indiscriminate application of major nutrients and intensive cropping, can result in a negative nutrient balance (Nadeem and Farooq, 2019). The sustainability and productivity of the rice-wheat cropping system are jeopardized by declining soil health, leading to diminished grain yields (Kumar et al 2016, Singh et al 2015).

The continuous cultivation of intensive rice-wheat sequences, coupled with discriminatory nutrient application, has contributed to the degradation of soil fertility, including the depletion of primary, secondary, and micronutrients (Prasad et al 2019). Given rice's substantial reliance on

nitrogenous fertilizers, it is imperative to address the varied nitrogen (N) requirements in transplanted rice due to continuous flooding during cultivation. Substantial losses, exceeding 40–50% of applied nitrogen, occur through mechanisms such as ammonia volatilization, denitrification, leaching, and runoff after fertilizer application (Hakeem et al 2012, Fu et al 2023, Kumari et al 2022). These losses not only diminish yield and economic efficiency but also pose severe environmental ramifications (Houlton et al 2019, Xu et al 2020, Cowan et al 2021).

Therefore, it is imperative to ascertain optimal nitrogen levels and identify high-yield varieties. Limited research has been conducted on determining the ideal nitrogen levels for diverse rice varieties cultivated in calcareous lowland soil. Consequently, this study aims to evaluate the performance of different rice varieties in calcareous soil within the Indo-Gangetic Plains under varying nitrogen levels.

### MATERIAL AND METHODS

**Experimental site:** Two-year field experiment, encompassing the consecutive rainy seasons of 2020 and 2021, was executed at the ICAR-Indian Agricultural Research Institute, Regional Station, Pusa, Samastipur, Bihar, India. The experimental site is characterized by geographical coordinates of 25°58'49" N latitude, 85°40'48" E longitude, and an elevation of 52.12 meters above mean sea level. The region experiences a subtropical, humid climate characterized by hot summers and cold winters. The recorded rainfall during 2020 and 2021 was 1633.2 mm and 1883.6 mm, respectively (Fig. 1). The soil at the experimental site exhibited a sandy loam texture, low organic carbon content (0.36%), a pH of 8.3, an electrical conductivity (EC) of 0.25 dS/m, and an available nitrogen content of 185 kg/ha.

Treatment details and crop management: Twenty-five treatment combinations, involving five levels of nitrogen application (control, 50% RDN, 100% RDN, 150% RDN, and 200% RDN) in the main-plot and five rice varieties (PNR-381, Rajendra Sweta, Sugandha-5, Pusa Sambha-1850, and Pusa-44) in the sub-plot, were examined using a split-plot design with three replications. The experiment spanned two consecutive years, with rice planted in a fixed plot and treatments superimposed on the same plot each year. Seedlings, grown in a well-prepared nursery bed and transplanted after 21 days in the main field with spacing of 20 cm × 15 cm. The recommended nitrogen doses was applied, with half as basal and the remaining half top-dressed in two equal split doses at active tillering and panicle initiation stage. Uniform doses of phosphorus and potassium were applied at 60 kg P<sub>2</sub>O<sub>5</sub> and 40 kg K<sub>2</sub>O, respectively, as basal doses.

Growth parameters, yield and economics: Upon reaching maturity, one border row surrounding the experimental plots was harvested, and the remaining net plot area was manually harvested using sickles to determine grain and straw yields. Dry matter efficiency (DME) was calculated using the methodology proposed by Kumar et al (2017). By multiplying the grain yield with the minimum support price (MSP) and incorporating the market value of by-products (such as straw) for the specific year, the gross return was determined. Net income, denominated in ₹/ha, was then computed as the



Fig. 1. Monthly average rainfall (mm), average maximum and minimum temperature (°C) in study locations (Pusa). Data were collected from the weather stations of the IARI, RS Pusa Bihar. Lines with filled triangles and squares represent the average minimum and maximum monthly temperatures, respectively, and bars represent the monthly average total rainfall

disparity between gross returns and cultivation costs. **Energy calculation:** Energy-use efficiency (EUE) was subsequently calculated as per Mittal and Dhawan (1988).

Net energy (MJ/ha) = Energy Output (MJ/ha) - Energy input (MJ/ha)

**Statistical analysis**: The data were analyzed by analysis of variance using OPSTAT Software (Sheoran et al 1998).

#### **RESULTS AND DISCUSSION**

Growth parameters: Diverse nitrogen levels exerted a pronounced influence on the height of rice plants, exhibiting a range from 72 cm to 102.1 cm across the various treatments (Table 1). Specifically, plots treated with 200% of the recommended dose of nitrogen (RDN) dosage, equivalent to 240 kg/ha of nitrogen, demonstrated the greatest plant height at 102.1 cm. This elevation in height was comparable to those observed with 100 and 150% nitrogen application, signifying a statistically significant increase compared to both the control and the 50% RDN treatment. The observed augmentation in plant height, attributed to the application of nitrogen, can be attributed to the efficient uptake and concentration of nitrogen, thereby enhancing rice growth and facilitating the production of dry matter. Comparable findings were reported by Hasanuzzaman et al (2012), Ritesh et al (2014), Nayak et al (2022), Zidan (2017). Conversely, control plots, characterized by an insufficient supply of nitrogen, exhibited the lowest plant height at 72.0 cm significantly lower than all other treatments. Rice varieties exhibited distinct variations in plant height at the maturity stage. Pusa Sugandha 5 manifested the tallest plants at 99.7 cm, followed by Pusa Sambha 1850 at 93.7 cm, both significantly surpassing the plant heights of other tested rice varieties. These differences are likely attributable to genetic variations influencing growth attributes, particularly plant height. In contrast, the PNR-381 variety displayed the lowest plant height at 86.3 cm, with Pusa-44 following closely at 87.7 cm.

**Yield attributing characters:** The panicle length demonstrated increase with elevated nitrogen levels, particularly at 200% RDN, suggesting a potential augmentation in nutrient availability conducive to panicle development (Bahuguna et al 2023, Mondal et al 2013). Panicle length exhibited a range from 21.3 cm in the control group to 27.1 cm at 200% RDN. Furthermore, distinct rice varieties displayed variability in panicle length, with Pusa Sugandha-5 exhibiting the longest panicles (28.0 cm), followed by Pusa-44, while Rajendra Sweta presented the shortest panicles (22.5 cm). Genetic variations likely

contribute to the observed differences in panicle length among diverse genotypes, corroborating findings in distinct genotypes (Sultana et al 2014). Similar trend was observed in earlier research (Bahuguna et al 2023, Sultana 2014, Mondal et al 2013, Nayak et al 2022 and Mohapatra et al 2024).

The influence of varying nitrogen levels on grain weight (1,000 grain weight) was notably apparent, with an increase from 18.1 g in the control to 21.3 g at 200% RDN, a value significantly higher than the control and equivalent to other nitrogen levels. The augmentation in panicle length, 1,000 grain weight, filled and total grains was particularly pronounced with elevated nitrogen levels up to 240 kg N/ha. This implies that nitrogen fertilization led to heightened metabolite synthesis in rice plants, contributing to the superior outcomes in terms of the number of filled grains per panicle, panicle length, and 1,000 grain weight. Conversely, control plots exhibited the highest number of chaffy or unfilled grains (47.2), whereas plots treated with 200% N/ha displayed the lowest count (Table 1). These findings resonate with previous studies (Hasanuzzaman et al 2012, Ritesh et al 2014, Zidan 2017, Ninju et al 2018, Bahuguna et al 2023). The rice genotypes exerted a substantial impact on both grain weight and the number of grains per panicle. Pusa Sugandha-5 demonstrated the highest grain weight (24.5 g/1000 grain), a statistically significant difference from other varieties, while Rajendra Sweta exhibited the lowest grain weight (15.7 g/1000 grain). This opinion aligns with the research of Renuka et al (2013), underscoring the role of genetics in determining rice test weight. Pusa Sambha-1850 documented the significantly maximum total number of grains per panicle (230.8/panicle), coupled with the highest count of unfilled or chaffy grains, surpassing even Rajendra Sweta. Pusa-44 displayed the lowest number of unfilled grains (27.2). This divergence in unfilled grain counts may contribute to the superior yield observed in Pusa-44.

Productivity: The influence of varying nitrogen levels on rice productivity is discernible through the data presented here. The augmentation in grain yield with the elevation of nitrogen levels, reaching its zenith at 200% of the recommended dose, registering at 4.94 t/ha (Table 2). Application of 200% RDN (240 kg N/ha) exhibited substantial elevated yield of grain (4.94 t/ha), straw (9.12 t/ha), and biological yield (14.05 t/ha), respectively, on par with 150% RDN (180 kg N/ha). The application of 200% RDN resulted in a 64.59, 31.42, 10.65, and 4.00% increased grain yield compared to control, 50%, 100%, and 150% RDN, respectively. The application of 150% RDN yielded 58.38, 26.36, and 6.39% higher grain yields compared to control, 50%, and 100% RDN, primarily attributed to heightened growth yield attributes with amplified levels of nitrogen application, as reported by Kumar et al (2018). The 4% increment in yield was with the fertilization of 200% RDN in comparison to 150% RDN and may be due to efficient vegetative growth and sink development, in alignment with earlier studies (Salahuddin et al 2009, Navak et al 2022, Kumar et al 2014. Consequently, overall yield

Treatment	Plant height	Panicle length	1000-grain		Grains/panicle		Harvest Index
	(cm)	(cm)	weight (g) -	Filled	Un-filled	Total	_
Nitrogen level							
Control	72.0	21.3	18.1	130.8	47.2	178.0	36.21
50% RDN	85.0	23.6	19.9	148.2	41.2	189.4	35.08
100% RDN	96.8	25.6	20.6	158.4	40.4	198.8	35.12
150% RDN	99.4	26.5	20.9	161.2	38.2	199.4	35.19
200% RDN	102.1	27.1	21.3	166.8	34.1	200.9	35.11
CD (p=0.05)	7.7	1.1	1.5	9.4	7.9	12.2	0.07
Varieties							
PNR-381	86.3	23.9	21.6	133.5	41.3	174.9	35.39
Rajendra Sweta	88.1	22.5	15.7	139.5	44.3	183.7	35.24
Pusa Sugandha-5	99.7	28.0	24.5	131.8	42.9	174.7	35.34
Pusa-44	87.7	25.2	21.8	175.2	27.2	202.4	35.35
Pusa Sambha-1850	93.7	24.4	17.3	185.4	45.4	230.8	35.40
CD (p=0.05)	3.5	1.2	0.9	12.9	5.7	14.8	0.04

 Table 1. Effect of nitrogen rates on growth and yield attributing characters of rice genotypes under transplanted condition in calcareous soil (Pooled data of 2 years)

Control (0 kg N), 50% RDN (60 Kg N), 100% RDN (120 Kg N), 150% RDN (180 Kg N), 200% RDN (240 Kg N)

increased as a result of an augmented number of panicles per unit area and heavier grains per panicle.

Similar to grain yield, straw yield exhibited an increase up to 200% RDN, leading to taller plants, augmented tiller density, and increased biomass production, consistent with the observations of Kumar et al (2018, 2019). Significantly heightened crop productivity (36.73 kg/ha/day) and DME (104.60 kg/ha/day) were recorded with the application of 200% RDN, comparable with 150% RDN, and significantly surpassed the remaining treatments. Incremental levels of nitrogen correspondingly improved crop productivity and DME up to the maximum applied nitrogen levels (200% RDN) due to higher yields within the same treatment.

In addition to nitrogen rates, rice varieties also exerted a significant influence on rice yields. Pusa-44 demonstrated substantial increased yield of grain (5.21 t/ha), straw (9.56 t/ha), and biological yield (14.77 t/ha) compared to remaining varieties tested. Pusa Sambha 1850 emerged as the second-highest yielding variety. These findings align with the results reported by earlier researchers (Danish et al 2022, Nayak et al 2022, Bahuguna et al 2023).

**Economics:** The aggregated data from two consecutive years indicates that the highest cultivation cost was observed in the 200% RDN treatment, amounting to ₹ 48,004/ha, followed by the 150% RDN. Conversely, the control plots, devoid of nitrogen application, exhibited the lowest cultivation cost., the application of 200% RDN resulted in significantly elevated gross returns (₹ 109,108/ha) and net

returns (₹ 61,104/ha) with a corresponding benefit-cost (B: C) ratio of 2.27, a performance that was on par with the 150% RDN treatment (Table 2). The 150% RDN application demonstrated 26.27 and 6.35% higher gross returns and 10.60 and 54.52% higher net returns compared to the 50% and 100% RDN treatments, respectively. However, the 200% RDN application yielded only a 5.84% increase in net returns compared to the 150% RDN treatment, possibly attributable to the higher yield achieved under the same treatment conditions. Similar inclination was previously reported by Kumar et al (2019). Significant disparities were evident among rice varieties concerning gross returns, net returns, and the B: C ratio. Pusa-44 exhibited notably higher gross returns (₹ 115,090/ha), net returns (₹ 68,602/ha), and a B: C ratio of 2.47, followed by Pusa Sambha-1850. The net returns of Pusa-44 surpassed those of Pusa Sambha-1850, Pusa Sugandha 5, Rajendra Sweta, and PNR-381.

**Energetics:** Combined data from two years demonstrated an escalating trend in input energy consumption corresponding to increasing nitrogen levels. The minimum input energy consumption was at lower nitrogen levels, and this increased progressively in the order of control <50% RDN <100% RDN <150% RDN <200% RDN, primarily attributed to heightened input requirements, notably nitrogen fertilization (Table 3). The highest energy input was registered at 200% RDN, followed by 150% RDN. Kumar et al (2018, 2019) also observed similar trend. The application of 200% RDN resulted in the highest net energy returns

 Table 2. Effect of nitrogen levels on yields and economics of rice genotypes under transplanted condition in calcareous soil (Pooled data of 2 years)

Treatment	Treatment Productivity (t/ha)		Crop	Dry matter	Cost of	Gross	Net	B:C ratio	
	Grain yield (t/ha)	Straw yield (t/ha)	Biological yield (t/ha)	(kg/ha/day)	efficiency (kg/ha/day)	Production (₹/ha)	returns (₹/ha)	returns (₹/ha)	
Nitrogen level									
Control	3.00	5.29	8.28	22.36	61.79	45235	65748	20513	1.45
50% RDN	3.76	6.95	10.70	27.95	79.65	45681	83045	37364	1.82
100% RDN	4.46	8.24	12.70	33.26	94.69	46390	98597	52207	2.13
150% RDN	4.75	8.74	13.49	35.38	100.54	47128	104862	57734	2.23
200% RDN	4.94	9.12	14.05	36.73	104.60	48004	109108	61104	2.27
CD (p=0.05)	0.21	0.39	0.61	1.62	4.61	-	4735	4735	0.10
Varieties									
PNR-381	3.55	6.50	10.05	30.86	87.41	46487.6	78364	31876	1.68
Rajendra Sweta	3.67	6.78	10.45	26.62	75.74	46487.6	81196	34708	1.74
Pusa Sugandha-5	4.12	7.56	11.68	32.46	91.96	46487.6	91021	44533	1.95
Pusa-44	5.21	9.56	14.77	35.22	99.79	46487.6	115090	68602	2.47
Pusa Sambha-1850	4.34	7.93	12.27	30.53	86.38	46487.6	95690	49203	2.05
CD (p=0.05)	0.30	0.55	0.84	2.25	6.39	-	6542	6542	0.14

Treatment	Energy input	Er	nergy output (MJ/I	ha)	Net energy	Energy ratio	Energy
	(MJ/ha) —	Grain	Straw	Total	— returns (MJ/ha)	(Energy use efficiency)	productivity (kg /MJ/ha)
Nitrogen level							
Control	13210	44044	66080	110124	96914	8.34	0.227
50% RDN	16870	55203	86818	142022	125152	8.42	0.223
100% RDN	20506	65557	102950	168507	148001	8.22	0.217
150% RDN	24142	69752	109270	179022	154880	7.42	0.197
200% RDN	27778	72543	113954	186497	158719	6.71	0.178
CD (p=0.05)	-	3150	4927	8077	8077	0.56	0.015
Varieties							
PNR-381	20501	52173	81283	133457	112956	6.60	0.176
Rajendra Sweta	20501	53993	84733	138726	118225	6.87	0.182
Pusa Sugandha-5	20501	60593	94464	155058	134557	7.80	0.208
Pusa-44	20501	76614	119459	196074	175573	9.67	0.258
Pusa Sambha-1850	20501	63725	99133	162857	142356	8.16	0.218
CD (p=0.05)	-	4351	6823	11173	11173	0.58	0.015

**Table 3.** Effect of nitrogen levels on energetics of rice genotypes under transplanted condition in calcareous soil (Pooled data of 2 years)

Table 4. Renewable and non-renewable input energy of rice genotypes under different nitrogen levels (Pooled data of 2-years)

Source of energy	Renewal	Renewable and non-renewable input energy (MJ/ha)						Percentage				
	N <sub>1</sub>	N <sub>2</sub>	N <sub>3</sub>	$N_4$	N <sub>5</sub>	$N_1$	$N_2$	$N_3$	$N_4$	$N_5$		
Direct renewable (Human & water)	4547	4571	4571	4571	4571	34	27	22	19	16		
Direct non-renewable (Diesel)	5321	5321	5321	5321	5321	40	32	26	22	19		
Indirect renewable (Seed)	625	625	625	625	625	5	4	3	3	2		
Indirect non-renewable (Machinery, fertilizer, chemicals)	2717	6353	9989	13625	17261	21	38	49	56	62		

N<sub>1</sub>: Control, N<sub>2</sub>: 50% RDN, N<sub>3</sub>: 100% RDN, N<sub>4</sub>: 150% RDN, N<sub>5</sub>: 200% RDN

(158719 MJ/ha) and gross energy output (186497 MJ/ha), comparable to 150% RDN and significantly superior to other nitrogen levels. Comparing 200% RDN with 150% RDN, the latter exhibited the highest EUE (7.42) and energy productivity (0.197 kg/MJ/ha), signifying a more judicious utilization of energy . This discrepancy is attributed to the intensified energy input consumption at higher nitrogen levels.

Within the array of rice varieties examined, Pusa-44 stood out with significantly elevated gross energy output (196074 MJ/ha), net energy returns (175573 MJ/ha), energy use efficiency (9.67), and energy productivity (0.258 kg/MJ/ha). This superiority is attributed to the variety's higher grain and straw yields coupled with lower energy utilization, consistent with findings reported by Kumar et al (2019). Examining different energy sources, the distribution among direct renewable, direct non-renewable, indirect renewable and indirect non-renewable sources varied across nitrogen levels. The percentage share of direct renewable, direct nonrenewable and indirect renewable energy sources decreased from control to 200% RDN. However, the share was increasing from control to 200% RDN (Table 4).

#### CONCLUSION

Based on the findings of the two-year investigation, it is deduced that cultivating the Pusa-44 rice variety in conjunction with 150% recommended dose of nitrogen (RDN) application proves to be a more advantageous and optimal strategy for attaining heightened levels of productivity, profitability, and energetic efficiency in the context of calcareous soils prevalent in the Indo-Gangetic Plains of India.

## **AUTHOR'S CONTRIBUTIONS**

Mohammad Hashim: Conceptualization, writing-original draft; Man Mohan Deo: Statistical Analysis; Sanjeev Kumar: Writing, reviewing and editing. The final version was approved by all authors.

#### references

- Bahuguna A, Singh DK, Supriya, Kumar A, Garg K, Verma P, Patel S and Sudarshan S 2023. Agronomic evaluation of rice (*Oryza* sativa) genotypes under varying fertility levels. *Indian Journal of* Agricultural Sciences 93(11): 1258-1261.
- Cowan N, Bhatia A, Drewer J, Jain, N, Singh R, Tomer R, Kumar V, Kumar O, Prasanna R, Ramakrishnan B, Kumar D, Bandyopadhyay S K, Suttan M and Pathak M 2021. Experimental comparison of continuous and intermittent flooding of rice in relation to methane, nitrous oxide and ammonia emissions and the implications for nitrogen use efficiency and yield. *Agriculture, Ecosystem & Environment* **319**: 107571.
- Danish M, Kumar M, Pramanick B, Singh SK and Hashim M 2022. Enhancing crop productivity, water and nitrogen use efficiency of *kharif* maize through planting methods, mungbean intercropping and nitrogen management practices. *Journal of Cereal Research* 14(1): 97-102.
- Devasenapathy P, Senthil Kumar G and Shanmugam PM 2009. Energy management in crop production. *Indian Journal of Agronomy* **54**(1):80-90.
- Fu Z, Zhang K, Zhang J, Zhang Y, Cao Q, Tian Y, Zhu, Y, Cao W and Liu X 2023. Optimizing nitrogen application and sowing date can improve environmental sustainability and economic benefit in wheat-rice rotation. *Agricultural Systems* 204(C): 103536.
- Hakeem K R, Chandna R, Altaf A and Muhammad I 2012. Physiological and molecular analysis of applied nitrogen in rice genotypes. *Rice Science* **19**: 213-222.
- Hasanuzzaman M, Ali MH, Karim MF, Masum SM and Mahmud JA 2012. Response of hybrid rice to different levels of nitrogen and phosphorus. *International Research Journal of Applied and Basic Sciences* 3(12): 2522-2528.
- Hashim M, Singh VK, Singh KK, Dhar S and Rathore SS 2021. Effect of foliar application of iron and zinc on performance of transplanted rice in middle gangetic plains of Bihar. *Journal of AgriSearch* 8(2): 72-78.
- Houlton BZ, Almaraz M, Aneja V, Austin AT, Bai E, Cassman KG, Compton JE, Davidson EA, Erisman JW, Galloway JN, Gu B, Yao G, Martinelli LA, Scow K, Schlesinger WH, Tomish TP, Wang C and Zhang X 2019. A world of cobenefits: Solving the global nitrogen challenge. *Earth's Future* 7: 865-872.
- Kumar A, Choudhary AK, Pooniya V, Singh VK, Singh U, Singh U, Praharaj CS, Singh SS and Singh N 2016. Soil factors associated with micronutrient acquisition in crops-biofortification perspective (New Delhi: Springer), pp 159-176.
- Kumar R, Mishra JS, Kumar S, Hans H, Bhatt BP, Srivastava AK and Singh S 2019. Production potential, economics and energetics of rice (*Oryza sativa*) genotypes as influenced by varying levels of nitrogen. *Indian Journal of Agricultural Sciences* 89(11): 94-97.
- Kumar R, Mishra JS, Dwivedi SK, Kumar R, Rao KK, Samal SK, Choubey AK and Bhatt BP 2017. Nutrient uptake and content in sorghum cultivars (*Sorghum bicolor L*) under summer environment. *Indian Journal of Plant Physiology* 22(3): 309-315.
- Kumar S, Kumar R, Mishra JS, Dwivedi SK, Prakash V, Rao KK, Singh, AK, Bhatt BP, Singh SS, Haris AA, Kumar V, Srivastava AK, Singh S and Yadav A 2018. Productivity and profitability of rice (*Oryza sativa*) genotypes as influenced by crop management practices under middle Indo-Gangetic Plains. *Indian Journal of Agronomy* **63**(1): 45-49.
- Kumar S, Srinivasa G, Raju M and Mahendra Kumar R 2014. Growth characteristics, yield attributes, grain yield and quality of rice hybrids as influenced by nitrogen fertilization. *Progressive Agriculture* **14**(1): 125-129.
- Kumari P, Devi LL, Kumar, A, Pandey A, Sinha SK and Singh A P 2022. Differential response of rice genotypes to nitrogen availability is associated with the altered nitrogen metabolism

Received 12 July, 2024; Accepted 11 September, 2024

and ionomic balance. *Environmental and Experimental Botany* **198**(3691369):104847.

- Mittal JP and Dhawan KC 1988. Research manual on energy requirements in agricultural sector. ICAR, New Delhi. pp. 20-23.
- Mohapatra SS, Behera SD, Behera B, Sahu G, Jena J, Giri KS, Behera SD, Sahoo RK, Mishra S and Panda M 2024. Yield and quality of aromatic rice (*Oryza sativa*) varieties under nutrient management in organic environment of Coastal Odisha. *Indian Journal of Ecology* 51(3): 552-558.
- Mondal S, Bauri A, Pramanik K, Ghosh M, Malik GC and Ghosh D C 2013. Growth, productivity and economics of rice as influenced by fertility level and plant density. *International Journal of Bioresource and Stress Management* 4(4): 547-554.
- Nadeem F and Farooq M 2019. Application of micronutrients in ricewheat cropping system of South Asia. *Rice Science* **26**(6): 356-371.
- Nayak K, Sar K and Mishra G 2022. Growth and yield of rice (*Oryza sativa* L.) varieties as influenced by nutrient management practices under irrigated-aerobic condition. *Indian Journal of Ecology* **49**(5): 1710-1713.
- Njinju SM, Samejima H, Katsura K, Kikuta M, Gweyi-Onyango J P, Kimani JM, Yamauchi A and Makihara D 2018. Grain yield responses of lowland rice varieties to increased amount of nitrogen fertilizer under tropical highland conditions in central Kenya. *Plant Production Science* **21**(2): 59-70.
- Prasad S, Agrawal KK and Kumar R 2019. Productivity, profitability, quality and nutrient uptake of heat tolerant wheat (*Triticum aestivum*) cultivars as influenced by staggered sowing and nutrition levels. *Indian Journal of Agricultural Sciences* **89**(4): 670-677.
- Renuka DK, Sudhakar P and Sivasankar A 2013. Evaluation of physiological efficiency and yield potential of rice under aerobic condition. *Bioinfolet* **10**(1B): 209-213.
- Ritesh S, Gangwar RK, Yadav V and Kumar R 2014. Response of Basmati (*Oryza sativa*) cultivars to graded nitrogen levels under transplanted conditions. *International Journal of Research in Applied, Natural and Social Sciences* **2**(9): 33-38.
- Salahuddin KM, Chowhdury SH, Muniram S, Islam M and Parvin S 2009. Response of nitrogen and plant spacing of transplanted aman rice. *Bangladesh Journal of Agriculture Research* **34**: 279-285.
- Sheoran OP, Tonk DS, Kaushik LS, Hasija RC and Pannu RS 1998. Statistical Software Package for Agricultural Research Workers. *Recent Advances in information theory, Statistics & Computer Applications* by D.S. Hooda & R.C. Hasija Department of Mathematics Statistics, CCS HAU, Hisar (139-143).
- Singh A, Singh Y, Singh R, Upadhyay PK and Kumar R 2019. Effect of cultivars and weed management practices on weeds, productivity and profitability in ZT direct seeded rice. *Indian Journal of Agricultural Sciences* 89(2): 353-359.
- Singh SS, Singh AK, Kumar S, Mishra JS, Haris AA, Sangle UR, Bhatt BP, Singh SK, Yadav A, Singh US and Singh S 2015. Performance of lentil under rice-lentil under different tillage in drought-prone rainfed ecosystem of Bihar. *Journal of AgriSearch* 2(4): 263-268.
- Sultana R and Islam MK 2014. Performance evaluation of two rice varieties under different levels of salinity stress. *Bangladesh Research Publication Journal* **2**: 61-64.
- Xu P, Chen A, Houlton BZ, Zeng Z, Wei S, Zhao C, Lu H, Liao Y, Zheng Z, Luan S and Zheng Y 2020. Spatial variation of reactive nitrogen emissions from China's croplands codetermined by regional urbanization and its feedback to global climate change. *Geophysical Research Letter* 47, 10.1029/2019GL086551.
- Zidan AA 2017. Performance of some rice genotypes to various nitrogen levels. *Journal of Plant Production,* Mansoura University **8**(1): 103-109.



# Source- Sink Regulation in Red Gram through Foliar Nutrition of Potassium and Growth Promoters in the Warm Humid Tropics of Kerala

# Madiki Aashiq and Sheeba Rebecca Isaac<sup>1</sup>

Department of Agronomy, Agricultural College, Bapatla-522 101, India <sup>1</sup>Regional Agricultural Research Station, Kerala Agricultural University-686 563, India E-mail: aashiqmadiki1997@gmail.com

**Abstract:** The study was conducted to assess the influence of potassium and growth regulators on the source-sink efficiency in red gram [*Cajanus cajan* (L.) Millsp.]. The field experiment was laid out at College of Agriculture, Vellayani during 2019-2021 with combinations of four levels of potassium (0.5 % K<sub>2</sub>SO<sub>4</sub>, 1.0 % K<sub>2</sub>SO<sub>4</sub>, 0.25 % KNO<sub>3</sub>, 0.5 % KNO<sub>3</sub>) and three growth regulators (gibberellic acid 75 mg kg<sup>-1</sup>, brassinosteroid 0.1 mg kg<sup>-1</sup> and humic acid 100 mg kg<sup>-1</sup>) as treatments, along with a control of conventional practice with no foliar nutrition. Foliar application of KNO<sub>3</sub> and GA<sub>3</sub> significantly enhanced the growth attributes. Seed yield (1081.33 kg ha<sup>-1</sup>) and haulm (4837.27 kg ha<sup>-1</sup>) yields were superior with foliar application of KNO<sub>3</sub> (0.5 %) and GA<sub>3</sub>. The interaction effects of KNO3 (0.5 %) x GA3 revealed the significantly highest values for yield attributes and seed yield revealing efficient source- sink regulation compared to the conventional practice of sole nutrient application in soil.

Keywords: Brassinosteroids, Gibberellic acid, Potassium, Red gram, Sink-efficiency

Pulses are climate smart crops can be imbibed in any cropping system and also act as catch crops on account of their deep rooting system, phenotypic plasticity, dehydration tolerance, high moisture retention capacity, and a wide range of photothermal sensitivity (Gull et al 2020). Red gram [Cajanus cajan (L.) Millsp.] belonging to the family Fabaceae is a perennial, woody shrub usually grown as an annual for its edible seeds. It is the second most important pulse crop after chickpea in India. Apart from its nutritional importance, red gram is also valued for its adaptability to arid conditions and is also considered a lifeline of subsistence agriculture. The crop enhances soil fertility by fixing atmospheric nitrogen and contributing organic matter and micronutrients through leaf fall (Bano 2015). Despite the several advantages, red gram is constrained by its low yield which is mainly attributed to its cultivation on poor soils with inadequate and imbalanced nutrient application, excessive vegetative growth, indeterminate growth habit and poor source-sink relationship (Patil et al 2018). Major physiological constraint limiting red gram productivity is abscission of flower and fruits (Kulkarni et al 2019). Among the major nutrients, potassium is of utmost significance, on account of its role in various plant growth functions such as enzyme activity, photosynthesis, mobilization of carbohydrates, fruit set, quality and disease resistance (Prajapati and Modi 2012). Use of plant growth regulators is one of the latest trends in agriculture. Plant growth regulators are chemically synthesized compounds,

which mimic natural hormones in their action and play an important role in shaping plant growth and development. The experiment was formulated to assess the influence of potassium and growth regulators on the source-sink efficiency in red gram.

#### MATERIAL AND METHODS

**Experimental location**: The field experiment was conducted at College of Agriculture, Vellayani, Thiruvananthapuram, Kerala during January to May 2021. The site located at  $8.43^{\circ}$  N latitude,  $76.99^{\circ}$  E longitude and at an altitude of 29 m above mean sea level, enjoys a humid tropical climate with the soil belonging to the order Ultisols. The soil nutrient status revealed medium organic carbon (1.21%), available K (121.6 kg ha<sup>-1</sup>), low available N (257.15 kg ha<sup>-1</sup>) and high available P (103.2 kg ha<sup>-1</sup>) status.

**Experimental details:** The experiment was laid out in randomised block design with 12 combinations of two factors, four levels of potassium ( $a_1 - 0.5 \% K_2SO_4$ ,  $a_2 - 1.0 \% K_2SO_4$ ,  $a_3 - 0.25 \% KNO_3$ ,  $a_4 - 0.5 \% KNO3$ ) and three growth regulators ( $b_1$ – gibberellic acid (GA<sub>3</sub>) 75 mg kg<sup>-1</sup>,  $b_2$  - brassinosteroid (BR) 0.1 mg kg<sup>-1</sup>.  $b_3$  - humic acid (HA) 100 mg kg<sup>-1</sup> as treatments in three replications. A control with recommended dose of nutrients without foliar application was also maintained short duration variety, APK 1 of Regional Research Station (RRS), Aruppokotai, Tamil Nadu was used for the study with an NPK dose of 40: 80: 40 kg ha<sup>-1</sup>

(Devaraj and Isaac, 2021). Foliar sprays were given twice, potassium, at flower bud and pod formation stages and growth regulators, 30 days after sowing (DAS) and at flower bud stage. Urea, rock phosphate and muriate of potash were used as the sources of N, P and K respectively. The entire dose of P was applied as basal, N and K, in two splits, basal and 30 DAS. Threshing and winnowing were done manually to separate seeds. The growth and yield attributes were recorded and the seeds weighed after harvest. The yields per net plot area were used to compute the per hectare yields.

**Statistical analysis:** OP STAT software developed by Chaudhary Charan Singh Haryana Agricultural University was used for data analysis. Control *vs* treatment comparison was statistically analysed as contrast analysis using GRAPES software developed by the Department of Agricultural Statistics, College of Agriculture, Vellayani.

## **RESULTS AND DISCUSSION**

#### **Effect on Growth Characters**

**Plant height:** Foliar application of potassium improved the various growth characters *viz.*, plant height, number of branches, leaf area and LAI in red gram (Table 1). The plant height at 30 DAS did not vary significantly with potassium treatments whereas at 60 DAS, plants were the tallest (94.59 cm) in  $a_4$  (KNO<sub>3</sub> @ 0.5 % application) on par with  $a_3$  (KNO<sub>3</sub> @ 0.25 %) and  $a_1$  (K<sub>2</sub>SO<sub>4</sub> @ 0.5 %). At 90 DAS, the significantly lowest plant height (139.96 cm) was in 0.25 per cent KNO<sub>3</sub> application and the maximum (148.87 cm) was in  $a_4$  (0.5 % KNO<sub>3</sub>). Growth regulators showed non-significant variations at 30 DAS, whereas  $b_1$  (GA<sub>3</sub> @ 75 mg kg<sup>-1</sup>) recorded the tallest plants at 60 and 90 DAS (100.84 and 150.31 cm respectively). It was significantly superior to BR and humic acid application. The interactions exerted significant

influence on plant height at 60 and 90 DAS. The combination,  $a_4b_1$  (KNO<sub>3</sub>@ 0.5 % x GA<sub>3</sub> 75 mg kg<sup>-1</sup>) recorded the maximum plant height (101.42 cm) at 60 DAS and was on par with  $a_1b_1$ ,  $a_3b_1$ ,  $a_2b_1$ , but at 90 DAS, plants were the tallest (153.94 cm) in  $a_4b_1$  and superior to all other combinations. Comparing the treatment effects with the control, at 30 DAS, the effect was non-significant. However, at 60 and 90 DAS, the treatments were significantly superior to the control.

**Number of branches per plant:** Foliar application of potassium sources at different concentrations recorded significant influence on number of branches per plant at flowering (Table 1). Superior number (8.73) was in  $a_4$  (KNO<sub>3</sub> @ 0.5 %) and the lowest number (7.64) in  $a_2$  (K<sub>2</sub>SO<sub>4</sub> @ 1 %). Among growth regulators,  $b_1$  produced the highest number of branches (9.95 per plant) and was significantly superior to BR and humic acid application. Among interactions the maximum number (11.81) was in  $a_4b_1$  and was significantly higher than the other combinations. The lowest number of branches (6.80) was in  $a_3b_2$ . The treatment effects were significantly superior to the control.

Leaf area per plant: Potassium sources applied on the foliage had significant influence on leaf area per plant. The significantly superior leaf area ( $1662.76 \text{ cm}^2$ ) was in  $a_4$  (0.5 % KNO<sub>3</sub>) and the lowest in  $a_2$  ( $1.0 \% \text{ K}_2\text{SO}_4$ ) ( $1430.59 \text{ cm}^2$ ). Growth regulators also had significant influence on leaf area. Leaf area ( $2068.75 \text{ cm}^2$ ) was maximum in  $b_1$  (GA<sub>3</sub>75 mg kg<sup>-1</sup>), followed by BR ( $1296.06 \text{ cm}^2$ ) and humic acid ( $1175.76 \text{ cm}^2$ ) and were markedly different from each other. The treatment interactions could not produce significant effect on leaf area per plant at flowering. However, the treatments were significantly superior to the control.

**Leaf area index:** The superior LAI (2.08) was in  $a_4$  (0.5 % KNO<sub>3</sub>) and the lowest LAI (1.79) in  $a_2$ (1.0% K<sub>2</sub>SO<sub>4</sub> (Tables 1,

Treatments	Pla (Day	nt height /s after sc	(cm) wing)	No. of branches	No. of Leaf area anches (cm <sup>2</sup> per		Number of pods	Average Average pod pod		Seed yield (kg ha <sup>-1</sup> )	Haulm yield (kg ha <sup>-1</sup> )
_	30	60	90	per plant	piant)	Index	per plant	length (cm	) weight (g	)	(купа)
Potassium (a)											
a <sub>1</sub> (0.5% K <sub>2</sub> SO <sub>4</sub> )	48.72	93.96	144.72	7.89	1451.33	1.81	43.57	5.43	0.39	1040.41	4399.17
a <sub>2</sub> (1.0% K <sub>2</sub> SO <sub>4</sub> )	47.51	92.76	144.84	7.64	1430.59	1.79	42.74	5.47	0.38	1006.13	4199.82
a₃ (0.25 % KNO₃)	48.70	93.53	139.96	7.94	1509.41	1.89	43.97	5.44	0.39	1014.44	4140.25
a₄ (0.5 % KNO₃)	48.23	94.59	148.87	8.73	1662.76	2.08	44.42	5.62	0.44	1081.33	4837.27
CD (p=0.05)	NS	1.205	0.730	0.407	124.816	0.156	0.998	0.049	0.023	14.804	189.769
Growth regulators (	(b)										
b <sub>1</sub> (GA <sub>3</sub> )	48.32	100.84	150.31	9.95	2068.75	2.59	48.24	5.64	0.45	1100.13	4780.82
b <sub>2</sub> (BR)	49.25	91.09	144.82	7.07	1296.06	1.62	45.37	5.37	0.38	1072.74	4618.95
b <sub>3</sub> (HA)	47.30	88.65	134.48	7.30	1175.76	1.47	37.41	5.46	0.37	934.02	3781.86
CD (p=0.05)	NS	1.043	0.633	0.352	108.094	0.135	0.864	0.042	0.020	12.821	164.345

 Table 1. Effect of potassium and growth regulators on growth and yield in red gram

2). Among growth regulators, LAI was maximum (2.59) in  $b_1$  (GA<sub>3</sub>75 mg kg<sup>-1</sup>). It was significantly superior to BR and humic acid application. Interactions could not exhibit any significant effect on LAI. Effects of treatments were superior to control of no foliar applications.

Potassium in plants has regulatory roles in numerous plant physiological processes viz. seed germination and emergence, stomatal regulation, phloem transport, cationanion balance, protein synthesis, photosynthesis, energy transfer, osmoregulation, enzyme activation, nutrient balance, and stress resistance (Marschner 2012). Foliar application over and above the recommended dose in soil had beneficial effects on the growth attributes in red gram. Potassium nitrate was found to be superior to K<sub>2</sub>SO<sub>4</sub>. Nitrogen is pivotal in promoting leaf growth, aids in photosynthesis and increases protein content (Olivar et al 2014). The significant increase in the plant height, number of branches and leaf area per plant might be due to the fact that nitrate of potash provided both potassium and nitrogen, two of the major nutrients that are crucial for crop growth. The positive interaction of the two nutrients within the plant when applied with the same source would also have bestowed favourable effect on growth.

Red gram being a leguminous crop is capable of nitrogen fixation which ensures the availability of the nutrient for the crop during its growth. However, after flowering the nodules are known to degrade that leads to a lowered nitrogen supply, and as vegetative and reproductive growth occurred simultaneously, this had an impact on the growth attributes. The foliar nutrition with KNO<sub>3</sub> at this juncture ensured an adequate supply of both the nutrients for proper growth, balancing the source and sink development. This is supported by the reports of Lenka (2020) which elucidate that, as the active nodulation of pulse crop stops after 45 to 50 DAS, a foliar feeding of fertilizers containing nitrogen helps in providing supplementary nitrogen to the crop, enhancing growth and yield. Thalooth et al (2006) observed that potassium and nitrogen added via KNO, augmented cell division and cell expansion in plants thereby stimulating the biological activity of photosynthetic pigments and enzymes, which in turn promoted the vegetative growth. The increment in the plant height and number of branches is possibly due to the favourable effects of GA<sub>3</sub> on cell division and internodal elongation. Similar results were elucidated by Giri et al (2018). The synergistic effect of gibberellin (GA) and potassium in combination has been reported by Dev et al (2020). The treatment effects on growth were superior to the control as the additional supply and effective absorption of the nutrients (nitrogen/sulphur and potassium) via foliar application and the inclusion of growth promoting substances favoured growth and growth attributes.

#### Yield Attributes and Yield

**Number of pods per plant:** The maximum number of pods (44.42) was in  $a_4$  (0.5 % KNO<sub>3</sub>) and comparable with  $a_1$  and  $a_3$ ,

Treatments	Pla (Day	int height /s after so	(cm) wing)	No. of branches	Leaf area (cm <sup>2</sup> per	Leaf area	Number of pods	Average pod	Average pod	Seed yield (kg ha <sup>-1</sup> )	Haulm yield
_	30	60	90	per plant	piant)	Index	per plant	iengin (cm)	) weight (g)		(kg na )
a <sub>1</sub> b <sub>1</sub>	47.06	100.87	149.86	9.81	1986.88	2.48	48.74	5.72	0.48	1114.67	4760.33
<b>a</b> <sub>1</sub> <b>b</b> <sub>2</sub>	51.66	91.17	150.05	6.91	1223.04	1.53	44.12	5.19	0.34	1071.83	4589.85
a <sub>1</sub> b <sub>3</sub>	47.46	91.72	134.27	6.92	1144.07	1.43	37.86	5.37	0.35	934.83	3874.33
<b>a</b> <sub>2</sub> <b>b</b> <sub>1</sub>	46.28	99.98	150.10	8.89	1877.99	2.35	46.49	5.75	0.49	1064.57	4214.60
$a_2b_2$	48.49	89.63	146.53	6.91	1287.11	1.61	44.37	5.43	0.35	1019.70	4489.06
a <sub>2</sub> b <sub>3</sub>	47.77	88.69	138.00	7.11	1126.68	1.41	37.37	5.25	0.33	932.53	3892.80
<b>a</b> <sub>3</sub> <b>b</b> <sub>1</sub>	49.60	101.10	147.33	9.28	2144.47	2.68	47.64	5.30	0.34	1071.83	4333.30
$a_3b_2$	47.17	91.76	136.90	6.80	1212.31	1.51	46.15	5.40	0.41	1034.20	4525.27
a <sub>3</sub> b <sub>3</sub>	49.32	87.73	135.65	7.73	1171.44	1.47	38.15	5.63	0.41	937.13	3562.19
a₄b₁	50.36	101.42	153.94	11.81	2265.67	2.84	50.11	5.78	0.51	1186.33	5815.05
$a_4b_2$	49.67	91.81	145.80	7.66	1461.77	1.82	46.84	5.46	0.42	1127.60	4871.63
a₄b₃	46.68	88.65	130.02	7.45	1260.84	1.58	36.30	5.61	0.42	930.10	3825.13
CD (p=0.05)	NS	2.087	1.265	0.704	NS	NS	1.728	0.084	0.040	25.642	328.689
Control (C)											
C (RDF)	46.78	84.33	120.55	6.05	1076.60	1.34	34.14	5.16	0.32	916.7	4049.5
Treatment vs Control	NS	S	S	S	S	S	S	S	S	S	S

Table 2. Interaction effect of potassium and growth regulators on growth and yield in red gram.

but superior to  $a_2$  (42.74). The growth regulators,  $b_1$  (GA<sub>3</sub> @ 75 mg kg<sup>-1</sup>) recorded the highest number of pods per plant (48.24) and was significantly superior to BR and humic acid application. Interaction of potassium and growth regulators exerted significant influence on the number of pods per plant, with the maximum number of pods (50.11) in  $a_4b_1$ , on par with  $a_1b_1$  (48.74). Pod number was the lowest number (36.30) in  $a_4b_3$ . The treatments were found to be significantly higher than control.

Average pod length: The significantly longest pods (5.62 cm) were in  $a_4$  and the shortest pods (5.43 cm) in  $a_1$ . Variations in pod length with growth regulators were also significant. GA<sub>3</sub> application proved superior with the longest pods (5.64 cm). In response to the interactive effects, pods were the longest (5.78 cm) in  $a_4b_1$  and was on par with  $a_1b_1$  (5.72 cm) and  $a_2b_1$  (5.75 cm) whereas, the shortest pods (5.19 cm) was in  $a_1b_2$ . Except in combination with 0.25 percent KNO<sub>3</sub>,  $b_1$  recorded the longest pods with  $a_1$ ,  $a_2$  and  $a_4$ . The treatments were significantly superior to control.

**Average pod weight:** Pod weight was significantly higher in  $a_4$  (0.5 % KNO<sub>3</sub>), GA<sub>3</sub> application (b<sub>1</sub>) and the combination  $a_4b_1$  (0.51 g). The lowest weight (0.33 g) was in  $a_2b_3$ . The treatments were significantly superior to control.

**Seed yield:** The highest yield (1081.33 kg ha<sup>-1</sup>) was with 0.5 % KNO<sub>3</sub> spray. The higher concentration of K<sub>2</sub>SO<sub>4</sub> (1.0 %) recorded the lowest yield (1006.13 kg ha<sup>-1</sup>). Among growth regulators, maximum seed yield (1100.13 kg ha<sup>-1</sup>) was in b<sub>1</sub> (GA<sub>3</sub> 75 mg kg<sup>-1</sup>) which was significantly superior to the BR and humic acid treatments. Seed yield was the lowest (934.02 kg ha<sup>-1</sup>) in b<sub>3</sub> (HA 100 mg kg<sup>-1</sup>). Interactions indicated significant variations with the maximum yield (1186.33 kg ha<sup>-1</sup>) in the combination of 0.5 % KNO<sub>3</sub> and GA<sub>3</sub> (a<sub>4</sub>b<sub>1</sub>) and was significantly superior. The lowest yield (930.10 kg ha<sup>-1</sup>) was in a<sub>4</sub>b<sub>3</sub>. The treatment mean was significantly superior to control.

**Haulm yield:** Haulm yields followed similar trends of growth and seed yields. The highest haulm yield (4837.27 kg ha<sup>-1</sup>) was in  $a_4$  (0.5 % KNO<sub>3</sub>) and the lowest (4140.25 kg ha<sup>-1</sup>) in  $a_3$ (0.25 % KNO<sub>3</sub>). The effect of growth regulators was also significant, the highest haulm yield (4780.82 kg ha<sup>-1</sup>) was recorded in  $b_1$  (GA<sub>3</sub>75 mg kg<sup>-1</sup>) but statistically comparable to that in  $b_2$ . Among the treatment combinations,  $a_4b_1$  recorded the highest haulm yield (5815.05 kg ha<sup>-1</sup>) in line with the individual effects and was significantly superior to other treatment combinations. The lowest haulm yield (3562.19 kg ha<sup>-1</sup>) was recorded in  $a_3b_3$ . The significant difference between treatments means and control was also evident.

The indeterminate growth habit of red gram ensues a continuous competition for available assimilates between vegetative and reproductive sinks throughout the growth period. In pulses, yield is in general highly constrained by the lowered translocation of assimilates to the growing reproductive sinks. Insufficient partitioning of assimilates, poor pod setting due to flower abscission and lack of nutrients during critical stages of crop growth are crucial. In the present study, potassium nutrition was found to favourably influence the sink efficiency in red gram. Foliar application of KNO<sub>3</sub> at 0.5 per cent concentration was found to be superior. The results accords the reports on efficient utilisation of nutrients thereby reducing flower shedding and enhancing yields (Sathishkumar et al 2020, Singh et al 2021). The increased leaf area indicated higher source strength, increased photosynthesis and hence, higher photo assimilate production. Foliar application of potassium enhanced the rapid availability of the nutrient; redistribution of carbohydrates and translocation of assimilates to the economically important parts, the sink. In addition, potassium aids in acceleration of the physiological processes within, resulting in improved plant growth and yield attributes, and these were manifested in the seed yields. It is deciphered that the supply of potassium and nitrogen to supplement the soil contribution was effective in enhancing the production by virtue of the increased translocation of assimilates to the sinks, and a higher pod set, nearly 20 per cent greater than the lowest (24.18%) among potassium treatments. The impacts on the leaf area, number and weight of pods would have attenuated the higher yields.

Growth regulators had significant influence on average pod weight, seed and haulm yields and the values were superior in GA application which could be attributed to the significant role in stimulating physiological responses and altering the source-sink metabolism through their effect on photosynthesis and sink formation. Gibberellic acid is closely related to cell division and enlargement during fruit development (Zhang et al 2007) and the exogenous application increased seed weights. Application at 30 DAS and at pod formation stage might have improved the vegetative and reproductive development of red gram crop and supported efficient translocation of photosynthates from source to sink, as illustrated by Kumar and Sharma (2021). The increase in sink strength, number of pods, might be due to increased leaf area which leads to better utilization of sunlight and the plants remaining physiologically more active to build up sufficient food material for developing more number of pods. The enhanced sink potential and capacity through efficient translocation of photosynthates from source to sink with GA<sub>3</sub> application would also have complimented the higher yields. The results of the experiment are in line with the reports of Giri et al (2018) Kumar et al (2018) and Bhanu et al (2024).

#### CONCLUSION

The foliar application of KNO<sub>3</sub> and GA<sub>3</sub> significantly enhanced growth and seed yields in red gram. Growth attributes that represent the source strength, and seed yields, the sink capacity, were significantly superior with KNO<sub>3</sub> @ 0.5 % and GA<sub>3</sub> application. The individual effects were reflected in the interaction effects of KNO<sub>3</sub> (0.5 %) x GA<sub>3</sub> also. Therefore, source- sink regulation through the foliar applications of KNO<sub>3</sub> (0.5 %) at flower bud and pod formation stages and GA<sub>3</sub> (75 mg kg<sup>-1</sup>), 30 DAS and at flower bud stage, proved superior and can be recommended for cultivation of short duration red gram in the warm humid tropics of Kerala.

#### ACKNOWLEDGEMENT

Authors express their sincere gratitude to Kerala Agricultural University for the financial assistance and physical support provided for the M.Sc. thesis of the first author.

#### REFERENCES

- Bano DA, Singh RK, Singh NP and Waza SA 2015. Effect of cowpea bradyrhizobium (RA-5) on growth parameters of pigeon pea plant under various salt concentrations at different time intervals. *Indian Journal of Ecology* 42 (1): 179-182.
- Bhanu VS, Nimbolkar PK, Wangchu L and Siddhartha SR 2024. Influence of plant growth regulators, urea and micronutrient on growth, yield and quality of Rangpur Lime (*Citrus limonia Osbeck*). *Indian Journal of Ecology* **51**(2): 327-332.
- Dev R, Dayal D and Sureshkumar M 2020. Gibberellic acid and potassium nitrate promote seed germination and growth of greyleaved saucer-berry (*Cordia sinensis* Lam.) seedlings. *International Journal of Fruit Science* **20**: 937-954.
- Devaraj GA and Isaac SR 2021. Production potential of short duration redgram [*Cajanus cajan* (L.) Millsp.] in the Southern Laterites of Kerala. *Legume Research* **4361**: 1-5.
- Giri MD, Jaybhaye CP, Kanwade DG and Tijare B 2018. Effect of foliar application of gibberellic acid on pigeonpea [*Cajanus cajan* (L.)] under rainfed conditions. *Journal of Pharmacognosy and Phytochemistry* 7(2): 617-620.
- Gull R, Bhat TA, Sheikh AT, Wani OA, Fayaz S, Nazir A, Saad AA, Jan A, Zazir I and Nisah RU 2020. Climate change impact on

Received 12 July, 2024; Accepted 01 October, 2024

pulse in India: A review. Journal of Pharmacognosy and Phytochemistry 9(4): 159-166.

- Kulkarni P, Muniswamy S, Lokesha R, Girish G, Suma TC and Mahaling DM 2019. Floral abscission in pigeonpea (*Cajanus cajan* (L.) Millsp.): genotypic disparity in land races, hybrids and advanced breeding lines. *International Journal of Pure & Applied Bioscience* 7(1): 389-397.
- Kumar D, Singh RP, Somasundaram J, Simaiya V and Jamra S 2018. Effect of foliar application of nutrients on growth and development of blackgram [*Vigna mungo* (L.) Hepper] under rainfed vertisols of Central India. *International Journal of Chemical Studies* 6(1): 609-613.
- Kumar R and Sharma SC 2021. Influence of foliar application of gibberellic acid on growth, yield and economics of pigeonpea (*Cajanus cajan* L.). *Biological Forum* **13**(1): 227-231.
- Lenka B 2020. Effect of foliar nutrition of water-soluble fertilizers on pulse crop productivity [online] Available: https://www.research gate.net/publication/340983430\_Effect\_of\_Foliar\_Nutrition\_of\_ Water\_Soluble\_Fertilizers\_on\_Pulse\_Crop\_Productivity. [05 Dec. 2021].
- Marschner H 2012 *Marschner's Mineral Nutrition of Higher Plants*. (3<sup>rd</sup> Ed.). Academic Press, London, 672p.
- Olivar VT, Torres OGV, Patino MLD, Nava HS, Martinez AR, Aleman RMM, Aguilar LAV and Iran 2014. Role of nitrogen and nutrients in crop nutrition. *Journal of Agriculture, Science and Technology* **4** (2): 29-37.
- Patil JR, Thakur V and Raju T 2018. Influence of foliar application of pulse magic on seed yield and economics of pigeonpea grown under North Eastern Dry zone of Karnataka. *Advances in Research* **15**(3): 1-5.
- Prajapati K and Modi HA 2012. The importance of potassium in plant growth: A review. *Indian Journal of Plant Sciences* 1(02-03): 177-186.
- Sathishkumar A, Sakthivel N, Subramanian S and Rajesh P 2020. Productivity of field crops as influenced by foliar spray of nutrients: A review. *Agricultural Reviews* **41**(2): 146-152.
- Singh K, Kumar S and Kaur C 2021. Effect of foliar application of water soluble fertilizers on growth and yield of chickpea (*Cicer* arietinum L.) Indian Journal of Agricultural Research 55(5): 639-642.
- Thalooth AT, Tawfik MM and Mohamed HM 2006. A comparative study on the effect of foliar application of zinc, potassium and magnesium on growth, yield and some chemical constituents of mung bean plants grown under water stress conditions. *World Journal of Agricultural Sciences* **2**(1): 37-46.
- Zhang C, Tanabe K, Tamura FF and Itai A 2007. Role of gibberellins in increasing sink demand in japanese pear fruit during rapid growth. *Plant Growth Regulation* **52**(2): 161-172.



# Impact of Intercropping with Black Gram on Incidence of Stem Fly, *Melanagromyza sojae* (Zehntner) during the *Kharif* Season in Gujarat

N.P. Pathan, D.B. Sisodiya<sup>1</sup> and R.D. Dodiya<sup>2</sup>

Department of Plant Protection, College of Horticulture, S.D. Agricultural University, Jagudan-384 460, India Department of Entomology, BACA, Anand Agricultural University, Anand-388 110, India Department of Entomology, CPCA, S.D. Agricultural University, Sardarkrushinagar-385 506, India E-mail: naziya.p.pathan@sdau.edu.in

**Abstract:** Investigation on impact of intercropping with black gram on incidence of stem fly, *Melanagromyza sojae* (Zehntner) were carried out at Anand Agricultural University, Anand during *kharif* season (2017 and 2021). Black gram intercropped with either maize or sorghum (3:2) proved superior over rest of intercropping systems with lower infestation (22.20 %), tunnelling (8.00 %), larval population (0.14 /plant) and pupal population (0.19 /plant) of stem fly, *M. sojae*. Higher black gram equivalent seed yield was registered in all the treatments of intercropping systems *i.e.*, black gram + maize (3:2 and 5:1 ratio) and black gram + sorghum (3:2 and 5:1 ratio) in comparison to black gram as sole crop.

Keywords: Black gram, Maize, Sorghum, Intercropping and stem fly

Pulse crops are hugely important in India since they help with crop diversity, revenue production, food security, sustainable agriculture, resilience to climate change, and export potential. One of the main pulse crops farmed in India is black gram. In India, the total production of black gram is 30,59,990 tons with 546 kg/ha productivity from an area of 56,02,470 ha in 2018-19 (Anonymous 2019). On an average, 2.5 to 3.0 million tonnes of pulses are lost annually due to pest problems (Rabindra et al 2004). In India, 60 insect species are known to attack black gram at different stages of crop growth. Yield loss due to stem fly varies between locations and according to the plant growth stage. Gaur et al (2015) reported 100% infestation and 33.84% stem tunnelling caused by *M. sojae* in soybean at Pant Nagar in Uttarakhand. Pathan et al (2023a) from Anand, Gujarat reported that Melanagromyza sojae (Zehntner) severely damages black gram at the seedling stage and exhibited a highly significant positive association with bright sunshine hours and maximum temperature. The black gram crop sown during the fourth week of August demonstrated the highest level of infestation (53.68%) and tunnelling (15.44%) followed by the crop sown during the third week of August (Pathan et al 2023c). The crop sown late i.e. 1<sup>st</sup> week of April showed significantly higher infestation (64.28%) of stem fly (Pathan et al 2022b). In the Gujarat region, the dipteran insect species M. sojae (Diptera: Agromyzidae) has emerged as a novel pest affecting black gram crop. Stem fly infestation exhibited a highly significant positive association (r = 0.519\*\*

and  $0.655^*$ ) with bright sunshine hours (BSS) and maximum temperature (MaxT), respectively in *kharif* (Pathan et al 2023b) while minimum temperature (r =  $0.769^{**}$ ) and evening during summer respectively in summer (Pathan et al 2022a). SKNU-11-11 proved to be susceptible genotype against *M. sojae* (Pathan and Sisodiya 2023, Pathan et al 2023d). Presently, there exists a deficiency of effective pest management strategies beyond the application of chemical insecticides. Given this scenario, it becomes imperative to explore and implement eco-friendly sustainable alternatives for insect pest control such as adopting cultural practices like intercropping. The comprehensive investigations were conducted to assess the influence of intercropping involving black gram on the incidence of the stem fly, *M. sojae* during the *kharif* season.

#### MATERIAL AND METHODS

In order to study the effect of intercropping on the incidence of stem fly, maize and sorghum was used as an intercrop with black gram. The experiment was conducted during *kharif*, 2017 and 2021. The variety of maize *i.e.* GAWMH-2 and for sorghum GNJ-1 (Gujarat Navsari Jowar 1) were used as an intercrop. There were four intercropping combinations *viz*, black gram + maize (5:1), black gram + maize (3:2), black gram + sorghum (5:1), black gram + sorghum (3:2) and sole black gram. The experiment was laid out in a randomized block design with four replication at Anand Agricultural University, Anand. Blackgram (Gujarat

Urad 1) were sown with the spacing of  $45 \times 10$  cm with the plot size of  $2.70 \times 5.00$  m.

For stem fly infestation, ten randomly selected seedlings were uprooted from each plot and brought to the departmental laboratory. The roots were gently washed in tap water to remove adhering soil. Stem of each plant was dissected and observations on the length of the stem, length of the tunnel and the number of larva (e) and pupae (e) present in the stem were recorded. The number of stem fly infested plants in each sample was also recorded at weekly interval starting from one week after germination.

**Statistical analysis**: The data were analysed by following standard statistical technique (Steel and Torrie 1980). Crop equivalent yield tunnelling and infestation per cent were calculated (Laxmigudi et al 2014). The software GWBASIC command prompt was used.

## **RESULTS AND DISCUSSION**

**Pest infestation (%):** The impact of intercropping on pest infestation was significant as the treatments differed significantly (Table 1). The black gram crop intercropped either with maize or sorghum (3:2) registered 22.10 and 23.43% infestation of stem fly in black gram crop, respectively. Intercropping of black gram either maize or sorghum (5:1) exhibited 36.55 to 38.04% infestation of *M. sojae* but was at par. The black gram crop sown alone showed significantly higher infestation of the pest (51.33%). **Tunnelling (%):** The black gram intercropped with maize

(3:2 ratio) and sorghum (3:2) registered 7.63 and 7.95% tunnelling, respectively. These cropping systems proved best in significant reduction of tunnelling (%) due to stem fly in black gram. The intercropping systems of black gram with either maize or sorghum (5:1) also proved moderately effective but found significantly better than black gram crop grown alone. Significantly highest tunnelling was in sole black gram crop over the rest of the treatments (16.02%).

**Number of larva:** The black gram intercropped with maize or sorghum (3: 2) exhibited significantly low larval population (0.14 to 0.16 larva/ plant) over other treatments. Larval numbers were also suppressed moderately where black gram was intercropped with either maize or sorghum (5:1), but it was significantly better over black gram grown as sole crop.

**Number of pupa:** The pupal population of stem fly, *M. sojae* in black gram reduced significantly when it was intercropped either with maize (3:2) or sorghum (3:2). Promising effect of these intercropping systems also observed when it was evaluated as 5:1 ratio. Black gram crop grown as sole crop exhibited significantly highest (0.87 pupa/plant) pupal count over rest of the treatments of intercropping systems.

**Yield:** The maximum seed yield was harvested in black gram + maize (3:2) treatments (562 kg/ha) followed by black gram + sorghum (3:2), black gram + maize (5:1) and black gram + sorghum (5:1). All these treatments were at par and registered significantly higher yield in comparison to black gram as sole crop (460 kg/ha). The black gram intercropped either with maize or sorghum in 3:2 ratio yielded significantly

Treatments	Infestation (%)	Tunneling (%)	Larva (e)/plant	Pupa (e)/plant
Black gram + Maize (5:1)	37.20 (36.55)	20.22 (11.95)	0.94 (0.38)	0.99 (0.48)
Black gram + Maize (3:2)	28.04 (22.10)	16.43 (8.00)	0.80 (0.14)	0.83 (0.19)
Black gram + Sorghum (5:1)	38.08 (38.04)	20.66 (12.45)	0.97 (0.44)	1.02 (0.54)
Black gram + Sorghum (3:2)	28.95 (23.43)	16.79 (8.34)	0.81 (0.16)	0.84 (0.21)
Black gram as sole crop	45.76 (51.33)	23.59 (16.02)	1.10 (0.71)	1.17 (0.87)
CD (p=0.05) T	1.17	0.52	0.03	0.03
Р	1.81	0.80	0.03	0.03
Y	0.74	0.32	0.02	0.02
ТхР	NS	NS	NS	NS
ТхҮ	NS	NS	NS	NS
PxY	NS	NS	NS	NS
Τ x Ρ x Υ	NS	NS	NS	NS
C.V. (%)	13.83	11.18	8.30	8.52

Table 1. Effect of intercropping on incidence of stem fly, M. sojae infesting black gram (Pooled: 2017 and 2021)

Figures in parentheses are retransformed values and those outside are arc sine transformed values, NS = Not Significant

Treatments	Yield (kg/ha)									
-	Black gram		Ν	laize	Sorghum		Equivalent yield			
	Seed	Haulm	Seed	Dry fodder	Seed	Dry fodder	Seed	Haulm		
Black gram + Maize (5:1)	490	638	164	225	-	-	540	936		
Black gram + Maize (3:2)	465	548	319	392	-	-	562	1070		
Black gram + Sorghum (5:1)	496	624	-	-	126	159	534	834		
Black gram + Sorghum (3:2)	470	570	-	-	275	332	553	1013		
Sole black gram	461	682	-	-	-	-	460	681		
CD (p=0.05) T	-	-	-	-	-	-	34.74	75.45		
Y	-	-	-	-	-	-	NS	NS		
ТхҮ	-	-	-	-	-	-	NS	NS		
Black gram = ₹63 /kg	М	Price aize grain = ₹	19 /Ka	So	rahum araiı	n = ₹ 27 /Ka <sup>.</sup>				

Table 2. Yield of maize and sorghum intercropped with black gram (Pooled: 2017 and 2021)

Black gram haulm = ₹ 1.5 /Kg;

Maize fodder = ₹ 2 /kg;

Sorghum fodder = ₹ 2/kg

## Table 3. Economics of different intercropping systems evaluated in black gram (Pooled: Kharif 2017 and 2021)

Treatments	Yield (kg/ha)						Cost of	Income	Gross	Net	ICBR
	Black gram		Maize		Sorghum		treatment (₹/ha)	( <th>income (₹/ha)</th> <th>pront (₹/ha)</th> <th></th>	income (₹/ha)	pront (₹/ha)	
	Seeds	Haulm	Grain	Fodder	Grain	Fodder	_				
Black gram + Maize (5:1)	490	638	164	225	-	-	2847	35393	5327	2480	1:1.87
Black gram + Maize (3:2)	465	548	319	392	-	-	2865	36962	6896	4031	1:2.41
Black gram + Sorghum (5:1)	496	624	-	-	126	159	2229	35904	5838	3609	1:2.62
Black gram + Sorghum (3:2)	470	570	-	-	275	332	1629	38554	8488	6859	1:5.21
Sole black gram	461	682	-	-	-	-	2829	30066	-	-	-

See Table 2 for details

higher haulm yield than rest of the treatments. Black gram + maize and black gram + sorghum (5:1) produced 936 and 834 kg/ha haulm yield, respectively.

Economics: The maximum (₹ 6859 /ha) net profit was in black gram + sorghum (3:2) followed by black gram + maize (5:1). The black gram + sorghum (3:2) registered highest ICBR (1:5.27) followed by the treatment of black gram + sorghum (5:1) wherein it was 1:2.62 ICBR (Table 3).

The black gram crop intercropped either with maize or sorghum (3:2 ratio) registered low infestation of stem fly, M. *sojae* in black gram. Similarly, intercropping system of black gram with either maize or sorghum at 5:1 ratio was also promising in suppressing the infestation of stem fly. This finding is strongly supported by the report of Prodhan et al (2000) and Pathan et al (2024) where black gram crop intercropped with maize significantly reduced larval and pupal population, stem tunnelling, infested plants and produced higher yield over sole black gram crop. Peter et al (2009) observed that the population of Ophiomia spp. was higher in pure stands of common beans in Tanzania than in the intercrop with maize. Low counts of larvae and pupae were recorded in intercropping system.

#### CONCLUSION

The black gram crop intercrop with maize (3:2) proved superior over rest of the other treatments followed by black gram + sorghum (3:2) intercropping system by recording less number of larvae, pupae as well as lower percent infestation and tunnelling. Intercropping black gram with either maize or sorghum (5:1) suppressed the pest at moderate level. Black gram crop grown alone exhibited maximum infestation. Relatively higher black gram equivalent seed yield was registered in all the treatments of intercropping in comparison to seed yield recorded from black gram sown as sole crop.

#### REFERENCES

- Anonymous 2019. Season-wise area, production and productivity of Urad in India. India stat. Retrieved from:http://www. indiastat.com.
- Gaur N, Sharma P and Nautiyal A 2015. Seasonal incidence of major insect pests of soybean and their correlation with abiotic factors. Journal of Hill Agriculture 6(1): 75-78.
- Jagadish KS, Gowda G and Lakshmikantha BP 1995. Effect of intercropping on the incidence of stem fly, Ophiomyia phaseoli (Tryon) (Diptera: Agromyzidae) in cowpea [Vigna unguiculata (L.) Walp.]. Crop Research (Hisar) 10(1): 80-84.

Justin GLC, Anandhi P and Jawahar D 2015. Management of major

insect pests of black gram under dryland conditions. *Journal of Entomology and Zoology Studies* **3**(1): 115-121.

- Karel AK 1991. Effect of plant population and intercropping on the population patterns of bean flies on common beans. *Environmental Entomology* **20**(1): 354-357.
- Kayitare JS 1993. Infestation on Phaseolus vulgaris (L) by the bean fly Ophiomyia spp. (Diptera: Agromyzidae) and its management by cultural practices. Ph.D. thesis submitted to the Department of Crop Science of the faculty of Agriculture, University of Ghana, Legon.
- Laxmigudi R, Gopali JB, Hosamani A and Yelshetty S 2014. Estimation of avoidable loss due to stem fly, *Ophiomyia phaseoli* (Tryon) and its management by using new molecules as seed dressers in green gram. *Karnataka Journal of Agricultural Sciences* 27(1): 32-35.
- Pathan NP and Sisodiya DB 2023. Screening of different genotypes/cultivars of black gram against stem fly, *Melanagromyza sojae* (Zehntner) in *Kharif* season. *Journal of Agriculture and Ecology* **17**: 53-57.
- Pathan NP, Sisodiya DB and Patel RM 2024. Significance of intercropping with black gram on incidence of stem fly, *Melanagromyza sojae* (Zehntner) in summer season. *Journal of Eco-friendly Agriculture* **19**(2): 405-410.
- Pathan NP, Sisodiya DB and Bhatt NA 2023d. Screening of different genotypes/cultivars of black gram against stem fly, *Melanagromyza sojae* (Zehntner) in summer season. *International Journal of Statistics and Applied Mathematics* SP-8(6): 1278-1282.
- Pathan NP, Sisodiya DB, Dodiya RD and Kalola AD 2023b. Seasonal incidence of stem fly, *Melanagromyza sojae* (Zehntner) infesting

Received 29 June, 2024; Accepted 21 September, 2024

black gram (Vigna mungo L.) in kharif. Journal of Agriculture and Ecology **15**: 94-99.

- Pathan NP, Sisodiya DB, Dodiya RD and RK Thumar 2023c. Impact of sowing periods on incidence of stem fly, *Melanagromyza sojae* (Zehntner) in *kharif* black gram. *The Pharma Innovation Journal* **12**(6): 6260-6265.
- Pathan NP, Sisodiya DB and Raghunandan BL 2023a. First Report of Stem Fly *Melanagromyza sojae* (Zehntner) Infesting Black Gram (*Vigna mungo* L.) in India. *Biological Forum* – *An International Journal* **15**(4): 637-642.
- Pathan NP, Sisodiya DB, Gohel NM and Mohapatram AR 2022b. Impact of sowing periods on incidence of stem fly, *Melanagromyza sojae* (Zehntner) in summer black gram. *The Pharma Innovation Journal* SP-**11**(9): 783-787.
- Pathan NP, Sisodiya DB and Mohapatram AR 2022a. Population dynamics of stem fly, *Melanagromyza sojae* (Zehntner) infesting black gram (*Vigna mungo* L.) in summer season. *The Pharma Innovation Journal* SP-**11**(8): 2118-2121.
- Peter KH, Swella GB and Mushobozy DMK 2009. Effect of plant population on the incidence of bean stem maggot (*Ophiomyia* spp.) in common bean intercropped with maize. *Plant Protection Science* **45**: 148-155.
- Prodhan MZH, Haque MA and Hossain MA 2007. Stem fly management in black gram through intercropping. *Annals of Bangladesh Agriculture* **11**(1):67-75.
- Rabindra RJ, Ballal CR and Ramanujan B. 2004. *Biological options* for insect pests and nematode management in pulses. Kalyani Publishers, New Delhi, India, p. 487.
- Steel RGD and Torrie JH 1980. *Principles and procedures of statistics*. Publ. McGraw Hill Book Company, New York, USA, p. 137.



# Effect of Date of Sowing and Varieties on Performance of Summer Fodder Pearlmillet (*Pennisetum glaucum* L.) under North Gujarat Condition

# Manisha M. Prajapati, Vikash Kumar\*, J.K. Patel<sup>1</sup> and Veeresh Hatti

Department of Agronomy, <sup>1</sup>Department of Agricultural Chemistry and Soil Science, C. P. College of Agriculture, S. D. Agricultural University, Sardarkrushinagar-385 506, India \*E-mail: vky.iari@gmail.com

**Abstract:** Field experiment was conducted on loamy sand soil at S. D. Agricultural University, Sardarkrushinagar, Gujarat during summer season of 2021. The experiment was laid out in split plot design with fifteen treatment combinations replicated thrice comprised of three dates of sowing in main plots *viz.* 25<sup>th</sup> February, 12<sup>th</sup> March and 27<sup>th</sup> March and five varieties in sub plots *viz.* Gujarat Fodder *Bajra* 4, Gujarat Fodder *Bajra* 1, BAIF *Bajra* 1, TSFB 15-8 and Moti *Bajra.* Crop sown on 12<sup>th</sup> March recorded significantly higher total green fodder and dry fodder yield *i.e.*, 589.8 and 139.2 q/ha, respectively. The 25<sup>th</sup> February sown crop was statistically at par with the crop sown on 12<sup>th</sup> March. Among the varieties, Gujarat Fodder Bajra 4 recorded significantly higher total green fodder yield (625.6 q/ha) and dry fodder yield (148.5 q/ha). TSFB 15-8 was statistically at par with Gujarat Fodder *Bajra* 4. The maximum B: C ratio *i.e.*, 2.25 was obtained under 12<sup>th</sup> March sown crop as compared to other dates of sowing. In case of varieties, maximum B: C ratio *i.e.*, 2.39 was obtained in Gujarat Fodder *Bajra* 4.

#### Keywords: Date of sowing, Economics, Pearlmillet, Quality, Varieties, Yield

Livestock is becoming agriculture's most economically important sub sector and contributes 25% to the total agricultural income. One of the major reasons for low animal productivity in India is the shortage as well as poor quality of fodder. In Gujarat, total animal population is about 18.4 million and their optimum fodder requirement is 42.2 million tonnes, whereas, the availability is of only 20.0 million tonnes of forage during normal year (Pareek et al 2015). Cereals have major role to play in the fodder supply. Four major cereals viz. maize, barley, sorghum and pearlmillet account for approximately 44% of total cereals fodder production. Pearlmillet (Pennisetum glaucum L.) is an annual crop belongs to the family Poaceae (Gramineae), which is grown for food as well as fodder. It is a fast growing and short duration crop which has high biomass production potential. Pearlmillet as a fodder crop has some additional advantages over sorghum and maize because of high crude protein content (9.9 to 14%) and absence of hydrocyanic acid which makes feeding of green fodder of any crop stage safer to the cattle. It is nutritious, palatable and can be fed as green, dry or conserved in the form of silage or hay.

Date of sowing is an important non-monetary input affecting the fodder yield of summer pearlmillet. The weather parameters like rainfall, temperature and day length greatly influence the crop yield through their effect on phenological development of the crop. Late sowing coincides with high temperature, high wind velocity and early monsoon showers at reproductive stage adversely affect crop growth and yield, whereas, early sowing faces problems of low temperature during the initial stage of growth in North Gujarat. Moreover, late sowing does not leave enough time to prepare the land for *kharif* crop which resulted in delayed sowing of *kharif* crops. The sowing date is reported to have a significant impact on crop growth and development (Abd EI-Lattief 2011), which is then reflected in yield and quality. Accurate decision-making with regard to the sowing date is not only important to achieve the higher crop yield and better quality fodder but also for minimizing the risk of crop failure. Santos et al (2017) reported that timely sowing decreases overall farming costs by eliminating labour and re-sowing costs. Therefore, it was of prime importance to find out the optimum date of sowing of fodder pearlmillet.

Feed shortage is a major challenge in livestock production enhancement. There are many options to cover the gap between forage demand and supply, one of them is adoption of high yielding crop varieties (Hassan et al 2014, Babiker et al 2015). Adoption of high yielding short duration varieties play an important role in the maximization of pearlmillet productivity. Screening of varieties which are appropriate to that particular climatic condition can help in boosting the production of pearlmillet. The identification of suitable variety to enhance productivity and quality is crucial to mitigate the present shortage of fodder requirement in summer season. Keeping in mind these facts, experiment was planned and conducted.

#### MATERIAL AND METHODS

Field experiment was conducted during summer season of 2021 at S.D. Agricultural University, Sardarkrushinagar. The standard week-wise meteorological data for the period of this investigation recorded at the Agricultural Meteorological Observatory (Fig. 1). The soil of the experimental field was loamy sand in texture with slightly alkaline in reaction. The details of the soil physical and chemical properties of the experimental plot are given in the Table 1. The experiment was laid out in split plot design with three replications. There were fifteen treatment combinations comprised of three dates of sowing in main plots *viz*.  $25^{th}$  February (D<sub>1</sub>),  $12^{th}$ March (D<sub>2</sub>) and  $27^{th}$  March (D<sub>3</sub>) and five varieties in sub plots *viz*. Gujarat Fodder *Bajra* 4 (V<sub>1</sub>), Gujarat Fodder *Bajra* 1 (V<sub>2</sub>), BAIF *Bajra* 1 (V<sub>3</sub>), TSFB 15-8 (V<sub>4</sub>) and Moti *Bajra* (V<sub>5</sub>). The crop was sown using recommended agronomic practices.

The biometric observations were recorded from randomly selected five plants in each net plot. Actual numbers of plants per metre row length were counted before harvesting of each cut without considering the tillers per plants. The plant height was measured from base to the tip of the plant at each cut. The number of leaves per plant was recorded at each cut. The average of five plants was calculated and presented as number of leaves/plant. The length of five leaves was measured from the base of the stem to the tip of the leaf to calculate average leaf length (cm). Similarly, the width of five leaves of the plant measured from the middle of the leaf to calculate the average leaf width (cm). Leaf: stem ratio was calculated during each cut at the time of harvest. The leaves were separated from the stem and fresh weight of both leaves and stem were recorded separately and leaf stem ratio is computed by dividing the leaves weight by stem weight and expressed in ratio. Average stem girth was calculated using the girth of five stem of the plant measured from the base, middle and tip of the stem. Two cuts were taken for green forage. The first cut was taken at 55 DAS,



Fig. 1. Mean weekly weather parameters recorded during crop growth period of summer-2021

whereas, the second cut was taken 35 days after first cut (90 DAS). The ring area of each plot was first harvested and removed. The plants from the net plot were harvested keeping 5 cm stubble height from ground level and the fresh weight of harvested produce was recorded for each treatment separately at each cut and converted into quintal per hectare. After calculating green forage yield, one kg of green plant sample was weighed randomly from each net plot and kept in a brown paper bag. Thereafter, samples were first sun-dried then oven dried at 65°C. After oven drying, the dry forage yield in quintal per hectare was calculated on the basis of dry weight of sample for each treatment at each cut.

Total nitrogen content was determined by modified Kjeldahl method. Crude protein content of fodder was estimated by multiplying nitrogen percentage with 6.25 (Tsen and Martin 1971). The previously grinded and powdered samples were used to estimate the crude fibre (Mahadevan (1965). The residue left after ether extract was first boiled in 1.25 N  $H_2SO_4$  followed by boiling in 1.25 N NaOH. The respective residue was washed with hot water, dried in oven and weighed. Thereafter, it was ashed in muffle furnace and per cent crude fiber was calculated using the formula given below.

	Weight of silica crucible with oven dry residues	
Crude fibre content $(0/) = -$	۔ Weight of silica crucible with ash	
	Weight of oven dry sample	~100

To evaluate the most effective and remunerative treatment, the relative economics of each treatment was worked out in terms of gross, net realization and benefit: cost ratio. The gross realization in term of ₹/ha was calculated from the income received from dry matter of each treatment with the prevailing market price. The cost of cultivation was worked out considering the cost of all the operations and inputs right from the preparation of land to harvesting of the crop. The net realization was worked out by deducting the total cost of cultivation from the gross realization per hectare for each treatment and recorded accordingly. The benefit: cost ratio (BCR) was calculated.

Benefit: Cost Ratio = Total cost of cultivation (₹/ha)

**Statistical analysis:** Analysis and interpretation of data was done using MS excel programme.

#### **RESULTS AND DISCUSSION**

**Growth and yield attributes:** The plant height, number of leaves per plant, leaf length, leaf: stem ratio and stem girth

were influenced significantly by different dates of sowing (Table 2). Crop sown on  $12^{th}$  March (D<sub>2</sub>) produced significantly higher plant height, number of leaves per plant, leaf length), leaf: stem ratio and stem girth at  $1^{st}$  and  $2^{nd}$  cut, and was at par with  $25^{th}$  February sowing. The leaf width of fodder pearlmillet recorded at  $1^{st}$  and  $2^{nd}$  cut was non-significant due to different dates of sowing. This might be due to the sowing of pearlmillet at appropriate time which provided favourable climatic conditions in terms of temperature, humidity and bright sunshine hour during crop growth period, that had resulted in more cell division and enlargement which ultimately leads to more growth of crop. The present findings are in accordance

with earlier observations (Abd El-Lattief 2011, Reiad et al 2014, Chaudhary et al 2020). Pearlmillet varieties significantly influenced plant height, number of leaves per plant, leaf length, leaf: stem ratio and stem girth except leaf width. Gujarat Fodder *Bajra* 4 recorded significantly higher plant height, number of leaves per plant, leaf length, leaf: stem ratio and stem girth and was at par with TSFB 15-8. This might be due to difference in genetic constitution among the varieties (Manjanagouda 2015, Bramhaiah 2016, Chaudhary 2021).

**Fodder yield:** Green fodder and dry fodder yields were influenced significantly by different dates of sowing (Table 3).

Table 1. Phy	sico-chemical	properties	of the	experimental soil

Properties	Soil depth (0-15 cm)	Methods of analysis
Mechanical composition		
Sand (%)	83.50	International pipette method (Piper 1966)
Silt (%)	7.55	
Clay (%)	7.10	
Textural classes	Loamy sand	
Chemical parameters		
Soil pH (1:2.5, Soil: Water ratio)	7.60	Potentiometric method (Jackson 1973)
Electrical conductivity (dS/m) 1:2.5, Soil: Water ratio at 25°C)	0.17	Schofield method (Jackson 1973)
Organic carbon (%)	0.16	Walkley and Black's rapid titration method (Walkley and Black 1934)
Available N (kg/ha)	160.20	Alkaline potassium permanganate method (Subbiah and Asijia 1956)
Available P <sub>2</sub> O <sub>5</sub> (kg/ha)	32.47	Olsen's method (Olsen et al 1954)
Available K₂O (kg/ha)	282.00	Flame photometric method (Jackson 1973)

## Table 2. Effect of date of sowing and varieties on growth of fodder pearlmillet

Treatment	Plant height (cm)		No. of leaves/plant		Leaf length (cm)		Leaf width (cm)		L:S ratio		Stem girth (mm)	
-	1 <sup>st</sup> cut	2 <sup>nd</sup> cut										
Main plot: Date of	sowing (D	)										
D <sub>1</sub> : 25 <sup>th</sup> February	166.4	145.2	9.29	7.90	65.01	60.92	1.86	1.76	0.49	0.45	30.49	28.51
D <sub>2</sub> : 12 <sup>th</sup> March	181.5	158.0	9.65	8.26	67.74	64.18	1.88	1.79	0.51	0.47	31.82	29.84
D₃: 27 <sup>th</sup> March	146.9	124.4	8.57	7.13	59.21	55.85	1.81	1.72	0.44	0.40	28.24	26.12
CD (p-0.05)	15.57	14.16	0.81	0.64	5.99	6.08	NS	NS	0.05	0.04	2.66	2.60
Sub plot: Varieties	(V)											
V₁: Gujarat Fodder <i>Bajra</i> 4	177.6	155.4	10.41	8.80	69.94	67.24	1.91	1.83	0.55	0.51	31.63	29.63
V <sub>2</sub> : Gujarat Fodder <i>Bajra</i> 1	157.5	135.1	8.40	7.15	60.76	56.07	1.82	1.71	0.43	0.40	29.24	27.19
V₃: BAIF <i>Bajra</i> 1	154.9	133.3	7.63	6.56	57.64	52.36	1.79	1.68	0.39	0.35	28.65	26.55
V₄: TSFB 15-8	172.4	149.7	10.08	8.49	67.30	64.84	1.88	1.81	0.53	0.49	31.17	29.17
V₅: Moti <i>Bajra</i>	162.1	139.4	9.32	7.82	64.29	61.06	1.85	1.76	0.48	0.45	30.22	28.24
CD (p-0.05)	12.77	10.35	0.68	0.52	4.87	4.74	NS	NS	0.04	0.03	2.08	2.08
Interaction (D × V)												
CD (p-0.05)	NS											

Crop sown on 12<sup>th</sup> March produced significantly higher total green fodder yield (589.8 q/ha) and dry fodder yield (139.2 q/ha). The 25<sup>th</sup> February sown crop was statistically at par with the crop sown on 12<sup>th</sup> March. The increase in yields with normal sowing might be attributed due to favourable effect on growth attributes *viz.* plant height, number of leaves per plant, leaf length, leaf: stem ratio and stem girth. This might have ultimately resulted in higher photosynthetic activities and also in production of more photosynthates( Chandrika et al 2012, Salama et al 2020, Kaur and Oberoi 2021).

Pearlmillet varieties significantly influenced the green fodder and dry fodder yield. Gujarat Fodder *Bajra* 4 recorded significantly higher total green fodder yield (625.6 q/ha) and dry fodder yield (148.5 q/ha). TSFB 15-8 was statistically at par with Gujarat Fodder *Bajra* 4 with respect to green and dry fodder yield. Fodder yield was significantly higher under Gujarat Fodder *Bajra* 4 which might be due to the superiority of the genotype to produce better growth characters like plant height, number of leaves per plant, leaf length, leaf: stem ratio and stem girth. In earlier studies also similar trend was

Table 3. Effect of date of a	sowing and	varieties on	yield of fodder	pearlmillet
------------------------------	------------	--------------	-----------------	-------------

Treatment	Gre	en fodder yield (q	/ha)	Dry fodder yield (q/ha)			
	1 <sup>st</sup> cut	2 <sup>nd</sup> cut	Total	1 <sup>st</sup> cut	2 <sup>nd</sup> cut	Total	
Main plot: Date of sowing (D)							
D₁: 25 <sup>th</sup> February	314.0	244.5	558.5	76.66	55.06	131.7	
D <sub>2</sub> : 12 <sup>th</sup> March	332.0	257.8	589.8	81.16	58.06	139.2	
D <sub>3</sub> : 27 <sup>th</sup> March	290.2	216.3	506.5	70.72	48.70	119.4	
CD (p-0.05)	30.04	24.48	47.92	7.30	5.46	11.19	
Sub plot: Varieties (V)							
V₁: Gujarat Fodder <i>Bajra</i> 4	347.1	278.5	625.6	85.80	62.66	148.5	
V <sub>2</sub> : Gujarat Fodder <i>Bajra</i> 1	296.1	217.0	513.1	71.87	49.48	121.4	
V₃: BAIF <i>Bajra</i> 1	273.6	198.1	471.7	66.21	45.15	111.4	
V₄: TSFB 15-8	329.0	263.5	592.5	80.58	58.77	139.4	
V₅: Moti <i>Bajra</i>	314.5	240.5	555.0	76.43	53.63	130.1	
CD (p-0.05)	28.28	18.76	38.88	7.02	4.18	9.34	
Interaction (D × V)							
CD (p-0.05)	NS	NS	NS	NS	NS	NS	

Table 4. Effect of date of sowing and varieties on quality of fodder pearlmillet

Treatment	Crude protei	n content (%)	Crude fibre content (%)		
	1 <sup>st</sup> cut	2 <sup>nd</sup> cut	1 <sup>st</sup> cut	2 <sup>nd</sup> cut	
Main plot: Date of sowing (D)					
D₁: 25 <sup>th</sup> February	7.85	7.62	27.33	23.30	
D <sub>2</sub> : 12 <sup>th</sup> March	8.00	7.78	27.61	23.47	
D <sub>3</sub> : 27 <sup>th</sup> March	7.73	7.51	26.98	23.08	
CD (p-0.05)	NS	NS	NS	NS	
Sub plot: Varieties (V)					
V₁: Gujarat Fodder <i>Bajra</i> 4	8.19	7.98	28.57	24.23	
V₂: Gujarat Fodder <i>Bajra</i> 1	7.69	7.47	26.39	22.32	
V₃: BAIF <i>Bajra</i> 1	7.59	7.37	26.24	22.16	
V₄: TSFB 15-8	7.99	7.77	28.02	24.01	
V₅: Moti <i>Bajra</i>	7.84	7.61	27.31	23.69	
C. D. at 5%	NS	NS	NS	NS	
Interaction (D × V)					
CD (p-0.05)	NS	NS	NS	NS	

Independent variable (x)	Dependent variable (y)	Correlation coefficient (r)	Regression equation	$R^2$
First cut				
Plant height (cm)	Green fodder yield (q/ha)	0.873**	y =45.20+1.62x	0.763
No. of leaves/plant		0.958**	y = 67.68+26.65x	0.917
Leaf length (cm)		0.976**	y = -32.77+5.39x	0.953
Leaf width (cm)		0.970**	y = -722.33+559.33x	0.942
Stem girth (mm)		0.924**	y = -157.74+15.56x	0.854
Green fodder yield (q/ha)	Dry fodder yield (q/ha)	0.999**	y = -4.85+0.26x	0.998
Second cut				
Plant height (cm)	Green fodder yield (q/ha)	0.859**	y =-13.62+1.78x	0.737
No. of leaves/plant		0.996**	y = -36.90+35.60x	0.992
Leaf length (cm)		0.995**	y = 76.59+5.24x	0.990
Leaf width (cm)		0.993**	y = -714.20+542.78x	0.987
Stem girth (mm)		0.922**	y = -219.80+16.31x	0.850
Green fodder yield (q/ha)	Dry fodder yield (q/ha)	0.998**	y = 2.16+0.22x	0.997
** = Significant at 1%				

 Table 5. Correlation and regression equations for various dependent and independent parameters of fodder pearlmillet as influenced by date of sowing and varieties

The variable x refers to the independent parameters listed in the column, variable y refers to the dependent parameters listed in the column

Table 6. Effect of date of sowing and varieties on economics of fodder pearlmillet

Treatment	Gross returns (₹/ha)	Cost of cultivation (₹/ha)	Net returns (₹/ha)	BCR
Main plot: Date of sowing (D)				
D₁: 25 <sup>th</sup> February	111700	52384	59316	2.13
D <sub>2</sub> : 12 <sup>th</sup> March	117960	52384	65576	2.25
D₃: 27 <sup>th</sup> March	101300	52384	48916	1.93
CD (p-0.05)	-	-	-	-
Sub plot: Varieties (V)				
V₁: Gujarat Fodder <i>Bajra</i> 4	125120	52384	72736	2.39
V <sub>2</sub> : Gujarat Fodder <i>Bajra</i> 1	102620	52384	50236	1.96
V₃: BAIF <i>Bajra</i> 1	94340	52384	41956	1.80
V₄: TSFB 15-8	118500	52384	66116	2.26
V₅: Moti <i>Bajra</i>	111000	52384	58616	2.12
CD (p-0.05)	-	-	-	-
Interaction (D × V)				
CD (p-0.05)	-	-	-	-

observed (Shroff and Patel 2017, Kaur and Goyal 2019, Chaudhary 2021).

Interaction effect between date of sowing and varieties: The interaction between date of sowing and varieties was non-significant for growth parameters, green and dry fodder yields as well as quality parameters.

**Quality parameters:** The crude protein content at  $1^{st}$  as well as  $2^{nd}$  cut of fodder pearlmillet crop did not differ significantly due to different date of sowing and varieties (Table 4). However,  $12^{th}$  March sown crop and Gujarat Fodder *Bajra* 4 variety had numerically high crude protein and crude fibre content values.

Correlation and regression analysis: There was

significant and positive relationship found between plant height, number of leaves/plant, leaf length, leaf width, stem girth with the green fodder yield of fodder pearlmillet during 1<sup>st</sup> cut and 2<sup>nd</sup> cut, as evident from correlation coefficient (r) analysis (Table 5). This indicates that the enhancement in all the independent variable will increase the green fodder yield (dependent variable). There was significant positive relationship between green fodder yield and dry fodder yield indicating direct positive relationship between both parameters. Regression equations also revealed that increase in plant height by 1 cm, 1 leaf/plant, 1 cm leaf length, 1 cm leaf width, 1 mm stem girth shall increase the green fodder yield of  $1^{st}$  cut by 1.62 q/ha, 26.65 q/ha, 5.39 q/ha, 559.33 q/ha and 15.56 q/ha, respectively. Similarly, the increase in plant height by 1 cm, 1 leaf/plant, 1 cm leaf length, 1 cm leaf width, 1 mm stems girth shall increase the green fodder yield of  $2^{nd}$  cut by 1.78 q/ha, 35.60 q/ha, 5.24 q/ha, 542.78 q/ha and 16.31 q/ha, respectively. Increase in green fodder yield by 1 q/ha in  $1^{st}$  and  $2^{nd}$  cut shall increase the dry fodder yield of 0.26 and 0.22 q/ha, respectively.

**Economics:** Crop sown on  $12^{\text{th}}$  March gave the highest gross returns (₹ 117960/ha), net returns (₹ 65576/ha) with maximum B: C ratio (2.25) as compared to other dates of sowing. In varieties, Gujarat Fodder *Bajra* 4 recorded the highest gross returns (₹ 125120/ha), net returns (₹ 72736/ha) with maximum B: C ratio (2.39) followed by TSFB 15-8. Among the pearlmillet varieties and date of sowing, Gujarat Fodder *Bajra* 4 and 12<sup>th</sup> March, respectively adjudged best on the basis of economics.

#### CONCLUSIONS

The fodder pearlmillet variety Gujarat Fodder *Bajra* 4 or TSFB 15-8 should be sown on 25<sup>th</sup> February or 12<sup>th</sup> March to obtain higher fodder yield under North Gujarat condition.

#### REFERENCES

- Abd El-Lattief EA 2011. Growth and fodder yield of forage pearl millet in newly cultivated land as affected by date of planting and integrated use of mineral and organic fertilizers. *Asian Journal of Crop Science* **3**(1): 35-42.
- Babiker SA, Khair MA, Tahir IS and Elhag FM 2015. Forage quality variations among some Sudan pearl millet [*Pennisetum* glaucum (L.) R. Br] collection. Annual Research and Review in Biology 5(4): 293-298.
- Bramhaiah U 2016. Evaluation of fodder pearl millet (Pennisetum glaucum L.) varieties under varied nitrogen levels in Southern agro climatic zone of Andhra Pradesh. M.Sc. (Agri.) Thesis. Acharya N.G. Ranga Agricultural University, Tirupati, Andhra Pradesh.
- Chandrika V, Shashikala T, Shanti M and Reddy KL 2012. Production potential of multi cut fodder bajra genotypes under varied dates of sowing. *Journal of Research ANGRAU* **40**(3): 54-57.
- Chaudhary HD 2021. Effect of nitrogen management and phosphorus levels on summer fodder pearl millet (Pennisetum glaucum L.) varieties under North Gujarat agro-climatic conditions. M.Sc. (Agri.) Thesis. Sardarkrushinagar Dantiwada Agricultural University, Sardarkrushinagar, Gujarat.
- Chaudhary JK, Patel AG, Gohil NB and Chaudhary DG 2020. Response of nutrient content and quality of summer forage pearl millet (*Pennisetum glaucum* L.) on sowing date and nitrogen

Received 12 June, 2024; Accepted 21 September, 2024

level. International Journal of Chemical Studies 8(5): 841-844.

- Hassan M, Ahmad A, Zamir SI, Haq I, Khalid F, Rasool T and Hussain A 2014. Growth, yield and quality performance of pearl millet (*Pennisetum glaucum* L.) varieties under Faisalabad Pakistan conditions. *American Journal of Plant Sciences* 5: 2215-2223.
- Jackson ML 1973. Soil chemical analysis. Prentice Hall of India Pvt. Ltd., New Delhi.
- Kaur M and Goyal M 2019. Influence of different nitrogen levels on growth, yield and quality of forage pearl millet (*Pennisetum glaucum*L.) genotypes. *Forage Research* **45**(1): 43-46.
- Kaur M and Oberoi HK 2021. Heat unit requirement, fodder yield and quality of pearl millet varieties under different sowing windows in Central Punjab. In virtual National conference on Strategic Reorientation for Climate Smart Agriculture V-AGMET, 100-103.
- Mahadevan SA 1965. Laboratory Manual for Nutrition Research, pp 56-58.
- Manjanagouda SS 2015. Performance of dual purpose pearlmillet (Pennisetum glaucum L.) varieties as influenced by cutting and nitrogen management. M.Sc. (Agri.) Thesis. University of Agricultural Sciences, Bangalore.
- Olsen SR, Cole VC, Wetanale FS and Dean LA 1954. *Estimation of available phosphorus in soil by extraction with sodium bicarbonate.* United States Department of Agriculture Circular, No. 939.
- Pareek P, Patel MR, Patel HK and Patel PM 2015. Effect of irrigation and nitrogen levels on forage yield and quality of pearlmillet (*Pennisetum glaucum* L.). *International Journal of Agricultural Sciences* **11**(2): 264-267.

Piper CS 1966. Soil and Plant Analysis. Hans Publisher, Mumbai.

- Reiad MS, Maha MA, Hamada, Abd EL-Maaboud MS and Khalil MH 2014. Forage growth and productivity of pearl millet as affected by soil mulching and planting date under salinity conditions. *Egyptian Journal of Agronomy* **36**(1): 75-94.
- Salama HSA, Shaalan AM and Nasser MEA 2020. Forage performance of pearl millet (*Pennisetum glaucum* [L.] R. Br.) in arid regions: yield and quality assessment of new genotypes on different sowing dates. *Chilean Journal of Agricultural Research* **80**(4): 572-584.
- Santos RD, Boote KJ, Sollenberger LE, Neves AL, Pereira LG, Scherer CB and Goncalves LC 2017. Simulated optimum sowing date for forage pearl millet cultivars in multilocation trials in Brazilian semi-arid region. *Frontiers in Plant Science* **8**(2074): 1-11.
- Shroff JC and Patel PM 2017.Performance of dual purpose pearl millet as influenced by different cutting management practices and nitrogen levels. *International Journal of Chemical Studies* **5**(5): 601-603.
- Subbiah BV and Asijia GC 1956. A rapid procedure for the estimation of available nitrogen in soils. *Current Science* **25**: 259-260.
- Tsen CC and Martin EE 1971. A note on determining protein contents in various wheat flours and flour streams by the Kjeldahl and by neutron-activation methods. *Cereal Chemistry* **48**(6): 721-726.
- Walkley A and Black IA 1934. An examination of the digestion method for determining soil organic matter and a proposed modification of the chromic acid titration method. *Soil Science* **37**(1): 29-38.



Indian Journal of Ecology (2024) 51(5): 1037-1041 DOI: https://doi.org/10.55362/IJE/2024/4348 Manuscript Number: 4348 NAAS Rating: 5.38

# Optimizing Planting Geometry and Nutrient Management for Enhanced Growth and Yield of Teff (*Eragrostis tef* (Zucc.) Trotter) in Eastern Dry Zone of Karnataka

# Manjunath S. Melavanki, B. Boraiah<sup>1</sup>, Mahantesh B. Nagangoudar<sup>1</sup> and R.T. Chethan Babu<sup>2</sup>

Agronomy Section, ICAR-National Dairy Research Institute, Karnal-132 001, India <sup>1</sup>Department of Agronomy, College of Agriculture, University of Agricultural Sciences, GKVK, Bengaluru- 560 065, India <sup>2</sup>Department of Agronomy, Sri Venkateswara Agricultural College, Tirupati-517 502, India. *E-mail: manjumelavanki*366@gmail.com

**Abstract:** Teff (*Eragrostis tef (Zucc.*) Trotter is an introduced minor millet therefore, among prime areas of research is developing standard agro-techniques for higher productivity. Field experiment was conducted on optimizing planting geometry and nutrient management for enhanced growth and yield of teff during *kharif*-2021 at UAS, GKVK, Bangaluru in factorial randomized block design with ten treatment combinations consisting two planting geometry (30 cm × 10 cm and 45 cm × 10 cm) and five nutrient management (50, 75, 100 and 125 % recommended dose of fertilizers, (RDF) and absolute control). As per UAS, Bengaluru package of practices, RDF is 20 N: 20 P<sub>2</sub>O<sub>5</sub>: 20 K<sub>2</sub>O and FYM @ 6 t ha<sup>-1</sup>. Growth and yield parameters significantly varied with the treatments. Maintaining 30 cm × 10 cm spacing recorded significantly higher plant height (74.79 cm, 93.57 cm at 60 DAP and at harvest respectively), lodging (56.39%), grain yield (2.37 q ha<sup>-1</sup>) and straw yield (3.65 q ha<sup>-1</sup>). Application of 125 % RDF resulted in significantly higher plant height (80.05 and 99.33 cm) and number of tillers (16.45 and 23.07 per plant<sup>-1</sup>) at 60 DAP and harvest respectively. However, significantly higher grain yield (2.73 q ha<sup>-1</sup>) was at 100 % recommended dose of fertilizers

Keywords: Eragrostis tef (Zucc.) Trotter, Teff, Nutrient management, Planting geometry

Minor/small millets are the best examples of underutilized crops gaining more importance in recent days due to climatic smartness, nutritional richness, health benefits, low risk in production and lower production cost. Karnataka is majorly known for millet cultivation in India, primarily with respect to finger millet and foxtail millet among the other states. Nowadays farmers are much concerned in commercial agriculture and are replacing sustainable crops with cash crops, hence, the area under millets is declining day by day (Durgad et al 2019). Teff (Eragrostis tef (Zucc.) Trotter is an introduced minor millet originated and diversified in Ethiopia and cultivated on an area of 3.01 m ha, with an annual production of 5.01 m t and productivity of 1.664 t ha<sup>-1</sup> in Ethiopia (Lee, 2018). Simultaneously, interests in teff cultivation are spreading to other parts of the world viz., Australia, Canada, Cameroon, China, India, South Africa, The Netherlands, UK, Uganda, and the USA. Though teff grass is a boon to Indian farmers, little is known about its cultivation, production, and value addition. The limitations comprise lack of improved varieties, lodging of the crop at later stages of crop growth, lack of mechanization, lack of awareness, and the standard package of practice (agrotechniques). Premier areas of research are need of the hour to promote this crop at the global level. The standard agro techniques viz., planting method, plant geometry, nutrient management, weed management, pest management, and reduction in post-harvest losses must be intended at reducing the loss in the output. Among these agro techniques, planting geometry and nutrient management play a vital role in increasing yield levels of teff crop as they ensure optimum plant population and ensures an adequate supply of nutrition to the plants. Row spacing is the key component in maximizing crop yield through optimizing plant population, improving light availability, and reducing weed competition. Therefore, it is of principal importance to develop and recommend suitable row spacing for maximizing teff production (Mihretie et al 2020). Dryland soils are 'not only thirsty but hungry too' and will make it mandatory to supply nutrients through external sources viz., organic and inorganic fertilizers, these nutrients are to be applied wisely to achieve maximum yield potentiality of the crop with the least losses. The modern agronomic practices and technologies have the potential to substantially enhance teff productivity to ensure food security. The experiment was conducted with the objectives to study the effect of two levels of rows spacing, five levels of fertilizer rates and their interaction on growth parameters, yield components and yield.

#### MATERIAL AND METHODS

Field experiment was conducted during Kharif season of

2021 at Zonal Agricultural Research Station (ZARS), Gandhi Krishi Vignana Kendra, University of Agricultural Sciences, Bengaluru which comes under Eastern Dry Zone (Zone-5) of Karnataka. The geographical reference point of the experimental site was 13° 05' North latitude and 77° 34' East longitude and at an altitude of 924 m above mean sea level. The soil was red sandy loam in texture consisting of 53.6 per cent coarse sand, 14.7 per cent fine sand, 14.9 per cent silt and 14.8 per cent clay. The soil was acidic (6.85 pH) in reaction with an electrical conductivity of 0.40 dS m<sup>-1</sup>, low organic carbon (0.42 %) and medium in available nitrogen (315.2 kg ha<sup>-1</sup>), phosphorous (28.1 kg ha<sup>-1</sup>) and potassium (281.3 kg ha<sup>-1</sup>). During the crop season from August to November, a total of 881.2 mm of rainfall was recorded. The experiment was laid out in factorial randomized block design with ten treatment combinations and replicated three times. Treatments consisting of two factors planting geometry (S.: 30 cm × 10 cm and S<sub>2</sub>: 45 cm × 10 cm) and five nutrient management N<sub>4</sub>: 50 % recommended dose of fertilizers (RDF), N<sub>2</sub>: 75 %, N<sub>3</sub>: 100 %, N<sub>4</sub>: 125 % RDF and N<sub>5</sub> (absolute control). Farm yard manure was applied @ 6 t ha<sup>-1</sup> commonly to all the treatments except absolute control. In this experiment, as per the UAS, Bengaluru package of practices, RDF of little millet (20 N: 20 P<sub>2</sub>O<sub>5</sub>: 20 K<sub>2</sub>O and FYM @ 6 t ha<sup>-1</sup>) was taken as base for determining fertilizer application rates in teff as teff morphology is much more similar to that of little millet compared to other millets. Brown seeded type teff was used for sowing. The gross plot size was 4.5 m × 3.0 m and net plot size was 3.9 m × 2.8 m (30 cm × 10 cm), 2.6 m × 2.8 m (45 cm × 10 cm) and treatments were allocated randomly in the plots within the blocks. Recommended dose of fertilizers and farm yard manure were applied at the time of sowing. Periodical observations were taken on growth parameters at 30, 60 DAP and at harvest where, five plants were selected randomly from each net plot and tagged with a label for recording various biometric observations on growth and yield parameters. Crop was harvested at 110 DAS, threshed and yield of the individual plots recorded separately and expressed in terms of per hectare. Lodging percentage was calculated by using formula:

Lodging (%) =  $\frac{\text{Number of lodged effective tillers}}{\text{Total number of effective tillers}} \times 100$ 

**Statistical analysis:** The data recorded on growth and yield parameters were subjected to Fisher's method analysis of variance using FRBD in MS excel.

## **RESULTS AND DISCUSSION**

#### **Growth Attributes**

Plant height and number of tillers: Plant height was

superior under 30 cm × 10 cm (74.79 and 93.57 cm, respectively) at 60 days after planting and at harvest. It could be attributed to the fact that higher plant population would certainly reduce the amount of light availability to the individual plant, particularly to lower leaves due to greater shading. As the mutual shading increases at higher plant densities, the plant tends to grow taller. Due to the elongation of internodes in narrow spacing leads to increase in plant height. Similar findings were reported by Hulihalli and Shantveerayya (2018) in buckwheat genotypes. Significantly higher plant height at 60 DAP (80.05 cm) at harvest (99.33 cm) was under application of 125 per cent RDF due to increased availability of soil nitrogen which leads to improved meristematic activity in terms of increased cell enlargement and elongation. Greater cell elongation resulted in increased plant height. Comparable findings were also described by Prakasha et al (2018). The number of tillers increased with age of the plant, at slower rate during early stages and at rapid rate up to 60 DAP and very few tillers were produced at later stage of plant growth. Wider spacing of teff produced significantly greater number of tillers per plant (14.76 and 20.34 plant<sup>-1</sup>) at 60 DAP and at harvest respectively, as the individual plants could have effectively utilized the available resources such as space, foraging area for root system, light utilization etc. and thus enhanced the tiller production at wider spaced treatment. Similarly increased number of tillers plant under wider spacing was documented by Kumari et al (2015). Application of 125 per cent RDF resulted in significantly more tillers per plant at 60 and 90 DAP (16.45 and 23.07, respectively) among all the nutrient levels. This indicates that fertilization encouraged tiller formation in the crop due to increased uptake of available form of major nutrients present in the soil under increased fertilizer application. Parallel findings were reported by Kumari et al (2015).

Leaf area and Dry matter production: The wider spacing of 45 cm × 10 cm resulted in larger leaf area at 60 days after planting and at harvest (5.93 and 6.10 cm<sup>2</sup> plant <sup>1</sup>respectively) encouraging better nutrient availability and reducing competition for growth promoting factors (Table 1). Wider spacing also favoured higher dry matter production (4.17 and 29.46 g plant<sup>-1</sup>) at 60 DAP and harvest, attributed to increased tillers and better assimilating area (LAI). Improvement of LAI might have increased the radiation use efficiency and photosynthetic efficiency of crop and have induced to produce more DMP per plant under wider spacing. Application of 125 RDF significantly increased leaf area (6.85 and 7.12 cm<sup>2</sup> plant<sup>-1</sup>) and dry matter (4.95 and 35.46 g plant<sup>-1</sup>) at 60 DAP and harvest, respectively. This higher RDF provided more available nitrogen, phosphorus and
potassium leading to rapid cell division and leaf expansion, reflected in terms of increased leaf area is similar to the findings in finger millet by Veeresh and Ramachandrappa (2016) and Hebbal et al (2018). Also, the adequate nutrient supply enhanced root development, photosynthetic rate, and carbohydrate utilization for more dry matter accumulation as reported by Charate et al (2018).

**Lodging:** Significantly higher lodging (56.39 %) was observed under narrow spacing of 30 cm × 10 cm (Table 2) which was due to competition for light under narrow spacing to utilize more incoming solar radiation which made the stem lean and when the plant grows taller, the nodes near the ground are unable to keep higher above ground biomass which resulted in greater lodging of teff. Similarly, with increased nitrogen application, the plant height was increased linearly and lean stems became more succulent

and were unable to keep the plant in upright position. Due to this, lodging per cent was significantly higher under application of 125 per cent RDF than other lower nitrogen treatments. In contrary, under absolute control plants were short and stems were strong due to lack of sufficient nitrogen availability. Hence lodging per cent was least (39.89) in absolute control at harvest. Similar trend was observed by Mahantesh (2020).

#### **Yield Attributes and Teff Yield**

**Number of productive tillers:** Narrow spacing ( $30 \text{ cm} \times 10 \text{ cm}$ ) resulted in significantly a greater number of effective tillers ( $428.16 \text{ m}^2$ ) compared to wider spacing of  $45 \text{ cm} \times 10 \text{ cm}$  ( $383.07 \text{ m}^2$ ), Under wider spacing in spite of higher number of the tillers per plant, the number of productive tillers per unit area were less due to lower plant density. Significantly greater number of effective tillers were under

Table 1.	Effect of	planting	geometry	/ and nutrien	t management	on growth	parameters	of teff

Treatments	Plant he	ight (cm)	Number of ti	llers plant <sup>1</sup>	Leaf area (	cm² plant¹)	Dry matter	· (g plant⁻¹)
	60 DAP	At harvest	60 DAP	At harvest	60 DAP	At harvest	60 DAP	At harvest
Factor 1: Planting geometry (	S)							
S₁: 30 cm × 10 cm	74.79	93.57	12.95	18.13	5.45	5.62	3.92	27.61
S <sub>2</sub> : 45 cm × 10 cm	63.92	86.20	14.76	20.34	5.93	6.10	4.17	29.46
CD (p = 0.05)	10.86	7.24	0.51	0.47	0.18	0.11	0.15	0.70
Factor 2: Nutrient levels (N)								
N <sub>1</sub> : 50 % RDF (10:10:10)	66.98	86.75	12.74	17.91	5.10	5.16	3.53	26.10
N <sub>2</sub> : 75 % RDF (15:15:15)	67.65	88.00	14.02	19.33	5.70	5.89	4.06	28.28
N <sub>3</sub> : 100 % RDF (20:20:20)	78.23	95.33	15.65	21.03	6.23	6.61	4.38	32.02
N₄: 125 % RDF (25:25:25)	80.05	99.33	16.45	23.07	6.85	7.12	4.95	35.46
N <sub>5</sub> : Absolute control	53.87	80.00	10.40	14.83	4.57	4.54	3.33	20.83
CD (p = 0.05)	17.17	11.44	0.81	0.74	0.28	0.17	0.23	1.11

Table 2. Effect of planting geometry and nutrient management on Lodging, yield parameters and yield of teff

Treatments	Lodging (%)	Productive tillers (m <sup>-2</sup> )	Grain weight (g ear <sup>-1</sup> )	Grain yield (q ha⁻¹)	Straw yield (q ha⁻¹)
Factor 1: Planting geometry (S)					
S₁: 30 cm × 10 cm	56.39	428.16	0.287	2.37	3.65
S <sub>2</sub> : 45 cm × 10 cm	52.97	383.07	0.296	2.04	3.34
CD (p = 0.05)	2.55	44.74	0.007	0.30	0.11
Factor 2: Nutrient levels (N)					
N <sub>1</sub> : 50 % RDF (10:10:10)	44.84	385.50	0.277	1.90	2.88
N <sub>2</sub> : 75 % RDF (15:15:15)	55.37	423.17	0.290	2.23	3.47
N <sub>3</sub> : 100 % RDF (20:20:20)	62.60	485.41	0.314	2.73	4.08
N₄: 125 % RDF (25:25:25)	70.69	425.00	0.305	2.45	4.36
N₅: Absolute control	39.89	309.00	0.270	1.74	2.66
CD (p = 0.05)	4.03	70.74	0.012	0.48	0.18

application of 100 per cent RDF (485.41 m<sup>2</sup>), however, it was on par with application of 125 and 75 per cent RDF due to higher number of tillers per plant that were erect enough to produce harvestable panicles. On the contrary, with increased nitrogen application the plants were much prone to lodging prior to panicle initiation itself, resulting in higher number of non-panicle bearing tillers.

**Grain weight per ear:** Wider spacing resulted in significantly higher grain weight per ear (0.296 g) compared to narrow spacing (0.287 g) which is mainly attributed to greater efficiency of the plants through nutrient absorption, photosynthetic activity and photosynthates translocation within the plants under leisure availability of growth resources in a wide spaced environment. Significantly higher grain weight per ear was observed under application of 100 per cent RDF (0.314 g) and was on par with application 125 per cent RDF (0.305 g). This is mainly attributed to the poor panicle initiation at higher nutrient application due to lodging of crop. There was poor photosynthate formation and supply to economic parts under lower nutrient supply levels and under absolute control. Balappa (2021) also observed similar trend.

Grain yield and straw yield: Among different planting geometry, significantly higher (2.37 q ha<sup>-1</sup>) grain yield was attained under the narrow spacing of 30 cm × 10 cm compared to wider spacing (2.04 q ha<sup>-1</sup>) of 45 cm  $\times$  10 cm. The higher grain yield in narrow spacing might be due to enhanced yield attributes, thus forming larger sink size coupled with efficient translocation of photosynthates to the sink. Simultaneously, due to higher plant population under closer spacing at 30 cm × 10 cm might have contributed to maximum dry matter production per unit area which ultimately enhanced the straw yield. Similar trend was observed by Thakur et al (2019). Grain yield was significantly higher under application of 100 per cent RDF (2.73 q ha<sup>-1</sup>) as lodging was observed before grain filling stage under 125 per cent RDF causing severe yield reduction in teff. However, under application of 100 per cent RDF it was observed immediately after grain filling stage which has reduced impact on development of panicles. Unlike grain yield, the treatment supplied with highest fertilizers i.e, 125 per cent RDF resulted in statistically higher straw yield (4.36 q ha<sup>-1</sup>) due to a more effective photosynthetic structure that allowed for more photosynthates to be synthesized, accumulated, partitioned and translocated to different regions of the plant. The crop was able to grow and develop more effectively reflecting in higher straw yield. Shankar (2017) and Ambresha (2017) also found similar results in little and foxtail millet respectively. However, the influence was nonsignificant with respect to interaction of different planting geometry and nutrient management.

#### CONCLUSION

Maintaining narrow spacing of 30 cm × 10 cm allows efficient utilization of growth factors ultimately leading to improved growth and development of teff crop than wider spacing. Among varied nutrient levels, supply of 100 % RDF ( $20 \text{ N}: 20 \text{ P}_2\text{O}_5: 20 \text{ K}_2\text{O}$ ) along with farm yard manure @ 6 t ha<sup>-1</sup> results in statistically higher yield of teff crop. Further, increase in fertilizer levels is not recommended as it leads to greater lodging of the crop because of lean and long nature of the stem which cannot support the higher shoot biomass.

#### REFERENCES

- Ambresha 2017. Effect of nitrogen and potassium levels on growth and yield of foxtail millet (Setaria italica L.). M.Sc. (Agri.) Thesis, University of Agricultural Sciences, GKVK, Bangalore, Karnataka, India.
- Balappa PG 2021. *Response of transplanted teff [Eragrostis tef (Zucc.) Trotter] to plant population and nutrient levels.* M.Sc. (Agri.) Thesis, University of Agricultural Sciences, Raichur.
- Charate S, Thimmegowda MN, Rao GE, Ramachandrappa BK and Sathish A 2018. Effect of nitrogen and potassium levels on growth and yield of little millet (*Panicum sumatrense*) under dryland *Alfisols* of Southern Karnataka. *International Journal of Pure and Applied Bioscience* **6**(6): 918-923.
- Durgad AG, Amrutha TJ, Suresh SP, Hiremath GM, Goudappa SB and Ananda N 2019. Economics of foxtail and little millets production in Ballari and Koppal districts of Karnataka, India. *International Journal of Current Microbiology and Applied Science* **9**: 214-222.
- Hebbal N, Ramachandrappa BK, Mudalairiyappa and Thimmegouda MN 2018. Yield and economics of finger millet with establishment methods under different planting geometry and nutrient source. *Indian Journal of Dryland Agricultural Research and Development* **33**(1): 54-58.
- Hulihalli UK and Shantveerayya 2018. Evaluation of buckwheat genotypes to different planting geometries and fertility levels in Northern Transition Zone of Karnataka. *International Journal of Agricultural and Biological Engineering* **12**(2): 64-68.
- Kumari CR, Shanthi P, Niveditha M, Sudheer KVS and Reddy B S 2015. Growth and yield of bajra hybrid as influenced by spacing and nitrogen levels in rainfed *Alfisols*. *Andhra Pradesh Journal of Agricultural Sciences* 2(2): 96-103.
- Lee H 2018. Teff a rising global crop: Current status of teff production and value chain. *The Open Agriculture Journal* **12**(5): 16-24.
- Mahantesh BN 2020. Performance of teff (Eragrostis tef [Zucc.] Trotter) under different planting methods and nutrient management. M.Sc. (Agri.) Thesis, University of Agricultural Sciences, Dharwad, Karnataka.
- Mihretie F, Tsunekawa A, Bitew Y, Chakelie G, Derebe B, Getahun W, Beshir O, Tadesse Z and Asfaw M 2020. Teff [*Eragrostis tef* (Zucc.)] rainfed yield response to planting method, seeding density and row spacing. *Journal of Agronomy* **113**(1): 111-12.
- Prakasha G, Murthy KNK, Prathima AS and Rohani NM 2018. Effect of spacing and nutrient levels on growth attributes and yield of finger millet (*Eleusine coracana* L. Gaertn) cultivated under guni planting method in red sandy loamy soil of Karnataka, *International Journal of Current Microbiology and Applied Science* **7**(5): 1337-134.
- Shankar C 2017. Response of little millet (Panicum sumatrense) to levels of nitrogen and potassium. M.Sc. (Agri.) Thesis, Univ. Agric. Sci., Bengaluru, Karnataka, India.
- Thakur AK, Kumar P and Netam PS 2019. The effect of different nitrogen levels and plant geometry, in relation to growth

characters and yield of browntop millet (*Brachiaria ramosa* L.) at Bastar Plateau Zone of Chhattisgarh. *International Journal of Current Microbiology and Applied Science* **8**(2): 2789-2794.

Veeresh H and Ramachandrappa BK 2016. Influence of

Received 23 July, 2024; Accepted 06 October, 2024

conservation tillage and nutrient management practices on the performance of finger millet (*Eleusine coracana*) and weed growth under dryland *Alfisols* of Southern Karnataka. *Research Journal of Agricultural Sciences* **7**(3): 495-498.



# Assessment of Heavy Metal Pollution in Soil of Different Land Uses of in Semi-Arid Region of Jaipur, Rajasthan

Sonali Tiwari, Naveen Kumar, Priyanka Jatav and Archana Meena

Department of Botany, University of Rajasthan, Jaipur-302 004, India E-mail: sona435768@gmail.com

**Abstract:** Due to extensive farming, urbanization and rapid growth, heavy metal contamination in soil has now a major issue in India. The semi-arid Jaipur region of Rajasthan underwent an environmental geochemistry assessment to ascertain the impact of pollution in the study area seasonally in the 2022. Soil samples collected from agriculture, industrial and forest area (as a control) in Jaipur were examined for soil physicochemical characteristics and heavy metal content, i.e., Cr, Pb, Ni, Cd, Zn, Cu, Mn and Fe by using the APHA method through an atomic absorption spectrophotometer. The determined mean metal concentration is in the order of Zn > Fe > Mn > Ni > Pb > Cr > Cu > Cd. Heavy metal contamination factor , contamination degree, geoaccumulation index, enrichment factor and potential ecological risk index. Conclusively, the concentration of heavy metals in the soil samples is within permissible limits according to prescribed standards.

Keywords: Heavy metals, Pollution indices, Seasonal variation, Semiarid

Soil on the earth's surface is formed naturally, carries specific chemical, physical, mineralogical and biological characteristics, and varies from the earth's surface to its depths (Thakre et al 2012). In urban ecosystems, changes in land use have significantly impact soil properties. The origin of metals in soils could be geological (weathering and erosion) or anthropogenic (Sahu et al 2024), which could be differentiated as point sources (domestic, mining, refining, industrial and manufacturing) and non-point sources (fertilization, surface runoff, irrigation and agriculture). In semi-arid areas, where the supply of fresh water for irrigation continues to decline, the use of sewage water for cultivation is now a common practice, resulting in increased levels of heavy metals (trace elements) such as nickel (Ni), mercury (Hg), iron (Fe), manganese (Mn), zinc (Zn), copper (Cu), lead (Pb), chromium (Cr), arsenic (As), and cadmium (Cd), which have toxicity and extreme persistence, causing serious harm to humans and the environment (Kurniawan et al 2006, Singh et al 2020). In recent studies, various indices have been used to measure the level of metal contamination and understand its sources, distribution and allocation in soil including the geo-accumulation index (Igeo), enrichment factor (EF) and others like contamination factor, potential ecological risk index.

Jaipur, the state's capital city, has seen significant economic growth in the past two decades due to various industries such as textile, printing, dyeing, gems, jewellery, ready-made garments and marbles. However, the city poses a serious environmental risk because of the large amounts of water, chemicals and dyes required for production and commercial use (Chavan 2001). Dyes, bleaching chemicals, salts, acids and heavy metals like Cr, Cu, Pb, Zn and Fe are dumped constantly into Amanishah Nalla without treatment, mainly originating from the textile and dyes industries located in the Sanganer area of Jaipur (Marwari et al 2012). The present study aims to estimate the spatial and seasonal soil concentrations of eight heavy metals, viz. chromium, lead, nickel, cadmium, zinc, iron, manganese and copper and to assess the contamination level in soil using geochemical indices.

#### MATERIAL AND METHODS

Jaipur extends from 26°47'0.21" to 27°1'36.84" N and 75°41'15.19" to 75°54'36.27" and has hot summers with temperatures ranging from 20–32° C, low rainfall and low relative humidity. The minimum recorded temperature is 5°C in January and the maximum is 46°C in May and the average rainfall was 74.81 mm during the study period (IMD 2022). The Nahargarh forest range of Jaipur, which covers 52.40 km<sup>2</sup>, is mostly dominated by Dhok (*Anogeissus pendula,* which covers 80% of the total vegetation). The other associated tree species are *Jatropha curcas, Maytenus emarginata, Adina cordifolia, Prosopis cineraria, Balanites aegyptiaca, Acacia nilotica, Acacia senegal, and Tecomella unduleta.* For the present study, seven sampling sites were selected based on land use activities, level of contamination and water use for irrigation (Table 1).

Soil sampling and analysis: Soil samples were collected

for three seasons, i.e., pre-monsoon (March to May), monsoon (June to September) and post-monsoon (October to February) during 2022. Permanent 10×10 quadrats were used to gather soil samples from five different sites at a depth of 0 to 10 cm in each land use to make composites. To sample stainless-steel trowels were used to prevent metal contamination. Soil samples were manually cleaned, combined, and stored in zipper polythene bags. Therefore, three locations were selected for each land use at some distance, for a total of 20 samples, and each sample was tested in a triplicate manner. Soil samples were air dry, grounded and sieved to evaluate the physicochemical parameter and trace elements as per the recommended procedures (Table 2).

**Environmental pollution indices:** In study predicted heavy metal contamination risks using soil trace element concentrations and Shale's trace element values as baseline

data: Cr=90, Pb=20, Ni=68, Cd=0.30, Zn=95, Fe=47,200, Mn=850 and Cu=45 (Turekian and Wedepohl 1961).

**Geoaccumulation index:** Muller (1969) devised the geoaccumulation index (Igeo) to analyse the heavy metal pollution in soil. Compare trace element amounts with background values to see contamination.

Igeo = 
$$\log 2 \frac{Cn}{1.5Bn}$$

Where Bn is the average upper crust's (Turekian and Wedepohl 1961) trace element concentration, and Cn is the concentration in soil samples. The constant 1.5 accounts for environmental changes and human impact.

**Enrichment factor (EF):** The enrichment factor (EF) is used to measure the impact of human activity on soil composition. Sinex and Helz's (1981) equation was used to calculate EF by comparing the concentration of the element being studied (Cn) to the concentration of a reference element (CRef) in the

Table 1. Sampling sites description

Site name	Latitude	Longitude	Elevation (mm)	Vegetation	Site description
Sitapura	N26º47'32.04"	E75º49'39.36"	314	Azadirachta indica, Acacia nilotica, Nerium oleander	Industrial area
Bagru	N26º49'14.17"	E75º32'39.25"	308	Rabi: <i>Sinapis alba</i> L. (Sarso) Kharif: <i>Cenchrus americanus</i> L. Morrone (Bajra)	Industrial area, rainfed
Muhana	N26º47'36.52"	E75º45'4.59"	321	Rabi: <i>Coriandrum sativum</i> L. (Dhaniya), <i>Medicago sativa</i> L. (Ranjaka) Kharif: <i>Spinacia oleracea</i> L. (Palak), <i>Abelmoschus</i> <i>esculentus</i> (L.) Moench (Bhindi)	Agriculture field irrigated with nala's water
Vidhani	N26º46'7.40"	E75º52'4.73"	292	Rabi:Solanum melongena L. (Bengan), Sinapis alba L. (Sarso), Triticum aestivum L. (Wheat) Kharif: Cenchrus americanus L. Morrone (Bajra), Sorghum bicolor L. (Jwar)	Agriculture field irrigated with nala's water
Begas	N26º52'26.69"	E75º33'8.04"	344	Rabi: Triticum aestivum L. (Wheat), Allium cepa L. (Pyaj), Medicago sativa L. (Ranjaka), Lathyrus oleraceus Lam. (1779) (Pea) Kharif:Arachis hypogaea L. (Peanut), Cenchrus americanus L. Morrone (Bajra)	Agriculture field irrigated with tank's water
Chomu	N27º10'24.31"	E75º40'55.02"	405	Rabi: Triticum aestivum L. (Wheat), Allium cepa L. (Pyaj), Medicago sativa L. (Ranjaka) Kharif: Praecitrullus fistulosus (Tinda), Cenchrus americanus L. Morrone (Bajra)	Agriculture field irrigated with tank's water
Nahargarh	N26º58'56.0"	E75º48'01.0"	490	Anogeissus dominated	Forestarea

Table 2. Procedure for soil para	ameter analvs	IS
----------------------------------	---------------	----

Parameter	Method used	References
Trace elements	AAS (Analytikjena ZEEnit 700 P)	(APHA 2005)
SM	Gravimetric (Oven dry)	
pH (1:2)	Potentiometry (Eutech model 510)	(Jackson 1973)
EC (1:2)	Conductometry (El model 602)	(Jackson 1973)
SOC	Rapid titration method	(Walkley and Black 1934)
SN	Micro-kjeldhal method (Kelplus classic dxvatse)	(Subbiah and Asija 1956)
SP	Spectrophotometry (Systronics 1203)	(Olsen et al 1954)
Soil texture	Sieve analysis	(Gee and Bauder 1979)

SM soil moisture, EC electrical conductivity, SOC soil organic carbon, SN soil nitrogen, SP soil phosphorus, AAS atomic absorption spectrophotometer

soil. Common reference elements used are Al (Islam et al 2015) and Fe (Parvez et al 2023), and Mn (Fabretti et al 2009, Sakata et al 2011). In this study, Mn was used to calculate enrichment.

$$EF = \frac{\left(\frac{Cn}{CRef}\right) \text{sample}}{\left(\frac{Cn}{CRef}\right) \text{Background}}$$

**Contamination factor and degree of contamination:** Soil pollution is evaluated using two measures (Hakanson 1980): the contamination factor (CF) and the contamination degree (CD). The CF measures the concentration of a heavy metal in a soil sample compared to the normal level of that same metal in the environment. The CD determines the overall contamination level by adding up the values of all the contaminants present in the sample.

$$F = \frac{(\text{Sample concentration}) \text{ sample}}{(\text{Sample concentration}) \text{ background}}$$

**Potential ecological risk index:** This index reflects the toxicity of soil heavy metals, and the response of the environment is computed (Hakanson 1980):

$$RI = \sum_{i}^{n} E_{r}^{i}$$
$$E_{r}^{i} = T_{r}^{i} * C_{r}^{i}$$

Where Er is the potential ecological risk factor for heavy metals, Tr is the biological toxic metal response factor, and CF is the single element CF of the metal. The toxic response factors for Pb, Cd, Cu, Zn, Fe, Ni and Cr are 5, 30, 5, 1, 1, 1, 5 and 2, respectively (Gbadamosi et al 2018) and for Mn, 1 (Sinex and Helz 1981). Table 3 shows the categorization of pollution indices.

**Statistical analysis:** MS Excel was used to compile the data, and XLSTAT ver. 2023 was used to do Pearson's correlation matrix between intermetal and physicochemical parameters.

### **RESULT AND DISCUSSION**

**Physicochemical properties of soils:** Soil pH ranges from slightly alkaline to moderately alkaline (Table 4). The electrical conductivity varies from 0.08 to 0.24 ms/m. The soil moisture was higher in agriculture land (S4) where sewage water was used for irrigation as compared to other land uses. The mean soil organic carbon concentration was higher in industrial soil (S1) and sewage-irrigated agricultural land (S4). The soil nitrogen was highest in forest soil (S7). The soil phosphorus ranged from 14.57 to 44.81 kg/ha in all land use. The soil texture varies from sandy loam to clay.

Seasonal and spatial variations of heavy metals in soil: The spatial and seasonal mean metal concentration in the soil follows the order as Zn > Fe > Mn > Ni > Pb > Cr > Cu >Cd, respectively (Fig. 1, A-H).

The average seasonal concentrations of most of the metals were maximum during the post-monsoon for Cr, Pb, Ni, Cd and Zn (1.58, 1.60, 5.21, 0.35 and 31.40 mg/kg, respectively), and Fe, Mn and Cu (3.72, 2.65 and 0.48 mg/kg, respectively) was maximum during the monsoon (Fig. 1). During post-monsoon, the nonpoint sources mainly fertilization, surface runoff, irrigation, and agriculture after the rainy period could have added the metal load in the soils which eventually settled to the bottom (Hossain et al 2020). The spatial variations showed that the average maximum metal concentrations for Cr, Ni and Zn, i.e., 1.85, 3.54 and 17.52 mg/kg at the industrial site (S1). In agriculture soils, the highest concentrations were obtained for Cd and Pb (S3:

Table 3. Soil quality classification for multiple indices to assess heavy metals

Enrichme	nt factor (EF) <sup>1</sup>		Geoad	ccumulation index (Igeo) <sup>2</sup>	Contaminati	on factor (CF) <sup>3</sup>	Potential ecolog PEF	gical risk index RI)⁴
EF classes	Sediment quality	lgeo value	lgeo class	Sediment quality	CF Value	Pollution level	PERI	Indication
EF < 1	No enrichment	0-0	0	Unpolluted	CF < 1	Low	PER < 50	Low risk
EF < 3	Minor enrichment	0-1	1	Unpolluted to moderately polluted	1 ≤ CF ≥ 3	Moderate	50 < PER> 100	Moderate risk
EF 3-5	Moderate enrichment	1-2	2	Moderately polluted	3 ≤ CF ≥ 6	Considerable	100 < PER > 150	High risk
EF 5-10	Moderate severe enrichment	2-3	3	Moderately to highly polluted	CF > 6	Very high	150 < PER > 200	Very high risk
EF 10-25	Severe enrichment	3-4	4	Highly polluted			PER > 200	Extreme risk
EF 25-50	Extremely severe enrichment	4-5	5	Highly polluted to very highly polluted				
		5-6	> 5	Very highly polluted				

<sup>1 and 2</sup> Birch and Olmos 2008, <sup>3</sup> Hakanson 1980, <sup>4</sup> Guan et al 2014

1.26; 3.72 mg/kg), and Cu and Mn (S6: 0.55; 2.75 mg/kg), respectively. However, at forest was highest for Fe (4.33mg/kg).

During the pre-monsoon, the maximum concentrations of most of the metals (Cr, Cd, Cu: S3; Pb, Ni: S5) were in agricultural land uses. However, in monsoon season a higher concentration of Cr, Cd, Mn and Fe was reported from forest soils. In post-monsoon season also, an increased metal concentrations were observed from agriculture land uses. This increased levels of heavy metals in agriculture soil could be explained by the overuse of fertilisers, and proximity to textile industries adding the Cd load in the soils of sewage-



Fig. 1. (A-H) Seasonal and spatial variation in heavy metal concentration in the soil sample

irrigated agriculture lands (Bhuiyan et al 2011, Edogbo et al 2020). The higher values of Mn, Fe and Cu in Chomu agriculture site of Jaipur were also observed by Sharma (2023). The concentrations of Cd, Ni, Pb at industrial sites were in the range as reported by peri-urban soils of Bengaluru (Altaf and Subbarayappa 2022). The high Zn, Ni, and Mn content near Bagru industrial sites may be due to the wastewater generated by textile organizations (Pajot et al 2011).

The concentration recorded in this study was below the allowable limits, concerning the Indian (Awashthi 2000) and European (EU 2002) standard values. This could be explained by then the rejuvenation of the Dravyavati River (August 2016–August 2018), the ban on wastewater and garbage dumping and, five sewage treatment plants (STP) and cleaning the water of river. The limited anthropogenic and industrial activity during the COVID-19 pandemic could have resulted in reducing water and soil pollution sources, and diminish transfer of metal from water (river) to agriculture soil (sewage water irrigated).

#### **Pollution Indices**

**Enrichment factor (EF):** The mean enrichment (EF) factor for metals in the soil followed the order of Fe < Mn < Cr < Cu < Ni < Pb < Zn < Cd. The EF values indicated extremely severe enrichment of Cd and Zn (EF >25), severe enrichment of Pb (10-25), moderately severe enrichment for Ni (5-10), moderate enrichment of Cu (3-5), slightly Cr enrichment (EF < 3), Mn and Fe with no enrichment (EF < 1) (Table 3).

**Geoaccumulation index (Igeo):** The Igeo values indicated no contamination for Cr, Pb, Ni, Zn, Fe, Mn and Cu ( $\leq 0$ ), while moderate contamination for Cd (0-2) in all seasons for forest land use (S7) and sewage irrigated agriculture land (S3) (Table 3). The mean Igeo values for heavy metals followed an order: Cr > Cd > Zn > Cu > Pb > Mn > Ni > Fe. The average value of Cf< 1 indicated no pollution in the studied soils across various land uses. However, the Cf for Cd  $\geq$  3 showed moderate contamination in soils.

Contamination factor (Cd): The average contamination

factor (Cd) value for most of the metals, i.e., Cr, Pb, Ni, Zn, Fe, Mn, and Cu, was < 6, indicating a low degree of contamination, whereas the Cd values of cadmium lie between 6-12, indicating a moderate degree of contamination.

**Potential ecological risk:** The samples have a low potential ecological risk for Cr, Pb, Ni, Zn, Fe, Mn and Cu interpretd by the RI (risk index) value of less than 50 for these metals for all land uses in all seasons. However, the potential ecological RI of Cd lies between 190 and 380 (219), showing low to moderate pollution (Table 3).

During the monsoon season, metal levels decrease due to rainwater. The deposit of metals in the soil causes maximum enrichment during the post-monsoon. Distinct land use patterns, anthropogenic interruptions, past pollution record and its sources impact the soil pollution in various degree (Lee et al 2006). Previous study reports on increased EF values in urban soils are mostly due to industrial and road traffic emissions (Loska et al 2004, Ali and Malik 2010). The low metal background value (0.3 mg/kg) suggests an anthropogenic origin for the increased Cd concentration in the soil sample (Wedepohl 1991).

Intermetal correlation analysis: The inter-metal correlation analysis determined a significant positive correlation of Cr with Ni, Zn and Mn. In addition, a strong positive correlation was found between Cd, Pb, Ni, and Zn (R<sup>2</sup>> 0.8). However, there is a weak positive correlation between Pb and Cu (Table 5). The SOC shows a positive correlation with microelements (Fe, Zn and Cu) similar to the agriculture land of N.T.R district in Andhra Pradesh (Prasad et al 2023). A significant correaltion of Fe with SOC indicated pedogenic origin of Fe. The positive correlations between Cd, Pb, Ni and Zn indicate these metals come from similar anthropogenic sources, which could be the nearby industries, automobiles, Pb-Ni-based batteries, fertilizers etc. which might have contributed to the presence of these metals in the soil (Pajot et al 2011, Marwari et al 2012). The discharge of untreated wastewater into nearby land and water systems,

Table 4. Physicochemical characteristics of soil in semiarid region of Jaipur

14010 11	1 Hyeree entern	near enaracterior		innana region	orouipui			
Site code	e Site name	pН	EC (ms/m)	SM (%)	SOC (kg/ha)	SN (kg/ha)	SP (kg/ha)	Soil texture
S1	Sitapura	8.20±0.02	0.24±0	4.12±0.16	65.83±1.81	172.36±4.74	25.82±0.4	Clay
S2	Bagru	7.52±0.01	0.08±0	6.03±0.15	50.77±1.97	170.50±4.40	15.28±0.29	Clay
S3	Muhana	8.20±0.02	0.19±0	11.29±0.14	51.08±1.80	181.19±6.64	24.99±0.39	Clay loam
S4	Vidhani	7.53±0.03	0.20±0	11.46±0.23	65.52±2.25	183.51±4.06	44.81±0.39	Sandy loam
S5	Begas	7.79±0.04	0.16±0	5.91±0.16	50.66±1.91	165.85±5.46	32.93±0.33	Clay
S6	Chomu	8.24±0.02	0.16±0	8.53±0.12	56.38±1.59	175.15±4.99	26.26±0.42	Clay loam
S7	Nahargarh	7.64±0.02	0.08±0	10.12±0.11	43.75±1.74	194.43±5.20	14.57±0.31	Sandy loam

Values are in means ± standard error; EC electrical conductivity; SM soil moisture, SOC soil organic carbon, SN soil nitrogen, SP soil phosphorus

	pН	E.C	SM	SOC	SN	SP	Cr	Pb	Ni	Cd	Zn	Fe	Mn	Cu
pН	1	-0.02	-0.46	-0.01	-0.51	-0.52	0.67ª	0.25	0.36 <sup>b</sup>	0.00	0.43 <sup>b</sup>	-0.41	0.14	0.29
E.C		1	-0.08	0.79	-0.26	0.64	-0.43	0.20	0.02	-0.00	-0.21	0.17	-0.74	0.55ª
SM			1	-0.16	0.75	0.24	-0.21	0.40 <sup>b</sup>	-0.11	0.52ª	-0.10	0.43 <sup>b</sup>	0.05	0.25
SOC				1	-0.25	0.67ª	-0.11	-0.25	0.47 <sup>b</sup>	-0.51	0.21	0.23	-0.60	0.36 <sup>b</sup>
SN					1	-0.15	-0.32	0.07	-0.41	0.36 <sup>b</sup>	-0.25	0.77ª	0.27	-0.30
SP						1	-0.37	-0.06	0.27	-0.22	-0.04	0.04	-0.63	0.45
Cr							1	-0.23	0.58ª	-0.37	0.52ª	-0.27	0.59ª	0.22
Pb								1	-0.18	0.90 <sup>ª</sup>	-0.04	-0.22	-0.33	0.44 <sup>b</sup>
Ni									1	-0.52	0.89ª	-0.37	-0.20	0.26
Cd										1	-0.32	-0.00	-0.09	0.19
Zn											1	-0.42	-0.16	-0.03
Fe												1	0.20	-0.07
Mn													1	-0.20
Cu														1

Table 5. Intermetal correlation with soil parameters

EC electrical conductivity; SM soil moisture, SOC soil organic carbon, SN soil nitrogen, SP soil phosphorus. \*Correlation is significant at 0.01 level, \*Correlation is significant at 0.05 level

contaminates the soils and poses a threat to human health and the environment.

#### CONCLUSION

The present study evaluated the spatial and seasonal concentrations of heavy metals (Cr, Pb, Ni, Cd, Zn, Fe, Mn and Cu) in the various land uses, i.e., industrial, agriculture and forests in the semiarid region of Jaipur. Pearson's correlation suggests a common source for metals Cd, Pb, Ni and Zn, could be anthropogenic, whereas for Fe is paedogenic. The geochemical approaches indicated a low degree of contamination for most of the metals except for Cd showing moderate pollution. The study suggests that wastewater containing domestic and industrial waste must be monitored periodically by various pollution control agencies. The untreated or partially treated waste from industrial effluents should be treated properly and efficiently by sewage treatment plants to maintain the metal concentrations at prescribed standards.

#### **AUTHORS CONTRIBUTION**

Sonali Tiwari: conducted field sampling, laboratory analysis, and manuscript preparation, Naveen Kumar: collected the data and edited the manuscript, Priyanka Jatav: reviewed and interpreted the result, Archana Meena: analyzed and supervised the data.

#### REFERENCES

Ali SM and Malik RN 2010. Spatial distribution of metals in top soils of Islamabad City, Pakistan. *Environmental Monitoring and Assessment* **172**(1-4): 1-16.

- Altaf K and Subbarayappa CT 2022. Assessment of soil quality in rural and peri-urban areas of southern transect of Bengaluru by using principal component analysis. *Indian Journal of Ecology* **49**(6): 2076-2081.
- APHA (American Public Health Association) 2005. Standard Methods for the Examination of Water and Wastewater (21st ed.). Washington, DC.
- Awashthi SK 2000. *Prevention of Food Adulteration Act no 37 of 1954*. Central and State Rules as Amended for 1999, third ed. Ashoka Law House, New Delhi.
- Bhuiyan MAH, Suruvi NI, Dampare SB, Islam MA, Quraishi SB, Ganyaglo S and Suzuki S 2011. Investigation of the possible sources of heavy metal contamination in lagoon and canal water in the tannery industrial area in Dhaka, Bangladesh. *Environmental Monitoring and Assessment* **175**: 633-649.
- Birch G and Olmos MA 2008. Sediment-bound heavy metals as indicators of human influence and biological risk in coastal water bodies. *Ices Journal of Marine Science* **65**(8): 1407-1413.
- Chavan RB 2001. Indian textile industry-environmental issues. Indian Journal of Fibre and Textile Research 26: 11-21.
- Edogbo B, Okolocha E, Maikai B, Aluwong T and Uchendu C 2020. Risk analysis of heavy metal contamination in soil, vegetables and fish around Challawa area in Kano State, Nigeria. *Scientific African* 7.e00281.
- European Union EU. *Heavy Metals in Wastes*, European Commission on Environment 2002. http://www.ec.europa.eu/environment/waste/studies/pdf/heavymetalsreport.pdf
- Fabretti JF, Sauret N, Gal JF, Maria PC and Scharer U 2009. Elemental characterization and source identification of PM2.5 using Positive Matrix Factorization: The Malraux Road tunnel, Nice, France. *Atmospheric Research* **94**(2): 320-329.
- Gbadamosi MR, Afolabi TA, Ogunneye A, Ogunbanjo O, Omotola EO, Kadiri T, Akinsipo O and Jegede DO 2018. Distribution of radionuclides and heavy metals in the bituminous sand deposit in Ogun State, Nigeria: A multi-dimensional pollution, health and radiological risk assessment. *Journal of Geochemical Exploration* **190**: 187-199.
- Gee GW and Bauder JW 1979. Particle size analysis by Hydrometer: A simplified method for routine textural analysis and a sensitivity test of measurement parameters. *Soil Science Society of America Journal* **43**(5): 1004-1007.

- Guan Y, Shao C and Mei J 2014. Heavy metal contamination assessment and partition for industrial and mining gathering areas. *International Journal of Environmental Research and Public Health* **11**(7): 7286-7303.
- Hakanson L 1980. An ecological risk index for aquatic pollution control: A sedimentological approach. *Water Research* 14(8): 975-1001.
- Hossain MS, Ahmed MK, Sarker S and Rahman MS 2020. Seasonal variations of trace metals from water and sediment samples in the northern Bay of Bengal. *Ecotoxicology and Environmental Safety* **193**: 110347.
- Islam S, Ahmed MK, Raknuzzaman M, Habibullah-Al-Mamun M and Islam MK 2015. Heavy metal pollution in surface water and sediment: A preliminary assessment of an urban river in a developing country. *Ecological Indicators* 48: 282-291.
- Jackson ML 1973. Prentice Hall of Indian Soil Chemical Analysis (Pvt.) Limited, New Delhi.
- Sahu N, Pipalde D, Jain RC and Gupta SC 2024. Geo-statistical analysis of soil pollutants of Tannery industrial area, Kanpur. *Indian Journal of Ecology* 51(1): 113-117.
- Kurniawan TA, Chan YSG, Lo W and Babel S 2006. Comparisons of low-cost adsorbents for treating wastewaters laden with heavy metals. Science of the Total Environment 366(2-3): 409-426.
- Lee CSL, Li X, Shi WZ, Cheung SCN and Thornton I 2006. Metal contamination in urban, suburban and country park soils of Hong Kong: A study based on GIS and multivariate statistics. *Science of the Total Environment* **356**(1-3): 45-61.
- Loska K, Wiechuła D and Korus I 2004. Metal contamination of farming soils affected by industry. *Environment International* **30**(2): 159-165.
- Marwari R and Khan T 2012. Physiological and biochemical adverse effects of heavy metals on Brassica oleracea grown in Sanganer area, India. *Journal of Environmental Science and Engineering* **54**(2): 249-259.
- Muller GM 1969. Index of geoaccumulation in sediments of the rhine river. *GeoJournal* **2**(3): 108-118.
- Olsen SR, Cole CV, Watanbe FS and Dean LA 1954. *Estimation of available phosphorus in soils by extraction with sodium bicarbonate*. USDA. Circular **939**: 19-33.
- Pajot HF, Farina JI and De Figueroa LIC 2011. Evidence on manganese peroxidase and tyrosinase expression during

Received 22 June, 2024; Accepted 11 September, 2024

decolorization of textile industry dyes by *Trichosporon akiyoshidainum*. *International Biodeterioration and Biodegradation* **65**(8): 1199-1207.

- Parvez MS, Nawshin S, Sultana SS, Hossain MS, Khan MHR, Habib MA, Nijhum ZT and Khan R 2023. Evaluation of Heavy Metal Contamination in Soil Samples around Rampal, Bangladesh. *ACS Omega* 8(18): 15990-15999.
- Prasad PNS, Subbaiah VP, Rajasekhar N, Reddy VI and Rajanarasimha M 2023. Assessment of available major and micronutrient status of soils under varied cropping systems of N.T.R district, Andhra Pradesh. *Indian Journal of Ecology* **50**(2): 345-350.
- Sakata M and Asakura K 2011. Atmospheric dry deposition of trace elements at a site on the Asian-continent side of Japan. *Atmospheric Environment* **45**(5): 1075-1083.
- Sharma S, Hasan A, Kumar P, Salvi K and Kumawat L 2023. Assessment of heavy metals and physicochemical parameters of sewage water and soil in the different blocks of Jaipur district of Rajasthan, India. *The Pharma Innovation Journal* **12**(7): 3330-3333.
- Sinex SA and Helz GR 1981. Regional geochemistry of trace elements in Chesapeake Bay sediments. *Environmental Geology* **3**(6): 315-323.
- Singh H, Yadav M, Kumar N, Kumar A and Kumar M 2020. Assessing adaptation and mitigation potential of roadside trees under the influence of vehicular emissions: A case study of Grevillea robusta and Mangifera indica planted in an urban city of India. *PLoS One* **15**(1): e0227380.
- Subbiah BV and Asija GL 1956. A rapid procedure for the estimation of available nitrogen in soils. *Current Science* **25**: 259-260.
- Thakre YG, Choudhary MD and Raut RD 2012. Physicochemical characterization of Red and Black soils of Wardha region. *International Journal of Chemical and Physical Sciences* 1(2): 60-66.
- Turekian KK and Wedepohl KH 1961. Distribution of the elements in some major units of Earth's crust. GSA Bulletin **72**(2): 175-192.
- Walkley AJ and Black CA 1934. An examination of the method for determining soil organic matter and a proposed modification of the chromic acid titration method. *Soil Science* **37**: 29-38.
- Wedepohl KH 1991. Chemical composition and fractionation of the continental crust. Geologische Rundschau 80(2): 207-223.



Manuscript Number: 4350 NAAS Rating: 5.38

# Diversity and Phosphate Solubilization Potential of Rhizospheric Fungi from different Land-use of Mokokchung district, Nagaland, India

## Imlimenla Jamir, Wati Temjen\* and Tali Ajungla

Department of Botany, Nagaland University, Lumami-798 627, India E-mail: temjen.wati29@gmail.com

**Abstract:** This study investigates the rhizospheric fungal communities in soil samples from Mokokchung, Nagaland, India, collected from five land-use types: natural forest (NF), tree plantation (TP), bamboo plantation (BP), jhum fallow (JF), and shifting cultivation (SC). Various soil parameters, including temperature, pH, organic carbon, moisture, available nitrogen, phosphorus, and potassium, were analyzed. Soil fungal species were isolated in rose Bengal agar (RBA) and potato dextrose agar (PDA), and the diversity, richness, evenness, and abundance were estimated using diversity indices such as Shannon, Simpson, Berger-Parker, and Pielou's evenness. Results depict significant variations in soil properties influencing fungal populations across the different land-use types. NF exhibited the highest fungal diversity, taxa, and richness, while SC had the lowest, as reflected by the diversity indices. *Aspergillus* and *Penicillium* were the predominant genera. Additionally, eight fungal isolates demonstrated the ability to solubilize phosphate, with *Penicillium* emerging as a particularly promising candidate for further evaluation due to its high solubilization potential. This study highlights the impact of land-use on fungal diversity and the potential of phosphate-solubilizing fungi for sustainable agriculture.

Keywords: Land-use, Fungal diversity, Phosphate solubilizing fungi, Diversity indices, North East India

The plant root system influences the soil rhizosphere and micro-organisms present in the soil are subsequently influenced by the plant-root metabolites (Sharma and Shrivastava 2017). Therefore, depending on the land-use, different fungal flora is present. The fungal population regulates the ecosystem, stabilizes habitat, and controls various soil processes (Frac et al 2015). However, reports on the loss of biodiversity globally, which ultimately lowers ecosystem functionality, extinctions, and even ecosystem collapse in extreme situations are reported (Dunne and Williams 2009). The primary driver of this rapid loss of fungal biodiversity is converting forest land to agricultural systems and land-uses characterized by regular anthropogenic disturbances (Temjen et al 2021). Therefore, inventory is a valuable resource and ensures natural resource efficiency and sustainable utilization to prevent further biodiversity loss. The main instrument for ensuring ecological monitoring and addressing the biodiversity crisis is the use of diversity indices, which enable measurement of the two essential aspects of an ecosystem, i.e., richness and evenness (Stirling and Wilsey 2001, Morris et al 2014).

Phosphorus constitutes about 0.2% of the plant's dry weight and is essential for plant growth and metabolism (Widawati and Suliasih 2006). However, plants can only utilize a trace amount of the chemical P and a substantial amount is immobilized and left inaccessible to plants (El Attar

et al 2022). The economically and environmentally beneficial solution would be a microbial inoculant capable of dissolving sparingly soluble inorganic soil P (Alori et al 2017). Phosphate solubilizing fungi (PSF) can solubilize insoluble phosphate in the soil. Fungi, in particular, have a more remarkable ability to solubilize insoluble phosphate than bacteria (Zhang et al 2018). Mokokchung district is a hilly region in Nagaland state, North-East India. The everincreasing population in the state has amplified the pressure on the land. There is a trend of rapid conversion of natural forests into different land-use systems, which negatively affects the fungal community (Miah et al 2010, Temjen et al 2021). Therefore, the present work aims to study the diverse rhizospheric fungal populations from various land-use sites in Mokokchung district, Nagaland, India and estimate those fungi with the capacity to solubilize P for sustainable agriculture.

#### MATERIAL AND METHODS

**Site selection:** Five land-use sites under Mokokchung district, Nagaland, India, were selected (Table 1). The major type of soil in the region have alluvial soil, non-laterite red soil, and forest soil, with an average temperature of 27°C and 2500 mm of rainfall annually (Temjen et al 2022).

Soil sample: Composite soil layers at 0-30 cm were collected from the rhizospheric region of each site during

spring 2023. Soil temperature and pH were recorded on-site, soil organic carbon (SOC) was estimated using air-dried sieved soil samples (Walkley and Black 1934), and soil moisture was measured via the gravimetric method. The available nitrogen (N) was estimated as per the Kjeldahl method (1883), the available phosphorus (P) as per Bray's no. 1 extract method (Bray and Kurtz 1945), and exchangeable potassium (K) as per Trivedy and Goel (1986). All tests were performed in triplicates. Soil fungal species were isolated in rose bengal agar (RBA) and potato dextrose agar (PDA) plates supplemented with streptomycin sulphate (0.03g/L) and prepared following the serial dilution method (Selman and Waksman 1921). PDA and RBA plates were incubated at 25±1°C for 5-7 days. The fungi were identified with the help of standard literature (Gillman 1957, Nagmani et al 2006, Webster and Weber 2007), and the colony-forming unit was expressed as CFU/g<sup>-1</sup>(Johnson and Case 2006).

**Phosphate solubilizing fungi (PSF):** Pikovskaya (PVK) agar (Himedia) was utilized to screen their ability for phosphate solubilization. PVK agar was autoclaved at 121°C, and the subsequently sterilized PVK agar was transferred to fixed Petri plates. Fungal isolates from the pure culture were transferred to the PVK agar in three replicates. The mean values of the clearing zone diameter/halo formed around the PSF colony were recorded on the 5<sup>th</sup>, 10<sup>th</sup> and 15<sup>th</sup> days of incubation. The phosphate solubilization index (SI) was recorded by measuring the area of the clearing

Table	1.	Descri	ption of	stud	/ sites
10010		000011		oluda	

zone/halo area (Premono et al 1996).

**Statistical analysis:** Data were analysed using SPSS version 26.0 to perform the Duncan's multiple range test (DMRT). PAST 4.03 was utilized to calculate the number of taxa (S), individuals (N), dominance (D), Simpson (1-D), Shannon (H), Pielou's evenness (J), and Berger-Parker indices to determine the diversity, richness, evenness, and abundance of the fungal population.

#### **RESULTS AND DISCUSSION**

Soil properties: Soil temperature varied significantly among the sites, with the highest at JF, followed by SC (Table 2). This increased temperature at JF and SC may be attributed to the loss of vegetation cover at both sites and the resultant exposure of the soil layer to the sun. The highest pH was observed at SC, while the lowest was reported at NF and TP, respectively. The increased pH level at SC is attributed to the burning of the soil, which has been reported to raise pH (Temjen et al 2021). In contrast, the decreased pH levels at NF and TP may be attributed to the increased organic matter input from aboveground biomass at the sites. There was an also significant variation in soil moisture. Site NF possessed the highest moisture content. This is attributed to the increased vegetation and water-holding capacity of the site. The decreased soil moisture at SC may be due to the heating of the soil by slash and burning activities. SOC was found to be highest in BP and NF, while the lowest was reported under

Site	Vegetation	Period	Disturbances
Natural Forest (NF) 26.31203°N, 94.49328°E	Ageratum conyzoides, Amaranthus sp, Angiopteris sp, Artemisia vulgaris, Azadirachta indica, Eupatorium sp, Sonchus wightianus, Macaranga denticulate, Mikania cordata, Pueraria sp, Terminalia myriocarpa and Thysanolaena maxima	12 <sup>th</sup> year of fallow	No anthropogenic disturbance
Tree plantation (TP) 26.3123°N, 94.4942° E	Duabanga grandiflora	5 <sup>≞</sup> year of plantation	Little or no anthropogenic disturbance
Bamboo plantation (BP) 26.3130°N, 94.4949°E	Dendrocalamus sp	5 <sup>th</sup> year of plantation	Little or no anthropogenic disturbance
Jhum fallow (JF) 26.3130°N, 94.4959° E	Ageratum conyzoides, Eupatorium sp, Amaranthus sp	1 <sup>st</sup> year of fallow	Little or no anthropogenic disturbance
Shifting cultivation site (SC) 26.31065°N, 94.49405°E	Manihotesculenta	4 <sup>th</sup> cycle of cultivation	High anthropogenic disturbance

<b>Table 2.</b> Soli parameters across the during study si
--

			• •				
Site	Temperature (°C)	pН	Moisture (%)	SOC (%)	N (Kg ha⁻¹)	P (Kg ha <sup>-1</sup> )	K (Kg ha⁻¹)
NF	22 <sup>ª</sup>	5.4 <sup>ab</sup>	24 <sup>d</sup>	3.42°	478 <sup>ª</sup>	190°	31°
TP	<b>23</b> ⁵	5.1ª	19°	3.01 <sup>b</sup>	316°	170 <sup>⁵</sup>	23°
BP	21ª	5.5ªb	17 <sup>b</sup>	3.37⁴	303°	<b>169</b> ⁵	24 <sup>bc</sup>
JF	<b>24</b> <sup>ª</sup>	5.6 <sup>b</sup>	18 <sup>b</sup>	2.64ª	<b>240</b> <sup>♭</sup>	116 <sup>ab</sup>	21 <sup>b</sup>
SC	24 <sup>b</sup>	5.7⁵	16 <sup>ª</sup>	2.47ª	114ª	98°	19ª

\*Values in the same column with different superscripts are significantly different at the 5% level by DMRT

SC. This increased SOC at BP and NF may be attributed to the increased build-up of organic matter from both root and aboveground biomass at the respective sites. Bamboos, in particular, have high carbon sequestration capacity owing to their large biomass density and fast growth rate (Devi and Singh 2021). Meanwhile, lower SOC under SC and JF may be due to high rates of continuous cultivation for an extended period without a sufficient fallow phase, reducing the amount of organic material available in the soil (Yimer et al 2007). Similar trend was reported for the macronutrients in the study, i.e., N, P, and K. We report higher values under NF, TP, and BP as compared to JF and SC. This may be attributed to the increased mineralization process under the soils of the undisturbed sites as compared to the disturbed sites, i.e., SC and JF (Temjen et al 2021).

**Fungal diversity and diversity indices:** Thirty-one fungal species were isolated from the various land-use sites (Table 3), and the CFU is depicted in Table 4. *Aspergillus* and *Penicillium* were the most dominant genera, while *Absidia* sp. and *Fusarium* sp. were the least occurring genera. The

Table 3.	Fungal	diversity	of the	different	land-use	sites

Fungal species			Land-use		
	NF	TP	BP	JF	SC
1. Absidia sp.*				+	
2. Acremonium murorum ***	+	+	+		
3. A. strictum *			+		
4. A candidus *	+				
5. A. flavus ****	+		+	+	+
6. A. fumigatus ***	+	+	+		
7. A. versicolor *	+				
8. <i>A</i> , ***	+	+			+
9. <i>Aspergillu</i> s sp. 1 **	+	+			
10. <i>Aspergillus</i> sp. 2	+		+		+
11.Chladosporium chaldosporiodes *****	+	+	+	+	+
12. Eupenicillium javanicum ****	+	+	+		+
13. <i>Fusarium</i> sp. *				+	+
14. Geotrichum candidum***	+	+	+		
15. <i>Humicola</i> sp. **			+	+	
16. <i>Mortierella</i> sp. *****	+	+	+	+	+
17. Mucor circinelloides	+			+	+
18. M.hiemalis ****	+	+	+		
19. Paecilomyces carneus***		+	+	+	
20. P. farinosus ****	+	+	+		+
21. Penicillium brevicompactum ***	+		+	+	
22. P.citrinum ***		+	+	+	
23. P. digitatum *	+	+			
24. <i>Penicillium</i> sp. 1 ***	+			+	+
25. Penicillium sp. 2 ***	+	+	+		
26. <i>Penicillium</i> sp. 3 ****	+	+	+		
27. Penicillium sp. 4 ***	+	+		+	
28. <i>Rhizopus</i> sp. ***	+		+	+	
29. Scopulariopsis sp. *	+				
30. Trichoderma viridie ***	+	+	+		
31. Trychophyton sp. ***		+	+	+	

Species distribution is categorized based on their presence across the land-use sites: \*\*\*\*\* = very broad (present in all five sites); \*\*\*\* = broad (present in four sites); \*\*\* = moderate (present in three sites); \*\* = narrow (present in two sites); \* = very narrow (present in one site)

distribution of the fungi in the present study may be attributed to a combination of their host-specificity and suitable environmental conditions that enable their establishment. Further, there are observations that fungal colonies with better sporulation features dominate and colonize the culture plates better, resulting in more spores (Jena et al 2015). NF had the highest fungal taxa, followed by BP, TP, JF, and SC (Table 5). The dominance value, indicating the evenness of taxa distribution, was highest at SC and lowest at NF. The Simpson index of diversity was highest at NF, suggesting greater species dominance, while SC had the lowest. The Shannon index, reflecting species richness and evenness, was also highest in NF. Pielou's evenness index showed the highest evenness at JF and the lowest at SC. The Berger-Parker dominance index indicated that the most abundant species were more dominant at SC compared to other sites. The most fungi live in symbiotic association with their respective host, a change in land-use is expected to change the fungal community. Overall, NF exhibited the highest fungal diversity, likely due to its undisturbed nature and diverse vegetation. In contrast, SC had the lowest diversity, potentially due to anthropogenic activities like slash-andburn practices, which lead to lower soil moisture and reduced fungal evenness. Miah et al (2010) also reported lower fungal populations in disturbed sites than in forests. The dominance of certain species at SC highlights the impact of land-use changes on fungal communities and underscores the importance of understanding the entire fungal population for

#### ecosystem sustainability (Dai et al 2013)

Phosphate solubilizing fungi (PSF): Eight isolates were identified from different land-use sites with the capacity for phosphate solubilization. The SI varied significantly among the isolates (Table 6). The largest halo zone was observed under Penicillium sp. 3, while the smallest was reported under Aspergillus sp. 1. Aspergillus and Penicillium genera were particularly notable for their phosphate solubilizing capabilities. Turan (2006) and Alori et al (2017) also reported that these genera significantly reduced pH during phosphate solubilization. The process is understood to occur due to organic acids lowering pH or by complexing cations bound to phosphates (Johan et al 2021). The NF site also had the highest concentration of PSF, while JF and SC had the lowest concentrations. The greater abundance of PSF in NF may be attributed to diverse vegetation and increased organic matter, while the reduced PSF population in SC is likely due to high anthropogenic disturbances and burning practices, making the environment less suitable for these fungi (Temjen et al 2021).

#### CONCLUSIONS

The present findings identified eight fungal species capable of solubilizing phosphate, with higher PSF recorded in the natural forest and lower in shifting cultivation and *jhum* 

 Table 6. Solubilization index of PSF

 Table 4. Fungal colony forming unit of the different land-use sites

Site	Fungal CFU
NF	76 x10⁴/gª
ТР	40.5 x10⁴/g⁵
BP	45 x10⁴/g⁵
JF	26 x10⁴/g°
SC	24.5 x10⁴/g°

\*Values with different superscripts are significantly different at the 5% level by  $\mathsf{DMRT}$ 

|--|

Telete et e etaletti Eattert			
PSF	Phosphate	solubilisation	index (cm)
	5 <sup>th</sup> day	10 <sup>th</sup> day	15 <sup>th</sup> day
Acremonium murorum	2.56±0.12	2.78±0.02	2.84±0.23
<i>Aspergillus</i> sp. 1	1.02±0.23	1.42±0.13	1.53±0.22
<i>Aspergillu</i> s sp. 2	1.05±0.53	1.65±0.02	1.71±0.31
Mucor hiemalis	1.56±0.11	1.72±0.13	1.81±0.09
<i>Penicillium</i> sp. 1	1.14±0.14	2.58±0.25	2.94±0.18
Penicillium sp. 2	1.81±0.13	2.22±0.36	2.52±0.17
Penicillium sp. 3	3.20±0.25	3.50±0.24	3.54±0.22
Rhizopus sp.	2.12±0.05	2.64±0.22	2.87±0.09

· · · · <b>)</b>					
Indices	NF	TP	BP	JF	SC
Taxa (S)	21	17	20	15	12
Individuals (N)	152	81	90	52	49
Dominance (D)	0.077	0.096	0.085	0.088	0.132
Simpson (1-D)	0.922	0.903	0.914	0.912	0.868
Shannon (H)	2.815	2.564	2.681	2.566	2.221
Pielou's evenness (J)	0.924	0.905	0.894	0.947	0.893
Berger-Parker	0.190	0.185	0.155	0.173	0.224

fallow sites. Among these, species from *Penicillium* sp. 3 demonstrated superior solubilization potential, making it a promising candidate for further evaluation. Given the rising costs and adverse effects of chemical fertilizers, PSF offers a sustainable alternative for agriculture. Developing high-quality inoculants and consistently monitoring biological resources is essential. Future research should focus on field trials and large-scale experiments to assess the viability of PSF.

#### REFERENCES

- Alori ET, Glick BR and Babalola OO 2017. Microbial phosphorus solubilization and its potential for use in sustainable agriculture. *Frontiers in Microbiology* **8**: 258916.
- Bray RH and Kurtz LT 1945. Determination of total, organic and available forms of phosphorus in soils. *Soil Science* **59**: 39-46.
- Dai M, Bainard LD, Hamel C, Gan Y and Lynch D 2013. Impact of land use on arbuscular mycorrhizal fungal communities in rural Canada. Applied and Environmental Microbiology 79. https://doi.org/10.1128/AEM.01333-13
- Devi AS and Singh KS 2021. Carbon storage and sequestration potential in aboveground biomass of bamboos in North East India. *Scientific Reports* **11**(1):1-8.
- Dunne J and Williams R 2009. Cascading extinctions and community collapse in model food webs. *Philosophical Transactions of the Royal Society B* **364**(1524): 1711–1723.
- El Attar I, Hnini M, Taha K and Aurag J 2022. Phosphorus availability and its sustainable use. *Journal of Soil Science and Plant Nutrition* **22**(2): 5036-5048.
- Frac M, Jezierska-Tys S and Takashi Y 2015. Occurrence, detection, and molecular and metabolic characterization of heat-resistant fungi in soils and plants and their risk to human health. *Advances in Agronomy* **132**: 161-204.
- Gillman JC 1957. A Manual of Soil Fungi, Oxford and IBH publishing Calcutta, Bombay, New Delhi.
- Jena SK, Tayung K, Rath CC and Parida D 2015. Occurrence of culturable soil fungi in a tropical moist deciduous forest Similipal Biosphere Reserve, Odisha, India. *Brazilian Journal of Microbiology* 46(1): 85-96.
- Johan PD, Ahmed OH, Omar L and Hasbullah NA 2021. Phosphorus transformation in soils following co-application of charcoal and wood ash. *Agronomy* **11**(10), 2010. https://doi.org/10.3390/agronomy11102010
- Johnson TR and Case CL 2006. *Laboratory Experiments in Microbiology*, Benjamin Cummings, San Francisco, California, United States.
- Kjeldahl J 1883. Neue methode zur bestimmung des stickstoffs in organischen korpern [New method for the determination of nitrogen in organic substances]. Zeitschrift für analytische Chemie 22: 366-382.

Miah S, Dey S and Sirajul Haque SM 2010. Shifting cultivation effects

Received 28 June, 2024; Accepted 21 September, 2024

on soil fungi and bacterial population in Chittagong Hill Tracts, Bangladesh. *Journal of Forestry Research* **21**(3): 311-318.

- Morris EK, Caruso T, Buscot F, Fischer M, Hancock C, Maier TS, Meiners T, Müller C, Obermaier E, Prati D, Socher SA, Sonnemann I, Wäschke N, Wubet T, Wurst S and Rillig MC 2014. Choosing and using diversity indices: Insights for ecological applications from the German Biodiversity Exploratories. *Ecology and Evolution* **4**(18): 3514-3524.
- Nagmani A, Kunwar IK and Manoharachary C 2006. *Handbook of soil Fungi*, I K International, New Delhi.
- Premono ME, Moawad A and Vlek P 1996. Effect of phosphatesolubilizing Pseudomonas putida on the growth of maize and its survival in the rhizosphere. *Indonesian Journal of Agricultural Science* **11**: 13-23.
- Selman A and Waksman A 1921. Method for counting the number of fungi in the soil. *Journal of Bacteriology* 7(3): 339-341.
- Sharma T and Shrivastava DK 2017. Isolation and Characterization of PGPR from Rhizospheric Soil. *The International Journal of Engineering Science* **8**(4): 54-58.
- Stirling G and Wilsey B 2001. Empirical relationships between species richness, evenness, and proportional diversity. *The American Naturalist* **158**(3): 286-299.
- Temjen W, Singh MR and Ajungla T 2022. Effect of shifting cultivation and fallow on soil quality index in Mokokchung district, Nagaland, India. *Ecological Processes* **11**(1): 1-16.
- Temjen W, Singh MR and Jungla T 2021. Impact of fallow on soil health in Mokokchung district, Nagaland, India. Journal of Environmental Engineering and Landscape Management 29(4): 410-417.
- Trivedy RK and Goel PK 1986. Chemical and biological methods for water pollution studies, Environmental Publication, Karad, India.
- Turan M, Ataoglu N and Sahın F 2006. Evaluation of the capacity of phosphate solubilizing bacteria and fungi on different forms of phosphorus in liquid culture. *Journal of Sustainable Agriculture* 28: 99-108.
- Walkley A and Black IA 1934. An examination of the Degtjareff method for determining soil organic matter, and a proposed modification of the chromic acid titration method. *Soil Science* 37(1): 29-38.
- Webster J and Weber RWS 2007. *Introduction to fungi*, Cambridge University press, New York, USA.
- Widawati S and Suliasih D 2006. Augmentation of potential phosphate solubilizing bacteria (PSB) stimulate growth of green mustard (*Brasica caventis* Oed.) in marginal soil. *Biodiversitas* **7**(1): 10-14.
- Yimer F, Ledin S and Abdelkadir A 2007. Changes in soil organic carbon and total nitrogen contents in three adjacent land use types in the Bale Mountains, south-eastern highlands of Ethiopia. Forest Ecology and Management 242(2-3): 337-342.
- Zhang Y, Chen S, Wu Q, Luan G, Zhang P, Fang M, Wan Z, Hu F and Ye R 2018. Isolation and characterization of two phosphatesolubilizing fungi from rhizosphere soil of moso bamboo and their functional capacities when exposed to different phosphorus sources and pH environments. *PLoS ONE* **13**(7). https://doi.org/10.1371/journal.pone.0199625



Manuscript Number: 4351 NAAS Rating: 5.38

# Studies on Four Species of Genus *Hypolampurus* Hampson (Lepidoptera: Thyrididae) from Western Ghats, India

Amit Katewa and P.C. Pathania<sup>1</sup>

National Center for Vector Borne Diseases Control, Ministry of Health & Family Welfare, Delhi-110 054, India <sup>1</sup>High Altitude Regional Centre, Zoological Survey of India, Saproon, Solan-173 211, India E-mail: amitkatewa@gmail.com

**Abstract:** The communications deals with the studies on the morphotaxonomy of four species i.e., *angulalis* Moore, *striatalis* Swinhoe, *langualis* sp. nov. and *neostriatalis* sp. nov. belonging to genus *Hypolampurus* Hampson under family Thyrididae of order Lepidoptera. The species i.e., *H. angulalis* Moore is recorded for the first time from India. The female genitalia of species *striatalis* Swinhoe have been studied for the first time which is the type-species of the genus *Hypolamprus* Hampson. Beside this, the details on the morphology of the adults, male and female genitalia, nomenclature of the wing venation, distribution and materials examined for all the species have been discussed and illustrated in full details. Adichotomous key for their interspecific discrimination is also provided.

Keywords: Morphotaxonomy, Lepidoptera, Thyrididae, Hypolampurus Hampson, Western Ghats

Lepidoptera are scaly winged insects (Moths, butterflies and skippers) in the class Insecta which are having 1,58,570 species on global basis which are approximately to 9% of total Animalia. About 15,000 species belonging to 84 families fall in this order out of which 13,359 species under 78 families are moths and remaining are butterflies from India (Chandra 2011). Under this order one of the Superfamily Thyridoidea, the family Thyrididae comprise the picture winged leaf moths which are sometime included in Superfamily Pyraloidea, but all these not supported by cladistic analysis.

The genus Hypolampurus Hampson is a genus of the family Thrididae under order Lepidoptera. This genus was proposed for its type-species i.e., Microsca striatalis Swinhoe (Hampson 1893). Hampson (1894) reported only seven species from the then limits of India including and only one species i.e., striatalis Swinhoe from Western Ghats of India. Eleven specimens belonging to four species of genus Hypolampurus Hampson were identified from the Lepidoptera collection of Western Ghats which are housed in different museums. Owing to characters such as position of labial palpi, type of antenna (which are minutely ciliated in male) and on the basis of wing maculation and venation, the phenon has been found to be identical to the genus Hypolamprus Hampson of family Thyrididae. Besides above, the materials were comparison with the reference collections housed in different national museums and consultation of relevant literature (Warren 1896, 1896a and Whalley 1971), the aforesaid phenon has been sorted out into four species i.e., H. angulalis Moore, H. striatalis Swinhoe, H. langualis sp.

nov and *H. neostriatalis* sp. nov. A dichotomous key for their interspecific discrimination is also prepared.

#### MATERIAL AND METHODS

The adults of this genus were exclusively collected with the help of portable light trap (Plate 1) or fluorescent lights at night hours from different localities of Western Ghats of India (Plate 2) and put in the glass bottles charged with the ethyl acetate. The collected specimens were stretched and dried in stretching boxes and afterwards preserved well fumigated wooden boxes. The external morphological characters like antennae, proboscis, labial palpi, colour of frons and vertex, eyes, markings on patagia and tegula, wing venation and maculation, colour and markings of thorax, abdominal characters, position and number of tibial spurs were studied at first instance. To study wing venation, permanent slides of wings were made. For the preparation of permanent slides of fore and hindwings, the method proposed by Common (1970) and advocated by Zimmerman (1978) and to study the external male genitalia, methodology given by Robinson (1976) were followed. The photographs of genitalia were taken in stereo zoom binocular and wing venation drawn on graph papers. Multiple dissections of the studied species were made to examine the population variations. The terminology given by Klots (1970) was followed for the nomenclature purpose. All the collections were deposited in the Insect Museum, Department of Zoology & Environmental Sciences, Punjabi University, Patiala and National PAU Insect Museum, Department of Entomology, Punjab

Agricultural University, Ludhiana, Punjab. The geographical information of the localities in the states/district of Karnataka, Kerala, Maharashtra and Goa are provided below (Table1).

#### **RESULTS AND DISCUSSION**

### HYPOLAMPRUS Hampson

# *Hypolamprus* Hampson, 1893, *Fauna of British India (Moths)* 1:364.

#### Type-species: Hypolamprus striatalis Swinhoe.

Labial palpus slight, upturned and reaching above vertex of head; antenna minutely ciliated in male; forewing with costa straight, convex near apex, apex slightly produced and acute, termen wavy or evenly curved, tornus rounded, anal margin nearly straight,  $R_3$ ,  $R_4$  stalked,  $M_1$  from middle of discocellulars; hindwing with costa straight, apex rounded, termen wavy or evenly curved, tornus rounded, anal margin straight.

#### Key to the species of the genus Hypolamprus Hampson

1.	Abdomen brown, irrorated with pale red scales	2
-	Abdomen not as above	3
2.	Head with frons and vertex velvety red tinged with brown	angulalis Moore
-	Head with frons and vertex fuscous	<i>striatalis</i> Swinhoe
3.	Legs furnished with red brown scales	<i>langualis</i> sp. nov.
-	Legs furnished with white scales	<i>neostriatalis</i> sp. nov.

#### Hypolamprus angulalis Moore

Morova angulalis Moore, 1888, Lepid, Atkin : 214. Hypolamprus angulalis (Moore) Hampson, 1892.

**Male:** Alar expanse: 24 mm (Plate 3). Vertex and frons velvety red tinged with brown; labial palpus slight, upturned and reaching above vertex of head; antenna minutely ciliated; forewing with costa straight, convex near apex, apex slightly produced and acute, termen wavy, tornus rounded, anal margin nearly straight, ground color dull coffee color, irregularly striated with light red; hindwing with costa straight, apex rounded, termen wavy, tornus rounded, anal margin straight, ground color dull coffee color, irregularly striated with light red; underside of both wings suffused with silvery scales; tarsi striped white and brown; abdomen brown, irrorated with pale red scales.

**Wing Venation:** Forewing with vein Sc ending beyond middle of costa,  $R_1$  and  $R_2$  from cell,  $R_3$  and  $R_4$  stalked,  $R_5$  from upper angle of cell,  $M_1$  from middle of discocellulars,  $M_2$  and  $M_3$  from lower angel,  $Cu_1$  very near to lower angle,  $Cu_2$  from middle of cell, 1A present, 2A forked with 1A at base; hindwing with Sc+ $R_1$  orignating from base of cell,

anastomosing upper nervure at angle and separates to meet costa, Rs and  $M_1$  from upper angle of cell,  $M_2$  and  $M_3$  from lower angle,  $Cu_1$  very near to angel of cell,  $Cu_2$  from middle of cell, 1A and 2A present.

**Male genitalia:** Male genitalia with uncus strongly built, sclerotized, broadens toward middle and again tapering towards distal end, tip ending to a small spine, gnathos absent; tegumen broad, vinculum very long, deep U-shaped; saccus present; valva simple, flap-like, without any projections, setosed with long setae, tip blunt; transtilla membranous; juxta small; aedeagus moderately long, much broad towards proximal end, cornuti absent, ductus ejaculatorious enters apically.



Plate 1. Portable light trap



Plate 2. Area surveyed

#### Female genitalia: Not examined.

Material examined: India: Goa: Distt. Sanguem, Molem, 110mASL, 23.ii.2004, 2♂; Distt. Ponda, Ponda, 85mASL, 28.ii.2004, 1♂, coll. A. Katewa.

#### Old distribution: Rangoon (Hapmson 1892)

**Remarks:** The species *angulalis* Moore has earlier been reported from Rangoon by Hampson (1892). During the present studies, three specimens were collected from two different localities in the state Goa. In view of this, it is clear that the species is new record not only for the Western Ghats but the country, as well. The external male genitalia of this species is being studied for the first time.

#### Hypolamprus striatalis Swinhoe

*Microsa striatalis* Swinhoe, 1886, *Proceeding of theZoologicalSociety, London*. 1885: 875.

Hypolamprus striatalis (Swinhoe) Hampson, 1892

**Male and Female:** Alar expanse: 17mm (Plate 4). Vertex and frons fuscus; labial palpus slight, upturned and reaching above vertex of head; antenna minutely ciliated; forewing with costa straight, convex near apex, apex slightly produced and acute, termen evenly rounded, not wavy, tornus rounded, anal margin straight, ground color pale red brown, evenly striated with brown lines, a dull spot on upper angle of cell and one below cell; hindwing with costa straight, apex rounded, termen straight, tornus rounded, anal margin straight, tornus rounded, anal margin straight, tornus rounded, anal margin straight, ground color pale red brown, evenly striated with brown lines, medial band present; legs white; abdomen pale red brown.

**Wing Venation:** Forewing with vein Sc ending beyond middle of costa,  $R_1$  and  $R_2$  from cell,  $R_3$  and  $R_4$  stalked,  $R_5$  from upper angle of cell,  $M_1$  from middle of discocellulars,  $M_2$  and  $M_3$  from lower angel,  $Cu_1$  very near to lower angle,  $Cu_2$  from middle of cell, 1A present, 2A forked with 1A at base; hindwing with Sc+ $R_1$  orignating from base of cell, anastomosing upper nervure at angle and separates to meet costa, Rs and  $M_1$  from upper angle of cell,  $M_2$  and  $M_3$  from lower angle,  $Cu_1$  very near to angel of cell,  $M_2$  and  $M_3$  from lower angle,  $Cu_1$  very near to angel of cell,  $Cu_2$  from middle of cell, 1A and 2A present.

Male genitalia: Not studied.

Female genitalia: Female genitalia with corpus bursae

rounded, weakly sclerotized; signum not distinct; ductus bursae long and narrow, sclerotized towards corpus bursae; ostium bursae marked with two sclerotized structures; apophyses small; papilla analis weakly sclerotized, setosed with fine setae.

Material examined: India: Maharashtra: Distt. Sindhudurg, Amboli, 850mASL, 11.x.2005, 3♀♀, coll.

**Distribution:** Dharamshala, Calcutta, Poona, Nilgiris, Ceylon (Hampson 1892)

**Remarks:** The species *striatalis* Swinhoe has earlier been reported from Poona (Maharashtra) near the Western Ghats by Hampson (1892). During the present collection tours, it has again been collected from a locality (Amboli) which is nearer to Poona, from the same state which is very close to Poona. The present distributional data is additional to the old one. It can be inferred that the species has restricted distribution. Also, the female genitalia of this species which is type-species of the genus *Hypolamprus* Hampson has been studied for the first time.

#### Hypolamprus langualis sp. nov.

**Male:** Alar expanse: 19mm (Plate 5). Vertex and frons fuscus; labial palpus slight, upturned and reaching above vertex of head; antenna minutely ciliated; forewing with costa straight, convex near apex, apex slightly produced and acute, termen wavy, cilia white, tornus rounded, anal margin straight, ground color red-brown, evenly striated with red lines; hindwing with costa straight, apex rounded, termen wavy cilia white tornus rounded, anal margin straight, ground color rusty-red, striated with brown lines; legs furnished with red brown scales; abdomen fuscus.

**Wing Venation:** Forewing with vein Sc ending beyond middle of costa,  $R_1$  and  $R_2$  from cell,  $R_3$  and  $R_4$  stalked,  $R_5$  from upper angle of cell,  $M_1$  from middle of discocellulars,  $M_2$  and  $M_3$  from lower angel,  $Cu_1$  very near to lower angle,  $Cu_2$  from middle of cell, 1A present, 2A forked with 1A at base; hindwing with Sc+ $R_1$  orignating from base of cell, anastomosing upper nervure at angle and separates to meet costa, Rs and  $M_1$  from upper angle of cell,  $M_2$  and  $M_3$  from lower angle,  $Cu_1$  very near to angel of cell,  $Cu_2$  from middle of cell, 1A and 2A present.

 Table 1. Geographical information of the localities

State	District	Locality	Geographical coordinates	Altitude
Karnataka	Uttar Kannada	Ganeshgudi	15.2843° N, 74.5302° E	480mASL
Kerala	Palakkad	Mukkali	11.0587° N, 76.5402° E	560mASL
	ldukki	Vallakadavu	8.4750° N, 76.9195° E	780mASL
Maharashtra	Sindhudurg	Amboli	15.9647° N, 74.0036° E	850mASL
Goa	Sanguem	Molem	15.3758° N, 74.2269° E	110mASL
	Ponda	Ponda	15.4027° N, 74.0078° E	85mASL

**Male genitalia:** Male genitalia with uncus long and narrow, sickle-shaped, sparsely setosed, tip ending to a curved spine; tegumen broad; vinculum long, broad U- shaped; saccus present; valva simple, costa and succulus well differentiated, a pair of small projections present towards distal end, tip blunt without any bifurcation, setosed with long hairs; transtilla membranous; juxta present; aedeagus long and narrow, curved, cornuti absent, ductus ejaculatorious enters laterally.

#### Female genitalia: Not studied.

Material examined:India: Kerala: Dist. Palakkad, Mukkali,560mASL, 22.ix.2004, 01♂; Karnataka:Dist. Uttar Kannada,Ganeshgudi, 480mASL, 16.x.2005, 02♂, coll.

**Etymology:** The name of the species as *Hypolamprusl angualis* sp. nov.is proposed as an anagram of its closely allied species *i.e.*, *Hypolamprus angulalis* Moore.

**Remarks:** *Hypolamprusl angualis* sp. nov. is allied to the species *i.e., striatalis* Swinhoe from which it can be easily differentiated on the basis of ground color which is red in the former species and grey in latter. The former species is smaller in size as compared to the latter.

#### Hypolamprus neostriatalis sp. nov.

**Male:** Alar expanse: 13mm (Plate 6). Vertex and frons white; labial palpus slight, upturned and reaching above vertex of head; antenna minutely ciliated; forewing with costa straight, convex near apex, apex slightly produced and acute, termen wavy, cilia fuscus, tornus rounded, anal margin straight, ground color red-brown, striated with red lines; hindwing with costa straight, apex rounded, termen wavy, cilia fuscus, tornus rounded, anal margin nearly straight, ground color rusty-red, few ill-defined dark-red striae present; legs white; abdomen scaled with few shite scales.

**Wing Venation:** Forewing with vein Sc ending beyond middle of costa,  $R_1$  and  $R_2$  from cell,  $R_3$  and  $R_4$  stalked,  $R_5$  from upper angle of cell,  $M_1$  from middle of discocellulars,  $M_2$  and  $M_3$  from lower angel,  $Cu_1$  very near to lower angle,  $Cu_2$  from middle of cell, 1A present, 2A forked with 1A at base; hindwing with Sc+ $R_1$  orignating from base of cell, anastomosing upper nervure at angle and separates to meet costa, Rs and  $M_1$  from upper angle of cell,  $M_2$  and  $M_3$  from lower angle,  $Cu_1$  very near to angel of cell,  $Cu_2$  from middle of cell, 1A and 2A present.



Figs. A=Adult, B=Forewing, C=Hindwing, D=Male external genitalia, E=Aedeagus, F=Uncus and Gnathos ventral view, G=Valva, H=Uncus and Gnathos ventral view

Figs. A=Adult, B=Forewing, C=Hindwing, D=Female external genitalia, E=Ovipositor lobes, F=Ductus Bursae and Apophyses,, G=Corpus Bursae

Plate 3. Hypolamprus angulalis Moore

Plate 4. Hypolamprus striatalis Swinhoe





Figs. A=Adult, B=Forewing, C=Hindwing, D=Male external genitalia, E=Aedeagus, F=Uncus and Gnathos ventral view, G=Valva, H=Uncus and Gnathos ventral view

Plate 5. Hypolamprus langualis sp. nov.

#### Male genitalia: Not studied.

**Female genitalia:** Female genitalia with corpus bursae elongated, bag-like, sclerotized; signum not distinct; ductusbursae very long and narrow, coiled towards ostium bursae; ostium bursae simple; apophyses long, almost of same length small; papilla analis semicircular, setosed with fine setae.

Material examined: India: Kerala: Dist. Idukki, Vallakadavu, 780mASL, 11.ix.2004, 01♀; Dist. Palakkad, Mukkali, 560mASL, 19.ix.2004,01♀, coll..

**Etymology:** The specific name is derived as *Hypolamprus neostriatalis* sp. nov. being sibling to *Hypolamprus angulalis* Moore.

**Remarks:** *Hypolamprus neostriatalis* sp. nov.is allied to *Hypolamprus angulalis* Moore sp. nov. but either of them can be easily separated by their alar expanse and general maculation of the wings.

### CONCLUSIONS

The present study has information on the taxonomy and distribution of four species of genus *Hypolampurus* Hampson

Figs. A=Adult, B=Forewing, C=Hindwing, D=Female external genitalia, E=Ovipositor lobes, F=Ductus Bursae and Apophyses, G=Corpus Bursae

#### Plate 6. Hypolamprus neostraiatlis sp. nov.

from Western ghats of India. Seven species from the then limits of India including and only species i.e., *striatalis* Swinhoe from Western Ghats of India was reported (Hampson, 1894). The collected materials have been sorted out into four species *i.e., angulalis* Moore (New record from India), *striatalis* Swinhoe and two new species i.e., *langualis* sp. nov and *neostriatalis* sp. nov. reported from the area under reference. A dichotomous key for their interspecific discrimination is also provided.

#### ACKNOWLEDGEMENTS

Authors are grateful to MoEFCC (GoI), New Delhi for funding the project on Microlepidoptera. Thanks are also due to Head, Department of Entomology, Punjab Agricultural University, Ludhiana for providing necessary facilities. The financial help given by Indian Council of Agricultural Research (ICAR) New Delhi for "Network Project on Insect Biosystematics" is greatly acknowledged. We are thankful to Dr. V.V. Ramamurthy, Principal Scientist (Retd.), Division of Entomology, for giving permission to consult reference collections housed at NPC, IARI, New Delhi.

#### REFERENCES

- Chandra K 2011. Insect fauna of states and Union territories in India. ENVIS Newsletter 14(1): 189-218.
- Common IFB 1970. Lepidoptera: Yponomeutidae of Heard Island. Pacific Insects Monograph, 23: 229-233.
- Gaede M 1937. Catalogue of Gelechiidae. *Lepidopterorum* Catalogus **79**:1-630.
- Hampson GF 1893. The Fauna of British India, including Ceylon and Burma 1 Fauna Br. India (Moths) 1: v-xxii, 1-527 Hampson, G.F. 1894. *Fauna of British India including Ceylon and Burma, Moths.* Vol. II. Taylor and Francis, London, xxviii + 546 pp.
- Hampson GF 1892. Fauna of British India including Ceylon and Burma, Moths. Vol. I. Taylor and Francis, London, xiii + 527 pp.
- Hampson GF 1895. Fauna of British India including Ceylon and Burma, Moths. Vol. IV. Taylor and Francis, London, xxiii + 588 pp.
- Hampson GF 1896. *Fauna of British India including Ceylon and Burma, Moths.* Vol. V. Taylor and Francis, London, xxviii + 594 pp.
- Klots AB 1970. Taxonomists Glossary of Genitalia in Insects. Munksgaard, Copenhagen Lepidoptera. pp. 115-139 in Tuxen.
- Robinson GS 1976. The Preparation of slides of Lepidoptera genitalia with special reference to microlepidoptera.

Received 03 June, 2024; Accepted 21 September, 2024

Entomologist Gazette 27(2): 127-132.

- Warren W 1897a. New genera and species of moths from the Old-World regions in the Tring Museum. *Novitates Zoologicae* **4:** 12-130.
- Warren W 1895. New genera and species of Pyralidae, Thyrididae and Epiplemidae. *The Annals and Magazine of Natural History* **16**(6): 460-477.
- Warren W 1896. New genera and species of Pyralidae, Thyrididae and Epiplemidae. *The Annals and Magazine of Natural History* **17**(6): 94-106, 131-150.
- Warren W 1896a. New species of Pyralidae from Khasia Hills. *The Annals and Magazine of Natural History* **18**(6): 107-119, 163-177, 214-232.
- Whalley PES 1964a. Catalogue of the World genera of the Thyrididae (Lepidoptera), with type selection and synonymy. *Annals and Magazine of Natural History* **13** (7):115-127.
- Whalley PES 1967. A revision of the Thyrididae of Madagascar. Faune de Madagascar, Insectes, Lépidoptères, Thyrididae. *Faune de Madagascar* 24:1-52, pls 1-14.
- Whalley PES 1976. Tropical Leaf Moths, A Monograph of the subfamily Striglinae (Lepidoptera: Thyrididae). London, British Museum (Natural History). 194 pp., 68 pls.
- Whalley PES 1971. A revision of the genus *Canaea* Walker (Lepidoptera: Thyrididae). *Bulletin of the British Museum* (*Natural History*) *Entomology* **26**(3):161-179.
- Zimmerman EC 1978. Microlepidoptera. *Ins. Hawaii*, vol. 9. University Press of Hawaii, Honololu. xviii + 1903pp.



# Potential of Twin Key Management Practices: Higher Colony Strength and Lower Honey Extraction Frequency in Improving Honey Quality of *Apis mellifera*

## Sumit Saini, O.P. Chaudhary, Vadde Anoosha<sup>1</sup> and Lalita<sup>\*</sup>

CCS Haryana Agricultural University, Hisar -125 004, India <sup>1</sup>Dr. YSRHU, Horticulture Research Station, Ambajipeta- 533 214, India \*E-mail: lalitaentohau@gmail.com

**Abstract:** Present study signifies the benefits of scientific management practices which eventually will earn more returns to beekeepers in India. Colonies with initial 10 frames strength and two extractions in the seasons were observed to produce honey of good quality and most economical. In 5 frame colonies moisture remained well below (17.91%) the maximum standards (20%) set for "special" grade honey and was still lower (17.80%) in medium strength (10 frames) and the strongest (15 frames) colonies (17.43%). However, under regular honey extraction, the moisture content was maximum in colonies of lowest strength (20.02%) compared to in 10 and 15 frame colonies. Other quality parameters like F: G ratio, sucrose, hydroxy methyl furfural (HMF) was observed above standards in 10 frame and two extraction colonies. Total reducing sugars was highest (82.23 %) in 15 frame single extraction colonies and 80.24 % in 10 frame colonies with similar extraction frequencies. Present findings clearly support the role of extraction frequencies in improving honey quality as we decrease frequency quality improves remarkable even in low strength colony. Strength is also a very important factor in deciding honey quality.

#### Keywords: Honey quality, Extraction, Quality Parameters, Moisture, Honey sugars

Beekeepers desire to make beekeeping more and more profitable and keep on extracting honey at regular intervals during honey flow season. Scientific knowledge of beekeeping to beekeepers is a major constraint and they follow either traditional or own developed non-scientific practices which degrades quality of honey. Regular extraction of honey at fixed interval of 7-10 days and extraction of un-ripened honey from brood chamber and reluctant to use supers for honey production are some causes of quality degradation. In India winter is the major honey flow season from mustard crop followed by few minor seasons up to early summer (December to May) followed by a long dearth from summer through rainy season till November (Chaudhary 2005). The Bureau of Indian Standards (BIS) standard (No. IS: 4941-1974), now Food Standard and Safety Authority of India (FSSAI 2010) through its regulation (No. 5.7.4) also recommends extracting honey from ripe and sealed honey combs and by default present Indian honey does not confirm to these standards. Besides poor quality, such practices are the root cause of low colony productivity. The production of honey and growth of colony is dependent over colony strength & honey extraction frequency. Right colony strength and extraction frequency gives better economical returns in beekeeping enterprise (Bhusal 2011, Saini et al 2018b, 2022). Honey quality parameters contain moisture content, ash content, total

soluble solids (TSS), acidity, reducing & non-reducing, fructose: glucose ratio and HMF content (Codex 2001). Beekeeping now days facing four major problems *viz*. low colony productivity, colony debilitation / mortality, extremely poor honey quality and ever increasing cost of production (labor and migration) resulting in non-stability of the beekeeping enterprise. Present study on colony strength and honey extraction frequencies was conducted to formalize some practices for beekeeping that improves colony health and honey quality.

#### MATERIAL AND METHODS

Present investigations were carried out on western honey bee, *A. mellifera* L. colonies at beekeeper's apiary during major honey flow season of mustard crop. Initial colony strength (ICS) and for equalization of colonies standard protocol were followed Delaplane et al (2013) and Saini et al (2022). After equalization colonies were migrated to the experimental site at RDS Farm of CCS Haryana Agricultural University, Hisar on the mustard crop which was about to flowering. Each colony with specified frame strength was labeled and placed in diamond orientation at 10 feet row and 5 feet colony distance. Bee colony with 5, 10 and 15 frames strength were the treatments, these colonies were then extracted at different frequencies (once, twice and regular) considering single colony as one replication and four replications were kept in each treatment, total 36 colonies were under experiment (12 in each colony strength)

**Honey extraction frequencies:** To determine the effect of honey extraction frequencies on honey quality four colonies in each colony strength of 5, 10 and 15 frame were extracted once for honey in the season, four were extracted twice and remaining four in each strength were extracted regularly or following beekeeper's practice. During the experiment, 36 colonies (Colony strength and honey extraction frequencies) were used.

**Collection of sample:** Samples of honey for each treatment were collected in wide mouth food grade plastic bottles (Cello) on the day of honey extraction in the field, specific number were given to each sample and stored in a closed big plastic box at home under favorable temperature condition so that any change in property could be avoided before going for physio-chemical analysis. All precautions were taken care to keep the quality unaltered during storage.

**Preparation of sample before processing:** Samples for quality testing, were placed in hot water bath at 40°C for liquefaction, strained through muslin cloth and stored in clean containers. Three honey sub samples from each treatment were drawn, then mixed together and a composite sub sample of 50 g was made for further processing. Honey left in the containers after making composite sample was again sealed in air tight jars. Three sets (replications) were made from the composite sample and subjected to testing for different honey quality parameters.

**Physio-chemical characteristics of honey:** The sub samples (in triplicate) were prepared from different treatments were analyzed as per standard procedures prescribed by the Bureau of Indian standard (IS4941:1994) unless specified in text. Standard procedures specified in FAO Manuals of Food Quality Control (FAO 1984), ISI Hand book of Food Analysis (Part II), and in testing manual of food safety and security authority of India (FSSAI 2010) were followed for honey quality testing.

#### **RESULTS AND DISCUSSION**

Honey quality in different colony strengths under different honey extraction regimes: In regular honey extraction, the moisture content was maximum in colonies of lowest strength (19.33%) compared to in 10 and 15 frame colonies where values were significantly lower but at par with each other (Table 1). The honey from lowest colony strength had least specific gravity (1.407) compared to the stronger colonies with similar values (1.409). On the contrary total reducing sugars was minimum in the lowest strength colonies 78.48 percent and increased progressively with increase in colony strength (79.22 and 79.43%). Colony

Table 1. Comparison of	honey qual	lity paramet	ers in colo	nies with varie	d frame str	ength under	different h	ioney extractior	n regime			
Quality parameters	Regula	ar extraction i. (frames/	n colony str colony)	engths	2 extraction	s in colony str	engths (fra	mes/colony)	1 extraction	in colony stre	ingths (fram	ies/colony)
I	5	10	15	CD (p=0.05)	5	10	15	CD (p=0.05)	5	10	15	CD (p=0.05)
Moisture (%)	19.33	19.05	19.05	0.06	19.10	18.80	19.00	0.06	18.60	18.40	17.40	0.07
Total reducing sugar (%)	78.48	79.22	79.43	0.20	78,96	79.65	79.78	0.05	79.87	80.24	82.23	0.05
Fructose (%)	40.06	41.18	40.85	0.07	40.60	40.80	41.07	0.02	40.66	41.15	41.89	0.04
Glucose (%)	38.42	38.04	38.59	0.16	38.36	38.85	38.71	0.06	39.21	39.09	40.34	0.08
Sucrose (%)	2.56	2.23	2.00	0.06	2.57	2.25	2.23	0.03	2.00	1.87	1.40	0.07
Acidity (%)	0.076	0.066	0.075	0.004	0.077	0.084	0.084	0.002	0.070	0.086	0.080	0.012
Ash (%)	0.091	0.082	0.093	0.005	0.088	0.095	0.094	0.003	0.086	0.095	060.0	NS
F/G ratio	1.042	1.082	1.059	0.005	1.058	1.051	1.061	0.002	1.036	1.053	1.038	0.003
H.M.F.	6.91	7.00	7.00	SN	7.103	7.038	7.000	0.048	7.107	7.190	7.190	NS
Specific gravity	1.407	1.409	1.409	0.000	1.408	1.410	1.409	0.001	1.410	1.411	1.414	0.001

#### Sumit Saini et al

strength under regular honey extraction had no effect on HMF content in the honeys. The moisture content was lowest in 10 frame colonies (18.80%) followed by 15 frames (19.00%). The increase in TRS was significant with increase in colony strength (78.96, 79.65 and 79.78%) from 5, 10 and 15 frame strengths, respectively. Fructose content also increased with colony strength while the glucose content was least in 5 frame colonies (38.36%) and maximum in 10 frame colonies (38.85%). The acidity and ash content was lowest in the weakest colonies (0.077 and 0.088%) that increased significantly in higher strength colonies.

Honey quality in different colonies strengths under different honey extraction regimes: In 15 frame colonies lowest moisture content 17.43%, signifying the highest honey quality in 15 frame colonies in the single honey extraction regime (Table 2). The specific gravity also had an increasing trend with increase in colony strength. TRS content increased from 80.38 percent in 5 frame colonies to 81.89% in 10 frame colonies and was the maximum in the strongest colonies (82.04%). The fructose content also exhibited similar increase from 40.90 to 41.71% while the glucose content was minimum in 5 frame (39.48%) to become maximum (40.57%) in 10 frame colonies. The sucrose exhibited reverse trend with colony strength being maximum in 5 frame (1.88%) and reducing in 10 (1.81) to become the least (1.26%) in strongest colonies. Acidity observed high in 5 frame colony (0.173) which decreases with increase in colony strength became minimum in 15 frame (0.076) and ash content where maximum in 15 frame colonies (0.093%) then 10 and 5 frame colonies (0.085 and 0.081, respectively).

Under twice honey extraction regimes, sucrose content was higher in 5 frame colonies which indicate un-ripened honey or nectar. F:G ratio showed pattern that was highest in the strongest colonies and lowest in weakest colony. Under regular honey extraction, the moisture content was maximum in colonies of lowest strength (20.02%) compared to in 10 and 15 frame colonies where values were significantly lower and varies significantly with each other. The honey from lowest colony strength had least specific gravity (1.405) compared to the stronger colonies which had value higher (1.409) than this. The total reducing sugars was minimum in the lowest strength colonies 78.54 percent and increased progressively with increase in colony strength (79.01 and 79.38%). Sucrose content was least in the strongest colonies (1.699%), increased to 2.01 % in 10 and 2.44 percent in 5 frame colonies. Ash content in honey was observed maximum in strongest colony (0.094%) which decreases significantly in 10 frame (0.089) and minimum in 5 frame colonies (0.082). The results for all the honey quality

Quality parameters	Reguli	ar extraction i (Frames,	in colony str /colony)	engths	2 extraction	s in colony stı	engths (Frai	mes/colony)	1 extractio	ר in colony str	engths (Frar	nes/colony)
I	5	10	15	CD (p=0.05)	5	10	15	CD (p=0.05)	5	10	15	CD (p=0.05)
Moisture (%)	20.022	19.809	19.652	0.016	19.860	19.697	19.345	0.021	17.907	17.807	17.433	0.115
Total reducing sugar (%)	78.542	79.011	79.384	0.049	78.433	79.385	79.597	0.019	80.387	81.897	82.047	0.025
Fructose (%)	40.051	40.931	41.241	0.034	40.183	40.713	40.879	0.021	40.900	41.390	41.713	0.060
Glucose (%)	38.492	38.080	38.144	0.037	38.248	38.675	38.718	0.028	39.487	40.573	40.333	0.146
Sucrose (%)	2.447	2.019	1.699	0.015	2.363	1.692	1.505	0.023	1.887	1.810	1.267	0.045
Acidity (%)	0.109	0.112	0.097	0.002	0.109	0.085	0.082	0.004	0.173	0.083	0.076	0.006
Ash (%)	0.082	0.089	0.094	0.002	0.079	0.085	0.079	0.002	0.081	0.085	0.093	0.004
F/G ratio	1.040	1.075	1.082	0.002	1.051	1.053	1.056	0.001	1.035	1.020	1.034	0.005
Hydroxy methyl furfural (mg/kg)	6.888	6.872	6.663	0.018	7.222	7.175	7.055	0.020	7.073	6.937	6.990	0.049
Specific gravity	1.405	1.408	1.409	0.001	1.399	1.404	1.410	0.001	1.412	1.413	1.415	0.001

**Annexure 1.** Various honey standards by agencies

Characteristics	P.F.A	BIS	6 (4941-19	994)	AG	MARK (20	08)	CODEX
		Special	"A"	Standard	Special	"A"	Standard	(2001)
Specific gravity at 27 C (Min)	-	1.37	1.37	1.37	1.40	1.37	1.35	
Moisture % (Max)	25	20	22	25	20	22	25	21
Reducing sugars % (Min)	65	70	65	65	70	65	65	65
Sucrose % (Max)	5	5	5	5	5	5	5	5
Ratio L/D (Min)	0.95	1	1	1	1	0.95	0.95	
Ash% (Max)	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.6
Acidity as formic acid % (Max)	0.20	0.20	0.20	0.20	0.20	0.20	0.20	40
Fiehe 's test		Neg	ative			Negative		
Aniline Chloride test	-	-	-	-		Negative		
Total pollen count and plant elements / g A Apiary honey (Max)	-	50,000	50,000	50,000	50,000 50,000	50,000 50,000	50,000 50,000	-
H.M.F. mg / Kg (Max)	-	80	80	80	80	80	80	40
Optical density % (Max)	-	0.3	0.3	0.3	0.3	0.3	0.3	-
Diastase (Min)	-	-	-	-	3	3	3	3
Water insoluble matters % (Max) for pressed honeys	-	-	-	-	0.50 0.10	0.50 0.10	0.50 0.10	0.1 0.5

BIS -If Fiesh's test is Positive and HMF content is more than 80 mg/kg then L/D ratio should be more than 1.00

1. If the pollen count is higher than 50,000/gm. Then honey may be categorized as squeezed honey.

2.If the Fiehe's test is positive but Hydroxyl Methyl Furfural (HMF) content is below 80ppm then honey may be accepted.

If granulated, it should be warmed at 60 till the crystals have dissolved completely

parameters were remarkable under single and two honey extractions over regular extractions regime. Similar high quality of honey was recorded in strongest (15) and 10 frame colonies over the weakest 5 frame colonies. The moisture content which is major quality criteria even in 5 frame colonies (17.91%) remained well below the maximum standards (20%) set for "special" grade honey and was still lower in medium (17.80%) and the strongest colonies (17.43%). The specific gravity, total reducing sugars, fructose, glucose, ash content (0.081 to also increased with increase in colony strength. The acidity and HMF content were least in these honeys and confirm to best quality honey. Reports of Szabo et al (1992) and Szabo & Lefkovitch (1990) about the effects of the frequency of honey extraction (4, 2 and 1 times) quantity and quality of honey on A. mellifera colonies indicating higher moisture content in colonies with applications of 5 supers and 4 honey removals (19.0%) and the lowest with 2 honey removals and 12 supers (16.7%), highest diastase (amylase) numbers for honey removed once (28.6) followed by that removed twice (26.5) and 4 times (23.2) amply support the present findings.

There is significant role of extraction frequencies in improving honey quality as we decrease frequency quality improves remarkable even in low strength colony. Strength is also a very important factor in deciding honey quality. In spite improving honey quality less number of honey extractions increases economic returns to the farmers (Saini et al 2022). The results for all the honey quality parameters were remarkable under single and two honey extractions over regular extractions. Colonies with initial 10 frames strength and two extractions in the seasons were observed to produce honey of good quality and most economical as per findings of the study. The moisture content which is major quality criteria even in 5 frame colonies (17.91%) remained well below the maximum standards (20%) set for "special" grade honey and was still lower in medium (17.80%) and the strongest colonies (17.43%). However, under regular honey extraction, the moisture content was maximum in colonies of lowest strength (20.02%) compared to in 10 and 15 frame colonies. If beekeepers in India want to improve honey quality and to earn more returns from this enterprise honey extraction should be limit to one or two in the season leaving enough food stores for bee to survive dearth period.

#### REFERENCES

- Bhusal SJ, Kafle L, Thapa RB and Shih CJ 2011. Effect of colony strength on the performance of honeybees (*Apis mellifera*) in Nepal (Hymenoptera: Apidae). *Sociobiology* **58**(2): 435-447.
- Chaudhary OP 2004. Prospects of beekeeping with *Apis mellifera* Linn. in Haryana. *Indian Bee Journal* **66**(3-4): 92-101.
- Chaudhary OP 2005. Incidence and intensity of *Varroa destructor* on *Apis mellifera* L. honey bees in Haryana migratory region. *Indian Bee Journal* **67**(3-4): 121-127.
- Codex 2001. Codex Alimentarius standard for honey 12-1981.

Agmark:

Revised Codex standard for honey. *Standards and standard methods* (Vol. 11). Retrieved D.

Delaplane KS, Van Der Steen J and Guzman-Novoa E 2013. Standard methods for estimating strength parameters of Apis mellifera colonies. *Journal of Apicultural Research* **52**(1): 1-2.

FAO 1984. Manuals of Food Quality Control. pp119

- FSSAI 2010. http://old.fssai.gov.in/Portals/0/Final\_Regulations\_ 2010.pdf. Food Standard and Safety Authority of India, Regulation 5.7.4 Honey
- Saini S, Chaudhary OP and Anoosha V 2018a. *Apis mellifera* colony productivity and growth influenced by initial frame strength: Farmer's perspective. *The Indian Journal of Agricultural Sciences* **88**(10): 1618-1623.

Saini S, Chaudhary OP and Anoosha V 2018 b. Relationship of

Received 22 July, 2024; Accepted 11 September, 2024

population size and extraction frequency with honey production in *Apis mellifera* colonies. *Journal of Entomology and Zoology Studies* **6**(3): 1374-1377.

- Saini S, Chaudhary OP and Anoosha V 2022. Maximizing income through beekeeping (*Apis mellifera*) by following right management practices. *Journal of Apicultural Research* **61**(1): 19-25.
- Szabo TI, Sporns P and Lefkovitch LP 1992. Effects of frequency of honey removal and empty comb space on honey quantity and quality. *American Bee Journal* **132**(12): 815-816.
- Szabo TI and Lefkovitch LP 1990. Effects of honey removal and supering on honey bee colony gain. *American Bee Journal* **130**(12):815-816.



# Morphological and Molecular Confirmation of *Thrips palmi*, Karny 1925 (Thripidae) and *Haplothrips tenuipennis* Bagnall, 1918 (Phlaeothripidae) of Order Thysanoptera in Muthalamada Mangoes of Kerala, India

Syed Mohamed Ibrahim S., Malini Nilamudeen<sup>1</sup>, Pratheesh P. Gopinath<sup>1</sup> and Linitha Nair<sup>2</sup>

Department of Entomology, College of Agriculture, Kerala Agricultural University, Trivandrum-695 522, India <sup>1</sup>Department of Agricultural Statistics, College of Agriculture, Trivandrum-695 522, India <sup>2</sup>Department of Agrl. Meteorology, RARS (SZ), Vellayani, Trivandrum - 695 522 E-mail: smis1997.s@gmail.com

**Abstract:** Periodic incidence of sucking pests in Muthalamada mangoes, Palakkad, Kerala has caused severe setback to mango growers. Symptoms of damage include malformation of mango panicles and bronzing of mango fruits which resulted in the reduced marketability of the produce. A random survey was conducted in mango orchards of these regions during the flushing, flowering, and fruit setting stage of crop during September 2022 to May 2023. Collected thrips specimens were subjected to morphological identification done with permanent slide technique of taxonomic key characteristics. Because *T. palmi* Karny and *Haplothrips tenuipennis* Bagnall has close physical resemblance to other thrips species, we used the mitochondrial cytochrome oxidase 1 (mtCO1) gene as a molecular marker. BLAST analysis of this sequence allowed us to identify the collected specimen as *T. palmi* and *H. tenuipennis*.

Keywords: Muthalamada, Mango city, Mitochondrial COI, Thrips palmi, Haplothrips tenuipennis

Palakkad district of Kerala is the mango hub ('mango city') with have maximum production of 55120 MT and area of 10068 ha (Ecostatkerala 2020). Kerala is also a hotspot for climate change and variability particularly Muthalamada experiences a lot of climatic variabilities which causes fluctuations in the population of insect pests mainly thrips. Thrips causes scars on the surface of fruit and foliage in addition to feeding and reproduction on inflorescences (Kumar et al 2012). T. palmi Karny and H. tenuipennis Bagnall was reported on mango inflorescence from India (Tandon and Verghese 1987 and Ramasubbarao and Thammiraju 1994). Krishnamoorthy and Visalakshi (2012) reported that of the several sucking pests infesting mango inflorescence, T. palmi and H. tenuipennis caused considerable loss. The unique properties of thrips make morphological key identification a laborious process that requires accuracy and speed (Asokan et al 2007). Nuclear markers such as ribosomal ITS have been utilized for molecular identification (Farris et al 2010). Nevertheless, it has been found that the best method for molecular identification within the genus Thrips is mitochondrial COI (DNA barcoding). The objective of paper is to confirm the presence of T. palmi and H. tenuipennis through morphological key characters and molecular DNA barcoding (Asokan et al 2007) and NJ (neighbour joining) or maximum likelihood analyses help for phylogeny study of such species.

### MATERIAL AND METHODS

**Sampling and morphological identification:** Random survey was conducted in five orchards (Table 1) of Muthalamada panchayat, Palakkad, Kerala during the year 2023 provided a sample, which thrips were collected from five randomly chosen trees in each orchard by adopting  $CO_2$  method (Aliakbarpour and Che Salmah 2010). The samples were subsequently stored in 70% alcohol, and a stereomicroscope was used to sort each individual thrips. Adult thrips were mounted using procedure of permanent slide technique (Mound 2005). The specimens were divided into suborders, Terebrantia and Tubulifera and placed in vials containing 70% alcohol before being transmitted to the NBAIR and ZSI for species confirmation.

#### DNA Barcoding using Universal Primers of COI

**Genomic DNA isolation:** Using the NucleoSpin® Tissue Kit (Macherey-Nagel) and the manufacturer's instructions, genomic DNA was extracted from the tissues. The extracted DNA stored at  $-20^{\circ}$  C. PCR was carried out with universal primer forward primer (LCO1490) 5'-GGTCAACAAATCATAAAGATATTGG-3' and reverse primer (HCO2198) 5'-TAAACTTCAGGGTGACCAAAAAATCA-3' (Vrijenhoek 1994), to amplify a 478 bp and 483 bp fragment of COI gene for *T. palmi* and *H.tenuipennis*. PCR amplification was carried out by following Thermo Scientific Phire Plant Direct PCR Master Mix product's instruction

(http://assets.fishersci.com/TFS-Assets/LSG/manuals/F== 160\_QR\_TS\_5.PDF). Purification of amplified product checked by 1.2 % agarose gel with ethidium bromide (Buckman *et al.* 2013 and Kumar *et al.* 2014). The products that were amplified were sent to the Rajiv Gandhi Centre for Biotechnology (RGCB) in Trivandrum, Kerala, for sequencing (India).

The National Center for Biotechnology Information (NCBI) database's Basic Local Alignment Search Tool (BLAST) was used to run a similarity search to determine the identity of the genomic DNA. The sequences were then compared with an existing database to confirm the species. Furthermore, additional sequences were obtained from GenBank and a multiple alignment was done using ClustalX version 2.1 (Thompson et al 1997). In addition, extra sequences were dragged from GenBank, and ClustalX version 2.1 (Tamura et al 2021) was used to perform a multiple alignment. Using the Jukes-Cantor distance model and the Maximum Likelihood approach with a bootstrap test of 1000 replications, the phylogenetic tree was studied using MEGA version 11.0. (Jukes and Contar 1969).

#### **RESULTS AND DISCUSSION**

Morphological Confirmation Phylum: Arthropoda Class: Insecta Order: Thysanoptera Family: Thripidae Sub family: Thripinae Genus: Thrips

**Species:** *T. palmi* (Karny) 1925: The following characteristics distinguish *T. palmi* from other known species of the genus Thrips during the morphological key characterisation process, which was carried out using created permanent slides of thrips (Fig. 1a-g). Head is

**Table 1.** Location of orchards covered during survey 2023

broader than long; Ocellar seta III faces lateral on anterior ocellus (Fig. 1g); antenna 7- segmented, I-II yellowish, III – V basally yellowish and distally brown, VI-VII brown (Fig 1b); Pronotum with 2 pair of long posteroangular setae (Fig 1d); Metanotum with two companiform sensilla (Fig 1e); Forewing yellow with three (sometimes two or four) setae on distal half of first anterior vein, Second vein with row of 15 setae approximately (Fig 1f). Abdomen tergite II with four lateral setae and microtrichia lacking on lateral thirds of tergites IV–VI; Tergite V–VII with paired ctenidia laterally; Tergite VII Posteromarginal comb complete, microtrichia long and slender (Fig 1c); Discal setae absent.

Phylum: Arthropoda

Class: Insecta

Order: Thysanoptera

Family: Phlaeothripidae

Sub Family: PhlaeothripinaGenus: Haplothrips

Species: H. tenuipennis Bagnall, 1918: 210: Haplothrips ceylonicus var. mangiferae Priesner, 1933: 359 syn.n. All members of the genus Haplothrips have four sense cones (rarely three) on segment IV of the antenna and one or two on segment III of the antenna. In Haplothrips tenuipennis, S1 setae in tergite not more than 0.75mm as long as tube. Antenna; the antennal segment III has a smaller sense cone and thinner. Sub basal S3 seta of forewing is acute (Mound 2019), which were confirmed the attack of mango inflorescence by T. palmi and Haplothrips tenuipennis. Zambrano et al (2021) first reported T. palmi on cotton in Ecuador by using the taxonomic key characters. Krishnamoorthy and Ganga Visalakshi (2009) reported T. palmi on flowers in his trail field to manage mango hoppers and thrips using entomopathogens. Additionally, H. tenuipennis was found in the same host from Andhra Pradesh, India by Ramasubbarao and Thammiraju 1994; Kannan and Rao 2006a, b). Earlier, S. rubrocintus has been

State	District	Panchayat	Locality	Latitude, Longitude	Host	Specimen	Date of collection	Name of collector	Material deposited
Kerala	Palakkad	Muthalamada	Adavumaram	10.5992614 76.7621706	Mango	<i>T. palmi</i> and <i>H. tenuipennis</i> (Adult thrips)	04/12/2023	Syed Mohamed Ibrahim	Insect museum, College of agriculture, Vellayani
			Vellaramkadavu	10.5740950 76.7725330			04/12/2023		
			Vellaramkadavu	10.5772010 76.7762450			08/12/2023		
			Anna nagar	10.5792830 76.7786070			08/12/2023		
			Chemmanapathy	10.57927 76.778582			13/12/2023		

observed in Kerala on inflorescence of mango (Ananathakrishnan and Muraleedharan 1974). But, till now no other thrips species was reported on mango in Kerala, India. Verghese et al (1988) and Krishnamoorthy and Visalakshi (2009) had previously reported on *T. palmi*'s attack on mango inflorescence in Tamil Nadu. On the other hand, Kerala is the first place where reports of these two species attacking mango inflorescences have been confirmed.

#### **Molecular Characterisation**

In the study, the NCBI database was used to BLAST sequences generated over incomplete mtCOI areas in order to obtain the accession numbers (OP963194 and OP957024) for *T. palmi* and *H. tenuipennis* from Muthalamada. With an E-value of 0 and a likelihood of 92.79% and 92.89% match, the Blast search using the NCBI BLAST connected the individuals to the *T. palmi* and *H. tenuipennis* sample from China and Pakistan, accession numbers MF6866687 and KP871477. The sum of branch



Fig. 1(a-g). Thrips palmi. a, Female; b, antenna; c, abdominal tergite; d, pronotum; f, meso and metanotum; g, forewing; e, ocelle setae

length of optimal maximum likelihood tree = 0.015689 and 0.015939 is displayed in Figure 2 & 3. Eight nucleotide sequences for T. palmi and H. tenuipennis were analyzed. First, second, third, and noncoding codon locations were covered. Every ambiguous location was removed for every sequence pair (pairwise deletion option). The completed dataset contained 478 and 483 positions in total. Evolutionary analyses of maximum likelihood tree and T. palmi mtCOI gene sequence (collected from GenBank) revealed that our samples were closest to the T. Palmi of China, Bangladesh and Indonesia formed as separate cluster. mtCOI sequence of T. major was taken as outgroup (Fig 2). Similarly, the maximum likelihood tree analyses for H. tenuipennis displayed that the sample from Kerala was close near to the Bangladesh which occupied in one clade and the sample from India and Pakistan of GenBank occupied on different clade. Haplothrips ganglbaueri as taken as outgroup (Fig. 3).

T. palmi morphologically resembles both T. alatus Bhatti and T. fabus Schrank. The location of the interocellar setae, which in T. palmi originate from outside the ocellar triangle but in T. fabus, originate from inside the ocellar triangle posterior to the anterior ocellus, supports differentiation (Mound 2009). Unlike T. palmi, where the metanotal sculpture is typically convergent on the posterior margin, T. alatus exhibits striate metanotal sculpture (Mound 2009). Likewise, female paratypes of H. ceylonicus var. mangiferae and syntypes of this species from northern India have been examined and compared. Mound (1968) established the first synonymy between these two species, whose females cannot be distinguished from one another. H. tenuipennis resembles members of the H. anceps group of species in general because it has dark pronotal major setae in common with them, but it differs from them in that it has a small, slender sense cone on the inner margin of antennal segment III that



Fig. 2. Maximum likelihood tree of *T. palmi* samples from Muthalamada, Kerala, and other locations retrieved from GenBank



Fig. 3. Maximum likelihood tree of *H. tenuipennis* samples from Muthalamada, Kerala and others extracted from GenBank

neither of the other two species have developed. In order to prevent misunderstandings, specimens that had been recognized morphologically were next subjected to molecular identification using mtCO1 gene PCR amplification to confirm that they were as *T. palmi* and *H. tenuipennis* not any other thrips species (Riley et al 2011).

#### CONCLUSION

The specimens collected on mangos were confirmed to be *T. palmi* and *H. tenuipennis* by both morphological and molecular evidence. This is the first report of *T. palmi* and *H. tenuipennis* on mango inflorescence at Muthalamada panchayat in Palakad, Kerala, India. The southern states of India are ideally suited for thrips' occurrences on mango, Since *T. palmi* and *H. tenuipennis* are spreading their host range, their occurrences in various regions of India require rigorous monitoring and IPM measures.

#### ACKNOWLEDGEMENTS

The authors are grateful to Dr. Rachana, R.R Scientist (Division of Germplasm Collection and Characterisation), ICAR-NBAIR, Bangaluru, Dr. Kumoud tyagi, Zoological Survey of India, Kolkata for morphological identification of pest and KAU for providing financial support to conduct study.

#### REFERENCES

- Aliakbarpour H and Salmah Md Rawi C 2010. Diurnal activity of four species of thrips (Thysanoptera: Thripidae) and efficiencies of three non-destructive sampling techniques for thrips in mango inflorescences. *Journal of economic entomology* **103**(3): 631-640.
- Ananthakrishnan T N and Muraleedharan N 1974. On the incidence and effects of infestation of *Selenothrips rubrocintus* (Giard) (Thysanoptera: Heliothripinae) on the free amino-acids of some susceptible host plant. *Current science* **43**: 216-218.

- Asokan RNK, Kumar KV and Ranganath HR 2007. Molecular differences in the mitochondrial cytochrome oxidase I (mtCOI) gene and development of a species-specific marker for onion thrips, *Thrips tabaci* Lindeman, and melon thrips, *T. palmi* Karny (Thysanoptera: Thripidae), vectors of tospoviruses (Bunyaviridae). *Bulletein of Entomological Research* 97: 461-470.
- Buckman RS, Mound LA and Whiting MF 2013. Phylogeny of thrips (Insecta: Thysanoptera) based on five molecular loci. *Systematic Entomology* **38**: 123-133.
- Farris RE, Ruiz-Arce R, Ciomperlik M, Vasquez JD and DeLeon R 2010. Development of a ribosomal DNA ITS2 marker for the identification of the thrips, *Scirtothrips dorsalis*. *Journal of Insect Science* **10**: 26.
- GOK [Government of Kerala]. 2020. Agricultural Statistics 2019-2020 [on-line]. Available https://www.ecostat.kerala.gov.in/ storage/publications/239.pdf
- Hulagappa T, Baradevanal G, Surpur S, Raghavendra D, Doddachowdappa S, Shashank PR, Mallaiah KK and Bedar J. 2022. Diagnosis and potential invasion risk of *Thrips parvispinus* under current and future climate change scenarios. *Peer Journal* **10**: p.e13868.
- Jukes TH and Cantor CR 1969. Evolution of protein molecules, pp 21-132. In: Munro HN (eds). *Mammalian Protein Metabolism*. Academic Press, New York.
- Krishnamoorthy A and Visalakshi PG 2012. Record of thrips on mango. *Journal of Horticultural Sciences* **7**(1): 110-111.
- Kumar V, Seal DR, Kakkar G, Mckenzie CL and Osborne LS 2012. New tropical fruit hosts of *Scirtothrips dorsalis* (Thysanoptera: Thripidae) and its relative abundance on them in South Florida. *Florida Entomology* **95**: 205-207.
- Kumar V, Tyagi K, Ghosh B and Singha D 2014. A new species of Taeniothrips (Thysanoptera: Thripidae) from India. Zootaxa 3884(2): 197-200.
- Mound LA 1968. A review of R.S. Bagnall's Thysanoptera collections. Bulletin of the British Museum (Natural History). *Entomology Supplement* **11**: 1-181.
- Mound LA 2005. The *Thrips orientalis* group from South-east Asia and Australia: some species identities and relationships (Thysanoptera: Thripidae). *Australian Journal of Entomology* **44**: 420-424.
- Mound LA 2019. Identification of *Haplothrips* species from Malesia (Thysanoptera, Phlaeothripinae). *Zootaxa* **4623**(1):41-50.
- Mound LA and Azidah AA 2009. Species of the genus Thrips (Thysanoptera) from Peninsular Malaysia, with a checklist of recorded Thripidae. *Zootaxa* **2023**: 55-68.

- Ramasubbarao V and Thammiraju N B. 1994. New record of blossom thrips *Megalurothrips distalis* on mango (*Mangifera indica*). The Indian Journal of Agricultural Sciences **64**(6): 417-418.
- Riley D, Shimat VJ, Srinivasan R and Diffie S 2011. Thrips vectors of tospoviruses. *Journal Integrated Pest Management* 1:1-10.
- Tamura K, Stecher G and Kumar S2021. MEGA 11: Molecular Evolutionary Genetics Analysis Version 11. *Molecular Biology Evolution* 38(7): 3022-3027.
- Tandon P L and Verghese A 1987. New insect pests of certain fruit crops. *Indian Journal of Horticulture* **44**(1&2): 120-121.
- Thompson J D, Gibson T J, Plewniak F, Jeanmougin F and Higgins D G 1997. The ClustalX windows interface: flexible strategies for

Received 06 June, 2024; Accepted 01 October, 2024

multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Research* **24**: 4876-4882.

- Verghese A, Tandon PL and Rao GP 1988. Ecological studies relevant to the management of *Thrips palmi* Karny on mango in India. *International Journal of Pest Management* **34**(1): 55-58.
- Vrijenhoek R 1994. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology Biotechnology* 3: 294-299.
- Zambrano ND, Arteaga W, Velasquez J and Chirinos DT 2021. Side effects of lambda cyhalothrin and thiamethoxam on insect pests and natural enemies associated with cotton. *Sarhad Journal of Agriculture* **37**(4): 1098-1106.



Manuscript Number: 4354 NAAS Rating: 5.38

# Life Table of Pod Borer, *Helicoverpa armigera* (Hubner) on Pigeonpea

### B.C. Patel, Bindu Panickar and M.R. Dabhi

Department of Entomology, C.P. College of Agriculture S.D. Agricultural University, Sardarkrushinagar-385 506, India E-mail: pbipin1@sdau.edu.in

**Abstract:** Life-table of *H. armigera* on pigeonpea was studied under laboratory condition. The output showed that out of total 100 eggs only the 89 eggs, 77 larvae and 70 pupae survived and finally emerged in to adults. The maximum durations of egg, larva, and pupa was 6, 20 and 8 days, respectively with pre-oviposition period varied from  $33^{rd}$  to  $35^{th}$  days of pivotal age. Female started laying eggs after  $35^{th}$  days and ceased after  $47^{th}$  days with lx (survival of female) values of 0.70 and 0.31 respectively. Females produced the maximum progenies (mx = 48.09) on the  $38^{th}$  day of pivotal age. The net reproductive rate (Ro) was 142.07 times multiplication of population per generation. Adults contributed only 0.76 percent of the population of stable age whereas eggs, larvae, and pupae contributed 59.31, 37.68 and 2.26 per cent, respectively. The expectancy of further life was 4.76 days at the time of adult emergence which showed that life expectancy of *H. armigera* declined with the advancement of development.

Keywords: H. armigera, Adults, Female, Eggs, Larvae, Pupae, Progenies, Legume, Life table

Pigeonpea [Cajanus cajan (Linnaeus) Millspaugh] is an important grain legume crop of the tropics and subtropics. Globally, pigeonpea is grown in an area of 6.03 million hectares with a production of 5.33 million tonnes and productivity of 883.40 kg/ha (FAO 2022). In India, it is grown on 5.05 million hectares with an annual production and productivity of 4.34 million tonnes and 859 kg/ha, respectively (Anonymous 2022). Among several insect pests infesting pigeonpea the lepidopteran pest viz., pod borer, Helicoverpa armigera (Hubner) is most serious. In India, the pod damage caused by Melanogromyza obtusa, H. armigera and Maruca vitrata was reported as 34.4 to 49.9, 9.4 to 18.1 and 5.7 to 12.4 per cent, respectively (Keval et al 2017). The maximum damage was recorded due to pod borer, H. armigera (34.90 %) followed by pod fly, M. obtusa spotted pod borer, M. vitrata and tur plume moth, Exelastis atomosa up to 23.64, 14.09 and 5.60 per cent, respectively (Muchhadiya et al 2024). Considering the above facts, the present investigation was carried out.

#### MATERIAL AND METHODS

To construct the life-table, the culture of *H. armigera* was maintained on pigeonpea tender plant for two consecutive generations at room temperature during October to March, 2021-22. The adults obtained from the culture were used for the further study. The freshly emerged male and female moth were kept for egg laying in wooden cages. The sides of the cage were covered with muslin cloth and the tender

pigeonpea branches were provided for egg laying. In order to construct life table, freshly laid 100 eggs were collected from the cage with the help of wet camel hair brush and placed in ten petri plate (10.0 cm diameter × 1.5 cm height) in batches of 10 (ten) were observed closely for recording the hatching percentage. After hatching, the first instar larvae were transferred individually into plastic vials. Fresh food was provided daily in morning. Vials were kept clean by removing frasses. Took daily observations on larval development, formation of pupae, emergence of adult and fecundity were recorded. Age-specific mortality in several developmental stages such as egg, larva, pupa and adult were also documented. To establish age-specific fertility, the total number of adults that emerged on the same day were placed in a cage for oviposition. Pigeonpea branches with leaves, buds, and pods were placed in conical flasks with fresh water and kept in cages until oviposition. The twigs were replaced every day and the number of eggs laid on the twigs, buds, pods and muslin material inside the cage were noted. Observations on fecundity were recorded until the female died. Assuming a 1:1 sex ratio, the number of eggs collected per female was divided by two to get the number of female births (mx). The methodology for life table as proposed by Howe (1953) followed by Atwal and Bains (1974), Dabhi et al (2009a), Dabhi et al (2009b), Patel et al (2016), Singh et al (2022) and Chaudhari et al (2023) was adopted. The stable age distribution was determined by observing the population schedule of birth and death rates (mx and lx) when grown in a

- x = Pivotal age in days
- Ix = Survival of female at age 'x'
- mx = Age schedule for female birth at age 'x'

Net reproductive rate (Ro): The values of 'x', 'lx', and 'mx' were calculated using the data provided in the life tables. The sum total of the products 'lxmx' is the net reproductive rate (Ro). The Ro is the rate of multiplication of population in generation measured in terms of females produced per generation. The formula used to compute the number of times a population would multiply per generation is as follows:  $Ro = \Sigma Ixmx$ 

**Mean duration of generation (T):** The approximate value of generation time (T) (the mean age of the mother in a cohort at the birth of female offspring) was calculated by using following formula (Atwal and Bains 1974)

Innate capacity for increase in numbers (rm): At each age interval, the total number of survivors and the average number of females emerged were recorded. The arbitrary value of 'rm (rc)' was determined from these data using the following formula (Loughlin, 1965 and Atwal and Bains 1974)

Where,

e=2.71828

T = Mean generation time

The intrinsic rate increase (rm) was calculated subsequently from the arbitrary 'rm' by taking two trial values; arbitrary selected on either side of it, differing in the second decimal place by establishing the following relationship as suggested by Atwal and Bains (1974).

 $\Sigma e^{7-rmx}$ . Ixmx =  $e^{7}$  = 1097.00

Where,

e=2.71828

The precise generation time (T) was calculated by using the following formula:

**Finite rate of natural increase** ( $\lambda$ )**:** The number of females per female per day *i.e.* finite rate of increase was determined as:

 $\lambda = anti \log e^{m}$ 

From this data the weekly multiplication of the population

was calculated. Hypothetical  $F_2$  females were also be worked out with the formula (Ro)<sup>2</sup>.

**Stable age distribution**: The stable age distribution (per cent distribution of various age groups) of *H. armigera* on pigeonpea was calculated using the knowledge of 'rm' and age-specific mortality of the immature and mature stages. The stable age distribution table was created using the methods proposed by Andrewartha and Birch (1954) and Atwal and Bains (1974). The 'Lx' (life table age distribution) was calculated from the 'lx' table using the following formula:

Per cent distribution of each age group (x) was calculated by multiplying the Lx with  $e^{-m(x+1)}$ . By putting together, the percentage under each stage *viz.*, egg, larval, pupal and adult stages, the expected per cent distribution were worked out.

#### **RESULTS AND DISCUSSION**

The study on the life table, age-specific distribution and life expectancy of *H. armigera* on pigeonpea were carried out at laboratory of Entomology from October to March during 2021-22 at Sardarkrushinagar Dantiwada Agricultural University, Sardarkrushinagar. The 89 per cent egg survived and 70 per cent reached the adult stage (Table 1). The longest durations of egg, larva, and pupa were 6, 20 and 8 days, respectively.

In the past, Deb and Bharpoda (2016) at Anand reported that total 92 eggs survived and 71 per cent reached adults. Basavaraj et al (2018) at Raichur (Karnataka) reported that 77 per cent survived from egg to adult emergence. The maximum duration of egg, larva and pupa was as 6, 28 and 9

 Table 1. Survival of different life stages of H. armigera during development on pigeonpea (Based on 10 eggs)

Egg	Larva	Pupa
	Number survived	
9	7	6
8	7	6
9	8	6
10	8	7
8	7	7
9	7	7
8	7	6
9	8	8
10	9	8
9	9	9
89	77	70

days, respectively. Thus, the findings of the current study are more or less consistent with those of the previous researchers. The pre-oviposition period occurred between the  $33^{rd}$  and  $35^{th}$  days of pivotal age (Table 2). Female began depositing eggs after  $35^{th}$  days and ceased after  $47^{th}$  days with lx of 0.70 and 0.31 respectively. Females produced the most progenies (mx = 48.09) on the  $38^{th}$  day of pivotal age, which declined day by day, thereafter.

The net reproductive rate (Ro) was 142.07 with a mean length of generation (Tc) 38.35 days. The intrinsic rate of natural increase in number (rm) was 0.1292 females per female per day with finite rate of increase ( $\gamma$ ) 1.14 females/female/day and the population was multiplied 2.48 times per week. The hypothetical F2 females were calculated to be 20183.8849 (Table 3).

Basavaraj et al (2018) reported that mean length of generation (T) was 41.40 days. The innate capacity (rm) and finite rate ( $\lambda$ ) of the in number was 0.13 and 1.14 females per female per day, respectively. Fathipour et al (2020) reported



Fig. 1. Survivorship curve (Ix) and age specific fecundity (mx) of *H. armigera* on pigeonpea

Pivotal age in days (x)	Survival of female at different age interval (Ix)	Age schedule for female births (mx)	lxmx	xlxmx
0-33		Immature	stages	
33-35		Pre-ovipositio	on stages	
36	0.70	31.03	21.72	781.96
37	0.69	46.99	32.42	1199.65
38	0.68	48.09	32.70	1242.65
39	0.67	32.38	21.69	846.09
40	0.65	24.31	15.80	632.06
41	0.64	13.03	8.34	341.91
42	0.61	7.12	4.34	182.41
43	0.55	4.31	2.37	101.93
44	0.47	2.94	1.38	60.80
45	0.39	2.31	0.90	40.54
46	0.34	1.17	0.40	18.30
47	0.31	0.00	0.00	0.00
			lxmx = 142.07	xlxmx = 5448.30

Table 2. Life table (Female) and age specific fecundity for *H. armigera*on pigeonpea

 Table 3. Mean length of generation, innate capacity for increase in numbers and finite rate of increase in numbers of *H. armigera*on pigeonpea

Population growth statistics	Formula	Calculated values	
Net reproductive rate	Ro = lxmx	142.07	
Mean length of generation (days)	Tc = xlxmx/Ro	38.3494	
Innate capacity for increase in numbers (Females/female/day)	rm = loge <sup>₨</sup> /Tc	0.1292	
Arbitrary 'rm' (rc)	0.12 and 0.13		
Corrected 'rm' (Females/female/day)	e <sup>7 - rmx. lxmx</sup>	0.1299	
Corrected generation time (days)	T = loge <sup>Ro</sup> /rm	38.1342	
Finite rate of increase in numbers (Females/female/day)	$(\lambda)$ = antilog e <sup>m</sup>	1.1387	
Weekly multiplication of population (times/week)	<b>(λ)</b> <sup>7</sup>	2.4820	
Hypothetical F <sub>2</sub> females	(Ro) <sup>2</sup>	20183.8849	

 Table 4. Stable age specific distribution of *H. armigera*on pigeonpea (rm = 0.1299)

Pivotal age in days 'x'	Lx	e <sup>-m(x+1)</sup>	Lx. e <sup>-m(x+1)</sup>	Percentage distribution
0	1.00	0.8781	0.8781	13.3646
1	1.00	0.7711	0.7711	11.7357
2	1.00	0.6771	0.6771	10.3054
3	1.00	0.5946	0.5946	9.0494
4	1.00	0.5221	0.5221	7.9464
5	0.99	0.4585	0.4539	6.9082
Eggs				59.3097
6	0.89	0.4026	0.3583	5.4534
7	0.88	0.3535	0.3111	4.7350
8	0.82	0.3104	0.2546	3.8744
9	0.81	0.2726	0.2208	3.3607
10	0.81	0.2394	0.1939	2.9511
11	0.81	0.2102	0.1703	2.5914
12	0.80	0 1846	0 1477	2 2475
13	0.80	0.1621	0.1297	1.9736
14	0 79	0 1423	0 1124	1 7114
15	0.78	0 1250	0.0975	1 4838
16	0.78	0.1098	0.0856	1 3029
17	0.77	0.0964	0.0742	1 1295
18	0.77	0.0846	0.0652	0 9918
10	0.77	0.0743	0.0002	0.8709
20	0.77	0.0653	0.0502	0.7648
20	0.77	0.0000	0.0302	0.7040
21	0.77	0.0503	0.0387	0.5897
22	0.77	0.0303	0.0340	0.5037
20	0.77	0.0442	0.0340	0.5170
	0.77	0.0500	0.0299	37 6750
25	0.77	0.03/1	0.0262	0 3003
25	0.77	0.0341	0.0202	0.3595
20	0.77	0.0299	0.0230	0.3000
21	0.77	0.0203	0.0202	0.3079
20	0.77	0.0201	0.0176	0.2374
20	0.77	0.0203	0.0130	0.2085
31	0.77	0.0176	0.0137	0.1831
30	0.77	0.0130	0.0120	0.1608
32	0.77	0.0137	0.0100	0.1008
Punao	0.77	0.0120	0.0035	2 2501
21	0.75	0.0106	0.0070	0 1207
25	0.75	0.0100	0.0079	0.1207
26	0.70	0.0093	0.0100	0.1522
30	0.70	0.0082	0.0037	0.0869
37 20	0.69	0.0072	0.0049	0.0752
30 20	0.00	0.0065	0.0043	0.0560
39 40	0.07	0.0055	0.0037	0.0503
40	0.05	0.0048	0.0032	0.0480
41	0.64	0.0043	0.0027	0.0415
42	0.61	0.0037	0.0023	0.0347
43	0.55	0.0033	0.0018	0.0275
44	0.47	0.0029	0.0014	0.0206
45	0.39	0.0025	0.0010	0.0150
46	0.34	0.0022	0.0008	0.0115
Adult				0.7553

Pivotal age (Days)	Number surviving to the beginning of age interval	Number dying during 'x'	Mortality rate per hundred alive at beginning of age interval [dx.100/lx]	Alive between age 'x' and 'x + 1' [lx + (lx+1)]/2	No. of the individual's life days beyond 'x'	Expectation of further life (Tx / lx) × 2
(x)	(lx)	(dx)	(100 qx)	(Lx)	(Tx)	(ex)
0-5	100	11	11.00	100.50	689.00	13.78
5-10	89	8	8.99	89.50	610.50	13.72
10-15	81	3	3.70	81.50	537.00	13.26
15-20	78	1	1.28	78.50	461.50	11.83
20-25	77	0	0.00	77.50	385.00	10.00
25-30	77	0	0.00	77.50	307.50	7.99
30-35	77	7	9.09	77.50	230.00	5.97
35-40	70	5	7.14	70.5	166.50	4.76
40-45	65	26	40.00	65.5	106.00	3.26
45-50	39	8	20.51	39.5	92.50	4.74
50-55	31	0	0.00	0.00	0.00	0.00

Table 5. Life table for computing life expectancy of H. armigeraon pigeonpea

that the net reproductive rate (Ro) was 147.40 female offspring. The mean length of generation (T) was 37.90 days. The innate capacity (rm) and finite rate ( $\lambda$ ) of the in number was 0.126 and 1.131 females per female per day, respectively. Maity et al (2020) reported that the net reproductive rate (Ro) was 133.83female offspring. The mean length of generation (T) was 43.7days. The innate capacity (rm) and finite rate ( $\lambda$ ) of the in number was 0.112 and 1.118 females per female per day, respectively. In current findings are similar trend was observed. The impact of each developmental stage and the stable age distribution indicated that adults showed just 0.76 per cent of the population of stable age whereas eggs, larvae and pupae contributed 59.31, 37.68 and 2.26 per cent, respectively (Table 4).

The life expectancy demonstrated that *H. armigera* life expectancy decreased significantly as development progressed. The life expectancy of newly placed eggs was 13.78 days (Table 5). The expectancy of further life was 4.76 days at the time of adult emergence.

#### CONCLUSION

Females produced the maximum progenies (mx = 48.09) on the 38<sup>th</sup> day of pivotal age. The net reproductive rate (Ro) was 142.07 times multiplication of population per generation. Adults contributed only 0.76 per cent of the population of stable age, whereas, eggs, larvae, and pupae contributed 59.31, 37.68 and 2.26 per cent, respectively. The expectancy of further life was 4.76 days at the time of adult emergence which showed that life expectancy of *H. armigera* was declined with the advancement of development. It is

important to increase the egg mortality and minimize the population of *H. armigera*.

#### REFERENCES

- Andrewartha HC and Birch CC 1954. *The distribution and abundance of ani*mals. University of Chicago Press p. 782.
- Anonymous 2022. *Statistics at a Glance 2022* Ministry of Agriculture and Farmers Welfare, Department of Agriculture and Farmers Welfare, Economics and Statistics Division, Government of India, Agricultural pp. 44-45.
- Atwal AS and Bains SS 1974. *Applied Animal Ecology*, Kalyani Publishers, Ludhiana pp. 11-35.
- Basavaraj K, Naik MI, Jagadish KS and Shadakshari YG 2018. Studies on age specific fecundity life tables for *Helicoverpa armigera* Hub. on sunflower (*Helianthus annuus* L.). *Journal of Entomology and Zoology Studies* **6**(2): 1364-1368
- Chaudhari P, Dabhi MR and Patel HC 2023. Life table of *Chrysoperla zastrowi sillemi* (Esben- Petersen) on coriander aphid, *Hyadaphis coriandri* (Das). *The Pharma Innovation Journal* **12**(10): 315-318
- Dabhi MR, Mehta DM and Patel CC 2009a. Life table of diamondback moth, *Plutella xylostella* L. on Cress (*Lepidium* sativum L.). International Journal of Agriculture Environment and Biotechnology 2(1): 80-82.
- Dabhi MR, Mehta DM, Patel CC and Korat DM 2009b. Life table of diamondback moth, *Plutella xylostella* (Linnaeus) on cabbage (*Brassica oleraceae* var. *capitata* L.). *Karnataka Journal of Agricultural Sciences* 22(2): 319-321.
- Deb S and Bharpoda TM 2016. Life-table parameters of fruit borer, *Helicoverpa armigera* (Hubner) hardwick in tomato, *Lycopersicon esculentum* Mill. *The Bioscan* **11**(1):09-14.
- FAOSTAT 2022. FAOSTAT Agriculture data. Available at http://faostat.fao.org/> accessed on 7<sup>th</sup> January, 2024.
- Fathipour Y, Baghery F, Bagheri A and Naseri B. 2020. Development, reproduction and life table parameters of Helicoverpa armigera (Lepidoptera: Noctuidae) on five main host plants. *Journal of Crop Protection* **9**(4): 551-561.
- Howe RW 1953. The rapid determination of intrinsic rate of increase of an insect population. *Annals of Applied Biology* **40**: 134-135.
- Keval R, Kumar R, Chakravarty S and Mishra VK 2017. Extent of damage caused by major insect pests on long duration pigeonpea (*Cajanus cajan* (L.) Millsp.) under natural conditions. *Plant Archives* 17(1): 643-646.
- Loughlin R 1965. Capacity for increase: a useful population statistics. *Journal of Animal Ecology* **34**: 77-91.
- Maity C, Mondal P and Mondal L 2020. Studies on age specific & female fertility life tables of *Helicoverpa armigera* under controlled condition. *Journal of Entomology and Zoology Studies* **8**(2): 585-591.

Received 12 August, 2024; Accepted 30 September, 2024

- Muchhadiya DV, Patel JJ, Patel DR and Patel RB 2024. Estimation of yield losses caused by insect pests on pigeonpea (*Cajanus cajan* (L.) *Millsp.*). International Journal of Plant and Soil Science **36**(3): 410-414.
- Patel HC, Borad PK and Dabhi MR 2016. Life fecundity table of *Maruca vitrata* on green gram. *Indian Journal of Plant Protection* **44**(1): 40-43.
- Singh NA, Dabhi MR and Mohapatra AR 2022. Life table of ladybird beetle, *Cheilomenes sexmaculata* (Fabricius) on cotton aphid. *The Pharma Innovation Journal* **11**(6): 2872-2875.



# Phenology, Productivity and Profitability with Phosphate and Zinc Solubilizing Microbes in Lentil (*Lens culinaris* L.) under *tarai* region of Uttarakhand

Monica Yaying, Anil Shukla, Supriya<sup>1\*</sup>, Sudarshan S.<sup>2</sup>, Shobhana Singh and Gunashekhar H.<sup>3</sup> and Chandra Bhushan<sup>4</sup>

Department of Agronomy, G.B. Pant University of Agriculture & Technology, Pantnagar-263 153, India <sup>1</sup>Agronomy Section, ICAR-National Dairy Research Institute, Karnal-132 001, India <sup>2</sup>Division of Agronomy, ICAR-Indian Agricultural Research Institute, New Delhi-110 012, India <sup>3</sup>Dairy Extension Section, ICAR-National Dairy Research Institute, Southern Regional Station, Bengaluru-560 030, India <sup>4</sup>Institute of Agricultural Sciences, BHU, Varanasi- 221 005, India \*E-mail: supriya.ndri5@gmail.com

**Abstract:** The field experiment was conducted at G. B. Pant University of Agriculture and Technology, Pantnagar during *rabi* 2020 on phosphorus and zinc efficacy with phosphate and zinc solubilizing microbes in lentil (*Lens culinarisL.*) in *tarai* region. Twelve different doses of phosphorus and zinc with phosphate and zinc solubilizing microbes in combinations were evaluated. Growth parameters *viz.* plant height, number of branches per plant, number of compound leaves per plant and dry matter accumulation were highest with application of 50% RDP (@ 24 kg ha<sup>-1</sup> +12.5 kg ha<sup>-1</sup> ZnSO<sub>4</sub> + Biophos (@ 5 ml kg<sup>-1</sup> seed + Biozinc (@ 5 ml kg<sup>-1</sup> seed (40.5 cm, 14 no. plant<sup>-1</sup>, 207 no. plant<sup>-1</sup>, 8.2 g plant<sup>-1</sup>, respectively) with respect to control (32.9 cm, 10.5 no. plant<sup>-1</sup>, 136 no. plant<sup>-1</sup>, 4.1 g plant<sup>-1</sup>, respectively). Grain yield, harvest index, net returns and B: C ratio were also highest with the application of 50% RDP (@ 24 kg ha<sup>-1</sup> +12.5 kg ha<sup>-1</sup> ZnSO<sub>4</sub> + Biophos (@ 5 ml kg<sup>-1</sup> seed (1735 kg ha<sup>-1</sup>, 0.53, 58460 € ha<sup>-1</sup>, 2.51, respectively) with respect to control (1318 kg ha<sup>-1</sup>, 0.34, 40231 € ha<sup>-1</sup>, 1.82, respectively).

Keywords: Biophos, Biozinc, Harvest index, Nitrogen fixation, Rhizobium

Lentil (Lens culinaris L.) belongs to the family Fabaceae and about 5% of the total area under pulses is lentil of which Asia accounts for 65% in terms of production. In India, lentil is a significant pulse crop, grown on approximately 1.42 million hectares, yielding around 1.28 million tonnes (Sah et al 2022). The lentil crop accounts for about 4.9% of the total pulse cultivation area and contributes 5.03% to the country's overall pulse production (2021-22). Phosphorus and zinc are essential elements for successful plant growth, particularly in pulse production (Siddigui et al 2015). Lentils use Rhizobium bacteria to fix atmospheric nitrogen, reducing the need for nitrogenous fertilizers to about 25-30 kg N ha-1 (Singh and Singh 2016). Proper phosphorus application is essential for improving lentil yield and soil quality, although excess application can lead to fixation and unavailability in acidic or alkaline soils. Introducing phosphorus-solubilizing microbes (PSB) into the rhizosphere enhances phosphorus availability to plants through the release of organic acids and enzymes, potentially increasing crop yields by 10-30% and supplementing phosphorus needs. Zinc-solubilizing microorganisms use various mechanisms, such as acidification and chelation, to make zinc more available in the soil. Inoculating plants with beneficial bacteria like Pseudomonas, Rhizobium strains, Bacillus and Azospirillum has shown improvements in growth, phosphorus and zinc content (Sindhu et al 2019, Chetan Babu et al 2023). Overall, these micronutrients and microbial interventions are crucial for optimizing crop health and yield in pulse production. The information available on the effect of phosphorus and zinc solubilizing microbes in the *Tarai* region of Uttarakhand is sparse and inadequate. With respect to this objective, the present study was conducted on phenology, productivity and profitability with phosphate and zinc solubilizing microbes in lentil.

### MATERIAL AND METHODS

**Experimental site:** The field experiment was carried out during the *rabi* season of 2020 at N.E. Borlaug Crop Research Centre of G. B. Pant University of Agriculture and Technology. The research center lies 30 km Southern end of foot hills of Shivalik Range of Himalayas at 29°N latitude and 79.3°E longitude and at an altitude of 243.83 m above the mean sea level in the *tarai* region of Uttarakhand.

**Experiment details:** The experiment was conducted with twelve treatments and three replications in randomized block design using lentil variety Pant L-8 with row to row spacing of 23cmand experimental area was about 812.16m<sup>2</sup> with gross plot size of 13.8m<sup>2</sup> (2.76 m x 5 m) each. The nutrients like N

and K were given to all treatments alike. But P, Zn and their solubilizing organisms were varied in different treatments. The other agronomic practices were followed as per the standard package of practices of lentil cultivation.

**Growth and development parameters:** At 30, 60, 90 days after sowing (DAS) and at harvest, the growth parameters were recorded. Height was measured from ground surface to the base of the apical leaf of five tagged plants from each plot. Total branch of branches of the five tagged plants were counted. The number of leaves form the five tagged plants were counted. Nodules collected from the five roots from of plants were dried in the oven at 70°C till the constant weight. Number of nodules were cut near the surface and dried in the hot air oven at 70°C till the constant weight.

**Yield parameters:** After threshing, the grain yield from each net plot was weighed and then multiplied with suitable conversion factor to get yield (kg ha<sup>-1</sup>). Straw yield was calculated by deducting the grain yield from the biological yield. Total produce of each net plot (excluding the root biomass) was dried in the sun in the field after harvest and weighed. Yield ha<sup>-1</sup> was computed by multiplying this with suitable conversion factor.

Harvest index = (grain yield /biological yield) x100 **Grain:** Straw ratio was calculated by dividing the grain yield by straw yield.

**Economic evaluation:** Selling price of grain and straw were multiplied to the grain and straw yield to obtain the gross returns. The cost was deducted from the gross returns to obtain the net returns. The net return was divided by the cost of cultivation to obtain benefit cost (B: C) ratio.

**Statistical analysis:** The experimental data were analyzed using OPSTAT for randomized block design which is programmed by HAU, Hisar, Haryana.

# RESULTS AND DISCUSSION

# Growth and Development

**Plant height (cm):** Plant height increased up to crop maturity (Table 1). The rate of increase of plant height for 30-60 DAS, 60-90 DAS and 90 DAS to maturity stage was 0.19, 0.72 and 0.31 cm day<sup>-1</sup>. Maximum increase was observed between 60-90 DAS stage. The  $T_{10}$  produced tallest plants at 30, 60, 90 and maturity *i.e.* 6.4, 12.2, 32.9 and 40.5 cm, respectivelyand was significantly superior over the other treatments at 30, while at 60 DAS and 90 DAS,  $T_{10}$ ,  $T_{11}$  and  $T_{12}$  were statistically on a par with each other, and significantly superior to other

Treatments		Plant he	ight (cr	n)	No. of branches plant <sup>-1</sup>				No. of compound leaves plant <sup>1</sup>		
	30 DAS	60 DAS	90 DAS	Maturity	30 DAS	60 DAS	90 DAS	Maturity	30 DAS	60 DAS	90 DAS
$T_1$ : Absolute Control (without P and Zn)	4.7	9.1	25.4	32.9	2.7	4.9	9.2	10.5	8.7	37	136
$T_2$ : RDP (Control) @ 48 kg ha <sup>-1</sup>	5.2	10.9	28.6	37.9	2.9	6.3	11.3	12.4	9.2	43	146
$T_3$ : Soil application of ZnSO <sub>4</sub> @ 25 kg ha <sup>-1</sup>	5.1	10.5	28.0	37.3	2.9	5.6	10.4	11.7	9.0	40	142
T₄: Biophos @ 5ml kg <sup>-1</sup> seed	5.0	10.4	27.8	37.0	2.8	5.5	10.1	11.1	9.0	39	142
$T_s$ : Biozinc @ 5ml kg <sup>-1</sup> seed	4.9	9.8	26.5	35.1	2.8	5.3	9.6	10.8	8.8	37	137
$T_{\scriptscriptstyle 6}\!\!:\!BiophosandBiozinc\textcircled{0}{0}5mleachkg^{\!-\!1}seed$	5.2	10.7	28.5	37.6	2.9	5.8	10.8	11.8	9.1	42	145
T <sub>7</sub> :50% RDP@ 24 kg ha <sup>-1</sup> + Biophos@ 5 ml kg <sup>-1</sup> seed	5.2	11.1	29.1	38.1	3.0	6.4	11.6	12.4	9.8	43	158
$T_8:12.5 \text{ kg ha}^{-1} \text{ZnSO}_4 + \text{Biozinc} @ 5 \text{ ml kg}^{-1} \text{ seed}$	5.0	10.1	27.7	36.4	2.8	5.5	9.6	11.1	8.9	39	139
$T_g:50\%$ RDP@ 24 kg ha <sup>-1</sup> + Biophos @ 5ml kg <sup>-1</sup> seed + Biozinc @ 5 ml kg <sup>-1</sup> seed	5.3	11.1	31.2	38.1	3.0	6.5	11.8	12.6	10.1	43	179
$T_{10}{:}50\%$ RDP@ 24 kg ha $^{1}$ + 12.5 kg ha $^{1}$ ZnSO4 + Biophos @ 5 ml seed $^{1}$ + Biozinc @ 5 ml kg $^{1}$ seed	6.4	12.2	33.9	40.5	3.4	7.0	13.6	14.0	13.4	47	207
T <sub>11</sub> :RDP@ 48 kg ha <sup>-1</sup> + Nutrient Mobilizer (LNm 43a) @ 20 g kg <sup>-1</sup> seed	5.7	11.3	32.5	38.4	3.1	6.6	12.2	13.7	10.9	43	187
$T_{12}50\%$ RDP@ 24 kg ha $^1$ + Soil application of 12.5 kg ha $^1$ ZnSO4 + Nutrient Mobilizer @ 20g kg $^1$ seed	5.8	12.0	32.9	39.2	3.1	6.7	13.0	14.0	11.3	44	203
CD (p=0.05)	0.5	1.0	2.7	3.4	0.3	0.9	2.7	3.0	1.4	4	26

 Table 1. Plant height, number of branches and compound leaves at different stages of crop growth as influenced by phosphorous and zinc treatments

treatments (Table 1). At maturity too,  $T_{10}$  produced the tallest plants being significantly superior over that of the other nutrient formulations except  $T_{12}$ ,  $T_{11}$ ,  $T_9$ ,  $T_7$ ,  $T_6$ ,  $T_2$  and  $T_3$  which produced statistically similar plants in terms of the plant height. This could be due to synergistic effect of various sources of nutrients known to have beneficial effects in lenti l(Singh et al 2017). Ganie et al (2009) in garden pea and Kant et al (2016) in blackgram observed same trend. Singh et al (2013) also reported variations in plant height of lentil crop due to different doses of phosphorus and zinc.

**Number of branches:** The number of branches increased steadily until maturity, with the growth rate slowing near maturity (Table 1). The highest growth rate occurred between 60 and 90 days. Treatment  $T_{10}$  consistently produced the most branches at all stages. The increased branch number was linked to higher nitrogen uptake during early vegetative stages, supported by phosphorus, which also enhanced root proliferation and the plant's nitrogen-fixing ability. Similar finding was also reported by Kalayu (2019) and Singh et al (2017). Singh et al (2013) also reported that the maximum number of productive branches plant<sup>-1</sup> obtained in case of zinc (0.04%) and minimum in control (no application of zinc) in lentil. Thus, zinc and phosphorus both applied at optimum dose increased the number of branches by improving root as well as shoot growth.

**Number of compound leaves:** Leaf count per plant increased up to the 90 day stage, with the most significant rise between 60 and 90 days, averaging 5.3 leaves per day, compared to 1.12 leaves per day between 30 and 60 days after sowing (Table 1). Treatment  $T_{10}$  consistently produced significantly more leaves than other treatments, except at 60 and 90 days, where it was comparable to  $T_{12}$ . Nitrogen, being

a key component of chlorophyll, played a crucial role in leaf development, while optimal phosphorus and zinc uptake enhanced apical branching, further increasing leaf numbers. Kasturikrishna et al(2000) observed that the application of  $26.6 \text{ kg P}_2\text{O}_5 \text{ ha}^{-1}$  significantly increased the number of leaves as well as the area of the leaves.

Dry matter accumulation (g plant<sup>-1</sup>): Dry matter accumulation increased with crop age and the rate of increase between 60 to 90 DAS was the highest at about 0.25 g day<sup>-1</sup> (Table 2). The maximum dry matter accumulation at 30, 60 and 90 DAS stages *i.e.*0.08, 0.7 and 8.2 (g plant<sup>-1</sup>), respectively, was associated with T<sub>10</sub>. At 30 DAS and 60 DAS stage,  $T_{11}$  and  $T_{12}$  were statistically on a par with the  $T_{10}$  while at 90 days, T<sub>10</sub> and T<sub>12</sub> were statistically on a par. Pandey et al (2010) also drew similar conclusion in lentil crop that the plants grown in soil supplied with zinc produced maximum dry matter yield over no zinc application. Similarly, Singh et al (2013) also confirmed that total above ground biomass was influenced by application of zinc. Tagore et al (2013) and Meena et al (2015) came to similar conclusion that the dry matter yield plant<sup>-1</sup> increased significantly with inoculation of phosphorus solubilizing microbes

**Days taken to flowering:** The effect on number of days taken to flowering due to various treatments was non-significant yet the  $T_1$  took the least number of days (73days) to flowering while  $T_{10}$  took the more number of days (80 days) (Table 2). Days taken to flowering is initiated later when the vegetative growth is lusher and more prolonged due to higher uptake of nitrogen. The treatments lacking in nutrients yields flower sooner to mitigate the unfavorable conditions.

**Days taken to maturity:** Number of days taken to maturity due to different phosphorus and zinc treatments were non-

Treatment		Dry matter (g plant <sup>-1</sup> )		Number of days taken to			
	30 DAS	60 DAS	90 DAS	Flowering	Maturity		
T <sub>1</sub>	0.041	0.34	4.1	73	131		
T <sub>2</sub>	0.052	0.51	5.7	76	134		
T₃	0.046	0.47	5.1	75	133		
T₄	0.046	0.46	4.3	75	133		
T₅	0.045	0.34	4.1	74	132		
T <sub>6</sub>	0.050	0.51	5.3	75	134		
T,	0.064	0.53	5.9	76	135		
T <sub>s</sub>	0.045	0.43	4.1	75	132		
T <sub>9</sub>	0.064	0.54	6.2	77	135		
T <sub>10</sub>	0.076	0.66	8.2	80	139		
T <sub>11</sub>	0.068	0.58	7.1	78	136		
T <sub>12</sub>	0.072	0.62	7.7	78	137		
CD (p=0.05)	0.012	0.09	1.0	NS	NS		

 Table 2. Dry matter accumulation at different stages of crop growth and days taken to flowering and maturity as influenced by phosphorous and zinc treatments

See Table 1 for details

significant (Table 2). The crop took minimum (131 days) number of days for maturity under  $T_1$  and maximum (138 days) days for maturity under  $T_{10}$ . Higher phosphorus application lengthened the period of crop maturity (Rasheed et al 2010).

**Grain yield (kg ha**<sup>-1</sup>): Grain yield ha<sup>-1</sup> differed significantly due to varying treatments of phosphorus and zinc (Table 3).Treatment  $T_{10}$  produced the maximum grain yield being significantly superior over all the other treatments. Combination of different organic and inorganic and timely availability of the nutrients owing to nutrient solubilizing microbes enhanced the plant growth and significantly increased the grain yieldover control (Singh et al 2017). Hussain and Ahmad (2015) reported similar results on effect of phosphorus and zinc on the grain yield.

**Straw yield (kg ha<sup>-1</sup>):** Straw yield ha<sup>-1</sup> was not influenced significantly due to varying treatments of phosphorus and zinc (Table 3). The highest yield was observed in  $T_{10}$ . Phosphorus aids in cell division and help in the development of new tissue and in energy transformation in plants. Enhanced plant growth due to proper availability of nutrients increased the straw yield too. The zinc helped the plant in chlorophyll production and directly contributed in plant growth. Meena et al (2017) observed that the application of phosphorus with zinc resulted in higher grain and straw yield.

**Biological yield (kg ha**<sup> $^{1}$ </sup>): The highest biological yield was obtained from T<sub>10</sub> and was statistically superior to all the other treatments (Table 3). Application of phosphorus and zinc to plants increased grain and straw yield and thus, the biological yield. The applied nutrients improved the various parameters of growth and development like plant height, number of

branches plant<sup>-1</sup>, number of compound leaves plant<sup>-1</sup>, dry matter accumulation plant<sup>-1</sup>, the cumulative effect of which ultimately resulted in higher yields of both straw and biological parts. Singh et al (2008) found that the biological yield increased significantly up to 50 kg  $P_2O_5$  ha<sup>-1</sup>. Different organic and inorganic nutrients might have helped in providing nutrients to the plants and ultimately improving the productivity of lentil (Singh et al 2017).

**Grain straw ratio:** The variations in grain to straw ratio owing to different treatments was non-significant (Table 3.) The highest ratio was found in treatment  $T_{10}$ ,  $T_9$  and  $T_7$ .

**Harvest index:** Harvest index varied non-significantly among the different treatment of phosphorus and zinc application (Table 3). The highest harvest index was in T<sub>8</sub> and T<sub>7</sub>. Improved crop growth from adequate phosphorus and zinc likely regulated the starch/sucrose ratio in reproductive organs and leaves. Phosphorus enhanced fruiting and facilitated better translocation of essential metabolites to yield-contributing parts, boosting grain yield. Large variations in harvest index are not influenced by any of the given treatments and this might be due to character, highly associated with genetic makeup of the crop (Singh et al 2011). Each increment of phosphorus from (25 to 75 kg ha<sup>-1</sup>) gave superior HI value of lentil (Fatima et al 2013).

### **Economic Studies**

**Gross returns (₹ ha**<sup>-1</sup>): The gross returns was highest in the case of  $T_{10}$  followed by the  $T_{12}$  (Table 3). These returns may be ascribed to the higher yield of the treatments due the balanced application of the nutrients. The higher growth parameters and yield attributes resulted in higher gross returns.

 Table 3. Effect of different treatments of phosphorus and zinc on grain, straw, biological yield, grain straw ratio, harvest index, cost of cultivation, net returns and B:C ratio

Treatments		Yield (kg ha <sup>-1</sup> )	)	Grain straw	Harvest index	Net returns	Cost of	B: C ratio	
	Grain	Straw	Biological	ratio		(₹ na )	cultivation (₹ na )		
T,	1318	2591	3909	0.52	0.34	40231	22058	1.82	
<b>T</b> <sub>2</sub>	1418	2894	4271	0.49	0.35	43906	23183	1.89	
T <sub>3</sub>	1381	2880	4198	0.52	0.33	42005	23308	1.80	
T <sub>4</sub>	1363	2731	4179	0.49	0.34	42091	22103	1.90	
<b>T</b> <sub>5</sub>	1322	2668	4049	0.49	0.35	40123	22103	1.82	
<b>T</b> <sub>6</sub>	1363	2816	4216	0.48	0.33	41791	22148	1.89	
Τ <sub>7</sub>	1522	2890	4298	0.53	0.31	49161	22666	2.17	
T <sub>8</sub>	1341	2708	4094	0.49	0.37	40410	22728	1.78	
T <sub>9</sub>	1531	2930	4475	0.53	0.33	49293	22711	2.17	
T <sub>10</sub>	1735	3424	5063	0.53	0.35	58460	23336	2.51	
T <sub>11</sub>	1544	2980	4502	0.52	0.32	53399	24183	2.21	
T <sub>12</sub>	1563	3333	4544	0.47	0.33	55101	24246	2.27	
CD (p=0.05)	191	NS	383	NS	NS	-	-	-	

See Table 1 for details

**Net returns (₹ ha<sup>-1</sup>):** The highest net returns was obtained in  $T_{10}$  whereas the minimum net return was in  $T_1$  (Table 3). Yadav et al (2017)observed that the application of phosphorus at 40 kg ha<sup>-1</sup> gave significantly higher net returns. **B: C ratio:** The highest B:C ratio was obtained in  $T_{10}$  followed by the  $T_{12}$  at 2.27 (Table 3). The minimum was in  $T_1$ . This indicates amount of money earned from investment of a unit amount of money. The higher B:C ratio may be ascribed to higher net returns rupee<sup>-1</sup> of cost. The nutrients applied caused better output when both phosphorus and zinc were combined along with the solubilizing microbes and led to higher gross and net returns which also led to higher B:C ratio (Singh et al 2019).

#### CONCLUSION

The application of. 50% RDP @24 +12.5 kg  $ZnSO_4$  + Biophos + Biozinc (5 ml kg<sup>-1</sup> seed each) proved effective in significantly enhancing the plant height, number of branches plant<sup>-1</sup>, number of compound leaves plant<sup>-1</sup>, dry matter accumulation plant<sup>-1</sup>, number of days taken to flowering and maturity and B: C ratio. The research demonstrates the significant impact of phosphate and zinc solubilizing microbes on growth and yield of lentil. The synergistic interactions between these beneficial microbes and phosphorus and zinc fertilizers not only enhances nutrient availability but also promotes sustainable agricultural practices.

#### **AUTHORS CONTRIBUTION**

Monica Yaying: Conceptualization and execution of field experiment and laboratory analysis, data analysis, writing; Anil Shukla: Conceptualization of research work, supervision; Supriya: Preparation of manuscript, editing; Sudarshan S: Data analysis and editing; Shobhana Singh: Editing; Gunashekhar H: Preparation, correction and revision of manuscript; Chandra Bhushan: Correction and revision

#### REFERENCES

- Chetan Babu RT, Mavarkar NS, Praveen BR, Singh M and Dileep R 2023. Effect of water-soluble fertilizer and PGPR on soil microbial population, nodule count and economics of black gram. *Indian Journal of Ecology* **50**(1): 95-98
- Fatima K, Hussain N, Pir FA and Mehdi M 2013. Effect of nitrogen and phosphorus on growth and yield of lentil (*Lens culinaris* M.). *Elixir Applied Botany* **57**: 14323-14325.
- Gaine A, Solanki RB and Allie FA 2009. Effect of biofertilizers on growth and yield of gardenpea. *The Asian Journal of Horticulture* **4**(2): 507-509.
- Hussain I and Ahmad B 2015. *Effect of nitrogen and zinc application on growth, yield and seed recovery of wheat crop.* MSc (Hons) thesis, KPKAgriculture University, Peshawar, Pakistan.
- Kalayu G 2019. Phosphate solubilizing microorganisms: Promising approaches as biofertilizer. International Journal of Agronomy 7.
- Kant S, Kumar A, Kumar S, Kumar V, Pal Y and Shukla AK 2016.

Effect of rhizobium, PSB and P-levels on growth, yield attributes and yield of urdbean (*Vigna mungo* M.). *Journal of Pure and Applied Microbiology* 10(4): 3093-3098

- Kasturikrishna S and Ahlawat IPS 2000. Effect of moisture stress and phosphorus, sulphur and zinc fertilize₹ on growth and development of pea (*Pisum sativum* L.). Indian Journal of Agronomy **45**(2): 353-356.
- Meena RJ, Singh RK, Singh NP, Meena SK and Meena VS 2015. Isolation of low temperature surviving plant growth promoting rhizobacteria (PGPR) from pea (*Pisum sativum* L) and documentation of their plant growth traits. *Biocatalysis and Agricultural Biotechnology* **4**(4): 806-811
- Meena SM, Laharia GS, Hadole SS, Sathyanarayana E, Dhule DT and Ghato PU 2017. Interactive effect of phosphorus and zinc on yield, quality and fertility status of soil after harvest of soybean. *Multilogic in Science* **6**(19)
- Pandey SK, Bahuguna RN, Pal M, Trivedi AK, Hemantaranjan A and Srivastava JP 2010. Effects of pretreatment and foliar application of zinc on growth and yield components of mungbean (*Vigna radiata* L.) under induced salinity. *Indian Journal of Plant Physiology* **15**(2): 164-167.
- Rasheed M, Jilani G, Shah IA, Najeeb U and Iqbal T 2010. Improved lentil production by utilizing genetic variability in response to phosphorus fertilization. *Soil and Plant Science* **60**: 485-493.
- Siddiqui SN, Umar S, Husen A and Iqbal M 2015. Effect of phosphorus on plant growth and nutrient accumulation in a high and a low zinc accumulating chickpea genotypes. *Annals of Phytomedicine* **4**(2): 102-105.
- Sindhu SS, Sharma R, Sindhu S and Phour M 2019. Plant nutrient management through inoculation of zinc-solubilizing bacteria for sustainable agriculture. *Biofertilizers for Sustainable Agriculture and Environment* 173-201.
- Singh AK and Bhatt BP 2013. Effect of foliar application of zinc on growth and seed yield of late-sown lentil (*Lens culinaris* M.). *Indian Journal of Agricultural Sciences* **83**(6): 622-626.
- Singh G, Virk HK and Khanna V 2017. Integrated nutrient management for high productivity and net returns in lentil (Lens culinaris). *Journal of Applied and Natural Science* **9**(3): 1566-1572.
- Singh G, Virk HK, Aggarwal N, Gupta RK and Khanna V 2019. Symbiotic parameters, growth, nutrient accumulation, productivity and profitability as influenced by integrated nutrient management in lentil (*Lens culinaris* M.). *Archives of Agronomy and Soil Science* **65**(3): 411-420.
- Singh G, Sekhon HS and Sharma P 2008. Studies on the effect of sources and levels of phosphorus and phosphate solubilizing bacteria on microbial traits and productivity of chickpea and lentil. *Journal of Plant Science and Research* **24**: 41-46.
- Singh G, Ram H, Sekhon HS, Aggarwal N and Khanna V 2011. Effect of nutrient management on nodulation, growth and yield of lentil (*Lens culinaris* M.) genotypes. *American-Eurasian Journal of Agronomy* 4: 46-49.
- Sah U, Rani R, Kumar H, Ojha J, Singh V, Dubey S and Dixit G 2022. Dynamics of lentil (*Lens culinaris*) production and trade: Global scenario and Indian interdependence. *The Indian Journal of Agricultural Sciences* **94**(3-1): 102-108.
- Singh OB, Shama HB and Singh VK 1999. Effect of nitrogen, phosphorus and rhizobium culture on yield and yield attributes of lentil under dry land conditions. G.B Pant University Agricultural & Technology Pantnagar, Uttarakhand, India.
- Tagore GS, Namdeo SL, Sharma SK and Kumar N 2013. Effect of rhizobium and phosphate solubilizing bacterial inoculants on symbiotic traits, nodule leghemoglobin and yield of chickpea genotype. *International Journal of Agronomy* **8**: 581627.
- Yadav M, Yadav SS, Kumar S, Yadav HK and Tripura P 2017. Effect of phosphorus and bio-fertilizers on yield, nutrient content and uptake of urban [*Vigna mungo* L.]. International Journal of Current Microbiology and Applied Science **6**(5): 2144-2151.

Received 01 July, 2024; Accepted 21 September, 2024



Manuscript Number: 4356 NAAS Rating: 5.38

# Morphological and Chemical Profile of *Pulicaria undulata* L. (Compositae) in Iraq

# Khansaa R. Al-Joboury

Iraq Natural History Research Center and Museum, University of Baghdad, Baghdad, Iraq E-mail: dr.khansaa@nhm.uobaghdad.edu.iq

**Abstract:** The morphological features of *Pulicariaundulata* (L.) C.A. Mey. were observed. This species is a perennial plant, twiggy subshrub 30-120 cm high with a deep root system, leaves are lanceolate, semi-amplexicaul and decurrent on one side, Inflorescence in irregular cymes, peduncles 4-5 cm long, Involucre 5-7 × 3-4 mm, phyllaries numerous, linear, brown, hairy or glabrous. Kaempferol, kaempferol 3-methyl ether, 6-methoxykaempferol, quercetin, quercetin 3-methyl ether, quercetin 3,7-dimethyl were estimated using HPLC chromatography.

Keywords: Compositae, Chemical study, Iraq, Morphological study, Pulicaria

Pulicaria undulata (L.) C.A.Mey. (1831) (syn. Francoeuria crispa subsp. crispa, F.crispa var. crispa, Pulicaria crispa subsp. crispa, P. crispa var. crispa, P. crispa var. gracillima Maire, P. undulata var. undulata, belongs to the family compositae (asteraceae) (GBIF Secretariat, 2020). This is native in Iraq, Iran, Yemen, Afghanistan, Algeria, Benin, Chad, Djibouti, Egypt, Eritrea, Ethiopia, Gulf States, India (Dawar et al 2002). This species very important medicinal plant known as Dethdath, and used medicinal plants to make traditional medicines all over the world since ancient times. The branches and flowers are used to prepare powders for sneezing which works to repel insects, also in the tea of herbal (Khansaa et al 2017). This species is used for a traditional medicine which act as tonic, antispasmodic, antihypoglycemic drugs and ingredients of perfumes (Ghazanfar and Edmondson 2019) and is rich source forlarge number of bioactive compounds such as flavonoids which are thesecondary plant metabolitesandare compounds of low molecular weight. They are chemically polyphenolic and their nature offers a common structure, benzo-y-pyrone (Ahmed and Ibrahim 2018). These flavonoids are used by the action of antioxidants, antimicrobials, photoreceptors, optical attracts (Khalid et al 2020). This study aimed to characterize the plant through morphologic characteristics and toevaluate the flavonoids present in it.

# MATERIAL AND METHODS

Twelve plant were collected from Al-Zafaraniyah/ Baghdad- Iraq in 2020during the flowering period in Aprilat 3 geographical locations, the coordinate of these stations was 33°15'49.0"N 44°29'16.6"E, 33°15'44.4"N 44°29'13.1"E and 33°15'34.7"N 44°29'17.4"E.. The taxonomical key was used to identify the specimens of this species for precise identification (Ghazanfar and Edmondson 2019). After being air dried in nature, the aerial parts of this species were ground in a mortar and flavonoids were isolated from *Pulicaria undulata*in pure form. Five grams of plant material separated in 100 ml of methanol using maceration (48 h). Then removed in the vacuum with the temperature at 50°C and the extracts were freeze-dried (Romanik et al 2007).

#### **RESULTS AND DISCUSSION**

Morphological study: Pulicaria undulatais perennial plant and twiggy subshrub were 30-120 cm high with adeep root. Plant branches from the base, white-floccose or light gray in color, densely glandular, and take the form of extensive cushions. Its branches are many and may intertwine. The leaves are very long at the bottom and shorter towards the top with shape as lanceolate, semi-amplexicaul and decurrent on one side. The margin of the leaves are very clearly undulating, 2-25×1-5mm, base almost auriculate (Fig. 1). Inflorescence is irregular cymes, peduncles 4-5 cm long, involucre 5-7 × 3-4 mm, phyllaries numerous, linear, brown, hairy or glabrous Ghazanfar and Edmondson (2019) also mentioned similar observation . Corolla yellow, 2.75-3 mm long, the ligule only 1 mm, and the disc florets 2.5-3.5 mm long, with glandular hairs. Cypsela 0.8 mm long, glabrous, brown, 5-ribbed, the bristles with a short narrow tuft of slightly longer hairs at the tip. Outer pappus is ring of scales and inner pappus of bristles (short stiff hair) joined at the base into a tube, each bristle with a narrow apical tuft of slightly longer hairs. The morphology gave as clear features of this species because assemblage for species and is very useful for the

delimitation and identification. There were two types in trichomes that spread densely in its parts. The non-glandular and glandular which spread and concentrated in all parts organs: leaves, stems and flowers (Abid and Qaiser 2002) Trichome diversity were divided.

**Non-glandular trichome:** These are found in all plant parts and are simple non-glandular unicellular trichomes, different lengths ranging from short to very long about 10-860µm, curved all with acute apex (Fig. 2. A 1- A 3) (Krak and Mraz 2008).

**Non-glandular multicellular trichomes**: These are also found in all parts of plants, some trichome fragmented from ending with a thin flagellum, others have basal cell broader,3-

7 cells long and the apical cell like a conical shape, others consists of two cells of different lengths, straight or curved and all with obtuse apex (Fig. 2. A 3- A7) (Al-dobaissi et al 2016).

**Stellate trichomes:** The sparse stellate hairs are on the involucral bracts (Fig. 2.A8).

*Glandular trichome:* These were multicellular divided into head and stalk (Fig. 2. B1) (Valkama et al 2003, Ciccarelli et al 2007).

**Chemical profile**: Six different flavonoids were extracted and identified: Kaempferol, kaempferol 3-methyl ether, 6methoxykaempferol, quercetin, quercetin 3-methyl ether, quercetin 3,7-dimethyl ether (Fig. 3). The peaks for target



Fig. 1. Morphological characteristics of *Pulicaria undulata*a: Whole plant, b: Root, c: the stem, d1-3: Different lengths of leaves, e-i: Inflorescence, j: Pappus calyx



**A**7



Fig. 2. Morphological characteristics for the trichomes of the species *Pulicaria undulata*A1-10:Non-glandular trichomes (300x), B1-5 Glandular trichomes(400x)



Fig. 3. Morphological form for the flavonoid compounds of the species P. undulata

83.22

40.32

Flavonoid compounds Retention time (mints) Area Concentration (mg/mL) Kaempferol 47.82 160.55 Kaempferol 3-methyl ether 52.38 50.32 6-Methoxykaempferol 44.71 40 13 Quercetin 34.41 36.66

Table 1. The flavonoids component with retention time and area of this species

flavonoidseluted at retention times 47.82, 52.38, 44.71, 34.41, 83.22 and 40.32 min, respectively, through the sample extract (Table 1). Fahmiet al (2019) also made similar observation.Kaempferol recorded the highest value of 160.55 but guercetin recorded the lowest value both in retention time and area which was 36.66 and 34.41, respectively. The highest concentration recorded for kaempferol was 0.26 mg/mL. Flavonoids protect plants against different biotic and abiotic stresses, and responsible for a very important role in resistance to frost, hardiness, drought resistance, also have a functional role for plant heat acclimation and freezing tolerance (Vieira et al 2003). Mierziak et al (2014) reported that the essential oil of the aerial parts for P. gnaphalodes has shown the presence of flavonoids, but studies on the concentrations of flavonoids are very rare in Iraq.

#### CONCLUSION

The study verified the morphological features of Pulicaria undulate (L.) C.A. Mey. (Asteraceae) which collected from Al-Zafaraniyah/ Baghdad- Iraq. Trichomes (hairs) weredivided into non-glandular and glandular. Various chemical kaempferol, kaempferol 3-methyl ether, 6methoxykaempferol, quercetin, quercetin 3-methyl ether, quercetin 3, 7-dimethyl were estimated by HPLC chromatography. This will be helpful for medicinal use and further investigations.

#### REFERENCES

- Ahmed SSand Ibrahim ME 2018. Chemical investigation and antimicrobial activity of Francoeuria crispa (Forssk) grown wild in Egypt. Journal of Materials and Environmental Science 9(1): 1-
- Abid RD and Qaiser M 2002. Cypsela morphology of Inula L. (s.str.) and its allied genera (Inuleae-Compositae) from Pakistan and

Received 18 June, 2024; Accepted 21 September, 2024

Kashmir. Pakistan Journal of Botany 34(3): 207-223.

90.71

110.53

Al-Dobaissi AM, Al- shimmery K and Almousawy AH 2016. Cuticular indumentum of some species from Asteraceae family in Erbil province Iraq. Iraqi Journal of Science. Special issue-part b: 251-262

0.26

0.11 0.05

0.04

0.15

0.20

- Ciccarelli D, Garbari F and Pagni AM 2007. Glandular hairs of the ovary: A helpful character for Asteroideae (Asteraceae) taxonomy. Annales Botanici Fennici 44: 1-7.
- Dawar R, Qaiser M and Perveen A 2002. Pollen morphology of Inula L. (s.str.) and its allied genera (Inuleae-Compositae) from Pakistan and Kashmir. Pakistan Journal of Botany 34(1): 9-22.
- Fahmi AA, Abdur-Rahman M, Naser AF, Hamed MA, Abd-Alla H I, Shalaby NM and Nasr MI 2019. Chemical composition and protective role of Pulicaria undulata (L.) C.A. Mey. subsp. undulata against gastric ulcer induced by ethanol in rats. Heliyon 5(3): e01359.
- GBIF Secretariat 2020. Pulicaria undulata (L.) C.A.Mey. https://www.gbif.org/species/3095567
- Ghazanfar SA and Edmondson JR 2019. The Flora of Iraq, 6. 394-395. Ministry of Agriculture Republic of Iraq by Royal Botanic Gardens. Kew.
- Khalid HA. Oraibi AG. Yahva HN and Al-Obaid JR 2020. Phytoremediation of Zinc Ion Using Faba Bean Plant Vicia faba in vitro. Indian Journal of Ecology 47(4): 965-968.
- Khansaa RA, Lubab GA and Sukeyna AA 2017. Morphological, anatomical and numerical taxonomy studies for some species of the fabaceae family. Journal of Biodiversity and Environmental Sciences 11(5): 117-123.
- Krak K and Mraz P 2008. Trichomes in tribe Lactuceae (Asteraceae)-Taxonomic implication. *Biologia* **63**(5): 616-630.
- Mierziak J, Kostyn K and Kulma A 2014. Flavonoids as important molecules of plant interactions with the environment. Molecules 19(10): 16240-16265.
- Romanik G, Gilgenast E, Przyjazny A, Kamiński M 2007. Techniques of preparing plant material for chromatographic separation and analysis. Journal Biochemical Biophys Methods 70: 253-261.
- Valkama E, Salminen JP, Koricheva J and Pihlaja K 2003. Comperative analysis of leaf trichome structure and composition of epicuticular flavonoids in finnish Birch species. Annals of Botany 91: 643-655.
- Vieira RF, Graver RJ and Paton A 2003. Chemical Profiling of Ocimum americanum using external flavonoids. Phytochemistry-\r: 000557.

Quercetin 3-methyl ether

Quercetin 3,7-dimethyl ether



# Impact of Western Disturbances on Stone Fruits in Mid Hill Zone of Himachal Pradesh

Prakriti Dadial, M.S. Jangra, S.K. Bhardwaj, Purnima Mehta and Akanksha Klate<sup>1</sup>

Department of Environmental Science and <sup>1</sup>Department of Social Sciences, Dr. YS Parmar University of Horticulture & Forestry Nauni, Solan-173 230, India E-mail: prakritidadial83@gmail.com

**Abstract:** The study on impact of western disturbances on stone fruits grown in mid hill zone of Himachal Pradesh" was done at Dr. YS Parmar University of Horticulture and Forestry, Nauni. The western disturbances (WD) data was collected for the period 1984-2022. The arrival and withdrawal of WD indicated that during the period of 38 years the WD arrived during October and withdrawal during May in Solan district of Himachal Pradesh. The WD duration and amount of rainfall received was decreasing at 0.29 Julian days/year and 1.65 mm/year but, the number of WD was increasing at a rate of 0.12 JD/year which indicated that the frequency, distribution and intensity of WD was increasing over time in mid hills of Himachal Pradesh. The regression relationship was developed between the number of WD and productivity at development and maturity stages of plum and apricot crops. The results depicted positive relationship between number of western disturbance and productivity at developmental stages with coefficient of determination (R<sup>2</sup>: 0.58 and R<sup>2</sup>: 0.51) whereas showed negative relationship at maturity stage of selected crop.

Keywords: Frequency, Western disturbance, Monthly variation, Regression, Crop

Western disturbances (WD) are cyclonic circulation/ trough in the mid and lower tropospheric levels or as a lowpressure area occurs in mid-latitude westerlies and originates over the mediterranean Sea, Caspian Sea and Black Sea and moves eastwards across north India. These are the most fundamental level, synoptic-scale vortical perturbations embedded in the subtropical westerly jet stream (Dimri et al 2016) and are most common during December to March, bring rain to the Western Himalayas as well as to the surrounding areas of north India, Pakistan and the Tibetan Plateau and also associated with weather hazards such as heavy snowfall, hailstorms, fog, cloudbursts, avalanches, frost, and cold waves (Hunt et al, 2024). The erratic and increasing pattern of rainfall in Karnataka district may lead to the flash floods and at times drought situations (Bharath and Venkatesh 2022).

Agarwal et al (2021) also analysed that in early 1950s, winter and monsoon rainfall was showing decreasing trend whereas the summer and post-monsoon was showing an increasing trend resulting into a westward shift, more variable and declining rainfall over the country. Himachal Pradesh is situated in the western Himalayas and is divided by altitudinal ranges of 350 m to 6975 m above sea level due to a complex geographical feature which results in a variety of climatic patterns, from hot and humid tropical climates in the south to frigid, alpine, and glacial climates in the eastern and northern mountain ranges. According to the report of Himachal Pradesh State Action Plan on Climate Change (2012) projects states that during the last 25 years, there has been a 40% decrease in rainfall. Kumar et al (2019) highlights the vulnerability of stone fruit cultivation in the face of such climatic variability. The study underlines the importance of understanding the specific impacts of western disturbances on stone fruit crops to devise effective adaptations and mitigation strategies.

Western disturbances exert a notable influence on stone fruit cultivation in the mid-hill zone of Himachal Pradesh, necessitating a comprehensive understanding of their impacts and the implementation of suitable measures to ensure the sustainability of agricultural practices. The main objective of the study is to investigate how WD influences phenological stages (flowering, fruit set, ripening) of stone fruit crops in the mid-hill zone of Himachal Pradesh.

#### MATERIAL AND METHODS

**Experimental site:** The study was conducted at Dr. Y.S. Parmar University of Horticulture and Forestry, Nauni-Solan (30°2240– 33°1240N and 75°4755-79°0420E) and 350 m to 6,816 m amsl during 2021-2023.

**Data collection:** The daily rainfall and rainy days data of 38 years (1984-2022) was collected from Agromet observatory, Department of Environmental Science, Nauni Solan whereas the data on area and production of stone fruit crops *viz.*, Plum and Apricot for last 30 years (1990-2020) for Solan district was collected from the Department of Horticulture, Govt. of Himachal Pradesh.

### Analysis of Western Disturbances

**Duration and Frequency of western disturbances:** The rainfall 2.5mm/day and more was considered as a WD spell. If the spell was continued for one day than WD spell is considered for 1 day. If the spell was continued for two days than WD spell is considered for 2 days and so on if the spell was continued for five days, than WD spell is considered for 5 days. The frequency of western disturbances was considered as the number of WD occurred during a month, season or year. Gill et al (2021) calculated the frequency of occurrence of WD duration for each month separately.

Frequency = Total rainfall / Total numbers of WD Linear regression analysis: Linear regression analysis is a parametric model and one of the most commonly used methods to describe a trend in a data series and develops a relationship between two variables by fitting a linear equation to the observed data. The data is first checked whether there is relationship between the variables of interest or not. This can be done by using the scatter plot. The linear regression model is generally described by the following equation:

Y=aX+b

Y= dependent variable, X= independent variable, a= slope of line, b= intercept constant.

### **RESULTS AND DISCUSSION**

Arrival, withdrawal and duration of western disturbances: The date of arrival of WD was decreasing (advancing) at a rate of 0.19 Julian days/year. Out of 37 years the arrival was below the normal for 21 years and above the normal during 16 years, it means that the WD arrival was early in 21 years and getting late during 16 years in the study area (Fig. 1) and the withdrawal date was increasing (delaying) at a rate of 0.11 Julian days/year (Fig. 2). Out of 37 years, 12 years showed the withdrawal below and remaining 25 years above the normal date of withdrawal of WD. The withdrawal of western disturbances was advanced in 12 years and delayed in 25 years in the study area. Dadial et al (2024) also studied the different behavioural characteristics of western disturbances (WD) like onset, withdrawal, durations and amount of rainfall in WD in Solan district of Himachal Pradesh. The annual duration of WD was decreasing at a rate of 0.29 Julian days/year, means decreasing at about 8 hours/year (Fig. 3).

Annual variation of number of WD and associated rainfall : The highest number of WD had been observed in one day duration (405) followed by two days duration (234) up to more than five days duration in the descending order whereas the associated rainfall was observed highest (4692.1 mm) for the WD of two days duration followed by one day duration and three days duration whereas lowest for the

WD of more than five days duration. The frequency of WD spell (61.8 mm) was received highest in five days WD spell followed by four days WD spell (61.1mm). Deviation from mean value was highest for two days duration and lowest for more than five days duration whereas coefficient of variation was calculated highest for more than five days duration and lowest for one day duration. The positive slope and trend were observed increasing for the WD of one to four days duration whereas decreasing for five and more than five days duration (Table 1).

The variation in annual number of WD was increasing at a rate of 0.12JD/year (Fig. 4) whereas, annual rainfall received from WD was decreasing at a rate of 1.66 mm per year (Fig. 5). Similar results were obtained by Jaswal et al (2015) indicated that seasonal and annual trends in rainfall over Himachal Pradesh for 37 stations from 1951 to 2015 significantly decreasing by 4.58 mm/year. All the 37 stations in Himachal Pradesh were not showing an increasing trend in



Fig. 1. Annual deviation (days) of WD from normal date of arrival



Fig. 2. Annual deviation (days) of WD from normal date of withdrawal



Fig. 3. Deviation in day from normal duration of western disturbances

Durations (Days)	Number of WD	Rainfall (mm)	Mean	SD (σ)	CV (%)	Slope (Year)	Trend	Frequency
1	405	3321.5	44.8	81.01	55.3	0.11	↑	8.2
2	234	4692.1	67.7	114.44	59.1	0.06	$\uparrow$	20.0
3	91	3309.2	71.9	80.71	89.1	0.16	$\uparrow$	36.3
4	39	2385.3	65.7	58.18	113.0	0.68	↑	61.1
5	10	618.6	41.8	15.09	277.6	-0.36	$\downarrow$	61.8
>5	6	358.3	25.3	8.74	290.4	-0.50	$\downarrow$	59.7

Table 1. Annual variation in WD of different durations from 1984-2022





Fig. 4. Annual variation in number of western disturbances

Fig. 5. Annual variation of rainfall received through western disturbances



Fig. 6. Effect of number of WD on crop yield occurred at development stage



Fig. 7. Effect of number of WD on crop yield occurred at maturity stage

annual rainfall. Harshavardhan et al (2020) investigated the spatiotemporal trends in rainfall in Krishna River Basin in India from 1965 to 2012 and observed that annual and postmonsoon precipitation exhibited a significant negative trend in magnitude from the normal. **Regression relationship of number of WD with productivity of stone fruit crops:** The linear regression model was developed between number of western disturbance and productivity of Plum and Apricot crop at development and reproductive stage. At development stage

both the crops showed the positive relationship with variation of 58 and 51 percent (Fig. 6) whereas showed negative relationship at reproductive stage with variation of 19 and 18 per cent (Fig. 7). The results inferred that the increase in the number of western disturbances caused detrimental effects on crop yield at a rate of -0.79 and -0.80 guintal per number of WD during reproductive to harvesting stage. The occurrence of heavy rainfall accompanied with high wind speed due to western disturbances at flowering stage will be washed out pollen from the stigma of the flower, resulting in poor or no fruit setting. Similarly, at the time of maturity, it leads to fruit dropping or poor quality of fruit. Mehta et al (2022) highlighted that erratic rainfall patterns led to a shorter cropping calendar, affecting agricultural practices. Over the past three decades, farmers have observed a mismatch in rainfall, with insufficient rain during crucial periods and excessive rain when it is not needed. Dadial et al (2024) reported that rainfall at development stage showed beneficial effects on wheat, barley, and peach crops whereas, rainfall during the months of April and May impacts the harvesting and threshing of wheat crop thus decreases the quality and quantity of crop. The hailstorm events in the month of April and May reduced the yield of peach crop in mid hill zone of Himachal Pradesh.

### CONCLUSIONS

The annual duration of western disturbance and amount of rainfall received from October to May depicted decreasing trend. While the western disturbance spells of day 1 to 4 showed increasing trend whereas day 5 and >5 spells showed decreasing trend thus indicated that the frequency, distribution of WD was increasing over time in mid hills of Himachal Pradesh. The regression model developed between the number of WD and productivity depicted positive relationship at developmental stages while showed negative relationship at maturity stage of selected crops. Hence, farmers should reschedule the plant protection

Received 23 July, 2024; Accepted 30 September, 2024

measures and irrigation activity according to the weather forecast during critical growth stages. The farmers should sow the crops at optimum date and avoid late sowing of crops to prevent the maturity and harvesting stages from coinciding with peak WD activities. With a decline in the amount of rainfall during WDs, ensure proper irrigation management to supplement water needs during critical growth stages. Rainwater harvesting systems can be developed to store water during periods of increased WD intensity and redistribute it when needed.

#### REFERENCES

- Agarwal S, Suchithra AS and Singh SP 2021. Analysis and interpretation of rainfall trend using Mann-Kendall's and Sen's Slope method. *Indian Journal of Ecology* **48**: 453-457.
- Bharath AL and Venkatesh B 2022. Precipitation Concentration Index and Rainfall Trend Analysis for South Western Districts of Karnataka India. *Indian Journal of Ecology* **49**(2): 462-469.
- Dadial P, Singh M and Mehta P 2024. Trend and frequency distribution of western disturbances and its impact on major crops of Solan district of Himachal Pradesh. *Journal Of Agrometeorology* 26(2): 190-195.
- Dimri A P, Yasunari T, Kotlia BS, Mohanty UC and Sikka DR 2016. Indian winter monsoon: Present and past. *Earth-Science Reviews* **163**: 297-322.
- Gill KK, Kaur S, Sandhu SS and Bhatt K 2021. Frequency of occurrence and duration of western disturbances in Punjab. V-AGMET conference paper.
- Harshavardhan PL, Nayak PC and Kumar S 2020. Spatio-temporal rainfall variability and trend analysis for Krishna River Basin in India. *Indian Journal of Ecology* **47**: 54-59.
- Hunt KMR, Baudouin JP, Turner AG, Dimri AP, Jeelani G, Pooja, Chattopadhyay R, Cannon F, Arulalan T, Shekhar MS, Sabin TP and Palazzi E 2024. Western disturbances and climate variability: A review of recent developments. *EGUsphere*: 1-106.
- Jaswal AK, Bhan SC, Karandikar AS and Gujar MK 2015. Seasonal and annual rainfall trends in Himachal Pradesh during 1951-2005. Mausam 66: 247-264.
- Kumar M 2019. Effect of Climate Change on Indian Economy and Agriculture. International *Journal of Scientific Research and Review* 07(01): 2279-543.
- Mehta P, Jangra MS, Bhardwaj SK and Paul S 2022. Variability and time series trend analysis of rainfall in the mid-hill sub humid zone: a case study of Nauni. *Environment Science and Pollution Research* **29**: 66-76.



# Standardization of Growing Media for Haworthia Pot Plant

Alka Singh, G.D. Patel, H.P. Shah, A.J. Bhandari, R.A. Gurjar and S.T. Bhatt

Department of Floriculture and Landscape Architecture, ASPEE College of Horticulture, Navsari Agricultural University, Navsari-396 450, India E-mail: alkaflori@nau.in

**Abstract:** Investigation was carried out to standardize growing media for *Haworthia attenuta*, a popular succulent pot plant at Navsari Agricultural University, Navsari. The experiment consisted of different growing media viz., $M_1$ - cocopeat + vermicompost + rice husk (6:1:3),  $M_2$ -cocopeat + vermicompost + perlite (6:1:3),  $M_3$ - cocopeat + vermicompost (9:1),  $M_4$ -local black soil + vermicompost+sand (6:1:3),  $M_5$ - cocopeat + vermicompost + charcoal (7:1:2),  $M_6$ -sand+vermicompost (9:1) and M7- white grit+vermicompost (9:1) in 10x10 cm pot. The observations were recorded at every three months interval, and pooled data analysis of three years was done. Among different growing media, sand: vermicompost (9:1) was highly suitable with regard to plant height, plant spread and number of leaves. Further, number of suckers as well as root length and number of roots were observed maximum in sand: vermicompost (9:1) growing media. Thus, growing media comprising of sand: vermicompost (9:1) was highly suitable for growing Haworthia as pot culture to obtain better plant growth and quality.

Keywords: Succulent, Haworthia, Pot plant, Growing media, Sand, Cocopeat, Vermicompost

Pot plant trading is recently gaining the impetus of an industry with the growing demand of guality ornamental plants in domestic as well as international trade. Rapidly growing urbanization and population explosion with hasty life style have created space paucity for living, wherein pot plants offers some respite (Singh et al 2016, Guddad et al 2022, Sindhuja et al 2022). In critical times that prevailed during the epidemic COVID-19, indoor gardening helped to enhance sense of emotional comfort and created sense of stress free happy environment (Altman 2011, Singh et al 2020). Succulent plants are increasingly in demand as indoor pot plant for urban horticulture (Dewir 2016, Cabahug et al 2018) because of specific characteristics like drought resistance (Baldwin 2013, Edwards and Ogburn 2013) and low light tolerance (Bell 2001, Nefzaoui 2007). Drought-tolerant plants are increasingly recognized as a resource to mitigate the consequences of climate change and need to be preserved (Grace 2019). Besides, succulents are known to exhibit crassulacean acid metabolism (CAM), resulting in CO<sub>2</sub> assimilation during night time (Lee 2010, Herrera 2009) and thus contribute in improving indoor air quality.

Haworthia is a small succulent ornamental plant native to South Africa, belongs to family *Asphodelaceae* and is native to South Africa. The cultivation of haworthia as pot plant is increasing in different countries owing to its small compact growth habit and drought tolerance characteristic (El Shamy et al 2018, Grace 2019). Its popularity and demand is further increasing for offices and homes as it can flourish at low light levels also. The successful potting media should provide good physical support for the plant, a reservoir for nutrients and water with sufficient drainage and aeration. Research on growing media standardization in succulents is meagre with no previous work on haworthia. Hence, research was conducted to standardize growing media for haworthia as pot plant.

#### MATERIAL AND METHODS

The present study was carried out at Navsari Agricultural University, Navsari-396450, Gujarat during 2017-2020 on popular succulent indoor plant Haworthia attenuta in a protected structure cladded with 200 micron UV stabilized film and 50 per cent shade net. The treatments consisted of seven growing media viz., M<sub>1</sub>- Cocopeat + vermicompost + Rice husk (6:1:3), M<sub>2</sub>- Cocopeat + vermicompost + perlite (6:1:3), M<sub>3</sub>- Cocopeat + vermicompost (9:1), M<sub>4</sub>- Local black soil+ vermicompost+ sand (6:1:3), M<sub>5</sub>- Cocopeat + vermicompost + Charcoal (7:1:2), M<sub>6</sub>-Sand+vermicompost (9:1) and M7- White grit+vermicompost (9:1) in 10x10 cm uv stabilised plastic pot. The experiment was laid out in completely randomized design with three repetitions for three consecutive years and each year new plantation was done. Uniform sized plants were planted in different growing media and uniform management practices were given for all treatments. As nutrient management, NPK (19:19:19) at 250 mg/l was given @ 50 ml per plant at monthly interval. Observations on different parameters like plant height, plant spread (E-W and NS), number of leaves and number of suckers were recorded at every three months of interval i.e.

after 3, 6, 9 and 12 months of planting. Further, number of roots and root length was measured at the beginning and after one year of period and the incremental change was determined. The experiment was repeated for three years. Chemical and physical properties of different growing media pH, EC and bulk density was estimated before the experiment (Table 4). The data of three years were pooled and statistically analysed by using the OP stat software (Sheoran et al 12)

#### **RESULTS AND DISCUSSION**

**Plant height and plant spread:** Among different growing media, plant height was significantly higher in haworthia grown in the media M6 i.e. sand: vermicompost (9:1) as recorded after different intervals i.e. during the followed by  $M_{s}$ - cocopeat + vermicompost + charcoal (7:1:2 (Table 1). The plant spread for both sides i.e N-S and E-W, it was significantly higher in haworthia grown in the media  $M_{6}$  i.e. sand: vermicompost (9:1) followed by M5 media while M4 media showed restricted plant spread. Haworthia, being

succulent does not prefer water stagnation. Sand in M6 media and cocopeat and charcoal in M5 media contributed to better provision of aeration to the rooting system of haworthia that resulted in its better growth. The restricted plant growth in M4 media was due to the heavy black soil that holds excessive water that is not suitable for succulents in general as per some earlier work (Song et al 2007, Kaur et al 2015, Singh et al 2017). Significant influence of growing media on pot plants have been earlier observed on plant growth in indoor plants (Rashidha et al 2021), Anthurium andraeanum(Basheer and Thekkayam 2012), Chlorophytum comosum (Dhanasekaran et al 2020), Dracaena reflexa (Kavipriya et al 2019), Epipremnum aureum (Khayyat et al 2007), Euphorbia milii (Kapadiya et al 2017) and ), Senecio sp (Lodhi et al 2021).

**Number of leaves and suckers:** Among different growing media, number of leaves were significantly higher in Haworthia grown in the media M6 i.e sand: vermicompost (9:1) at different intervals (Table 2). The number of suckers were significantly higher in Haworthia grown in the media  $M_6$ 

Table 1. Effect of different growing media on plant height and plant spread in haworthia (cm)

Treatments		Plant height				Plant spread (N-S)				Plant spread (E-W)			
	3 months	6 months	9 months	12 months	3 months	6 months	9 months	12 months	3 months	6 months	9 months	12 months	
M1	5.04	6.21	7.03	7.45	4.68	5.77	6.40	7.14	4.87	5.60	6.18	7.07	
M2	5.18	6.59	7.40	7.85	4.76	6.06	6.72	7.43	5.00	5.75	6.43	7.23	
M3	5.01	6.45	7.27	7.71	4.63	5.66	6.29	7.25	4.77	5.45	6.10	6.99	
M4	4.76	5.88	6.88	7.32	4.42	5.35	5.67	6.37	4.50	5.29	5.81	6.65	
M5	5.40	7.27	8.08	8.42	5.00	6.54	7.34	7.98	5.36	6.15	7.04	7.89	
M6	5.62	8.32	9.07	9.73	5.24	7.88	8.79	9.93	5.47	7.61	8.28	9.39	
M7	5.05	6.74	7.55	8.00	4.77	6.33	7.01	7.76	4.88	6.05	6.73	7.55	
CD (p=0.05)	0.22	0.43	0.53	0.68	0.19	1.26	1.20	1.41	0.33	1.12	1.02	NS	
CD (p=0.05) (YT)	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	

Table 2.	Effect of	different	growing	media (	on numbe	er of le	eaves and	number	of suckers	in Haworthia
----------	-----------	-----------	---------	---------	----------	----------	-----------	--------	------------	--------------

Treatments	Number o	f leaves per pla	ant after indicat	ted months	Number of suckers per plant after indicated months					
	3	6	9	12	3	6	9	12		
M1	16.66	21.36	26.88	30.33	0.79	1.17	1.42	1.80		
M2	17.84	21.87	26.22	30.73	1.10	1.32	1.63	2.09		
M3	16.84	21.85	26.22	29.84	0.83	1.23	1.34	1.75		
M4	15.36	19.85	24.32	27.17	0.24	0.53	0.90	1.13		
M5	19.15	24.22	30.52	33.63	1.20	1.49	1.79	2.26		
M6	19.36	25.17	33.62	41.62	1.46	2.12	2.45	3.37		
M7	17.14	21.62	27.70	30.47	1.12	1.34	1.74	2.28		
CD (p=0.05)	0.98	2.22	2.53	4.11	0.49	0.42	0.67	0.76		
CD (p=0.05) (YT)	NS	NS	NS	NS	NS	NS	NS	NS		

i.e Sand: Vermicompost (9:1) after 3 (19.36), 6 (25.17), 9(33.62) and 12(41.62) months (Table 2). Better plant growth in terms of leaves and suckers was the result of better rooting system indicated with more number of roots and longer root being developed in sand: vermicompost media (9:1). An appropriate growing media provides better rooting system contributes in more water and nutrients for successful in growth and development of the plants (Wazir et al 2004, Franco et al 2011, Gohil et al 2018). Sand is an important component of growing media owing to its physical properties of low EC, neutral pH and improve plant growth in a few succulents and cacti (Kapadiya et al 2017, Bell 2021, Lodhi et al 2021).

Number of roots and length of root: Among different growing media, number of roots (3.26) and length of root (3.42 cm) significantly increased in haworthia grown in the media  $M_6$  i.e. sand: vermicompost (9:1) at end of experiment (Table 3). Root can be considered a key parameter as it plays an important role for the plant to absorb the nutrient and capture water. Promotion or inhibition of root growth is known to be influenced by the physical and chemical characteristics of growing media like texture, structure, bulk

 
 Table 3. Effect of different growing number of roots and root length in haworthia

Treatments	Incremental number of roots	Incremental length of root (cm)			
M1	1.35 (1.41)	1.94			
M2	1.60 (2.15)	2.25			
M3	1.14 (2.68)	2.04			
M4	0.96 (0.44)	1.12			
M5	1.78 (2.74)	2.83			
M6	1.93 (3.26)	3.42			
M7	1.61 (2.16)	2.96			
CD (p=0.05)	0.33	0.52			
CD (p=0.05) (YT)	NS	NS			

Note: The values in parenthesis are original and outside are arcsine transformed values

**Table 4.** Effect of growing media on EC, pH and bulk density

Treatments	EC (mS/cm)	pН	Bulk density (g/cc)
M1	1.74	6.5	0.18
M2	1.33	6.8	0.30
M3	2.1	6.1	0.19
M4	1.56	6.9	0.25
M5	1.21	7.2	1.32
M6	0.16	6.7	1.70
M7	0.11	7.8	1.73

density, porosity, water-holding capacity, EC, pH, etc. (Larsen and Guse, 1997, Hartmann et al 1997, Giuliani 2024). Thus, the physical properties like low EC and neutral pH of sand+vermicompost (9:1) media and suitable bulk density contributed to better root growth in haworthia.

# CONCLUSION

Among seven different growing media combinations evaluated for haworthia, the growth of haworthia with regard to plant height, plant spread, number of leaves, number of suckers as well as root length and number of roots was found better in media comprising of sand:vermicompost (9:1). Thus, for growing haworthia as pot plant, media consisting of sand: vermicompost (9:1) is highly suitable.

# ACKNOWLEDGEMENT

The authors duly acknowledge the grant received under the project Advance technology centre of soilless system for various crops, B.H. 329/12041 supported by the state government, Gujarat.

#### REFERENCES

- Altman K 2011. At home with succulents, Altman Plants Inc. Vista, California, USA.
- Baldwin DL 2013. Succulents simplified: growing, designing and crafting with 100 easy care varieties, Timber Press Inc., Portland, London, UK.
- Basheer SN and Thekkayam SG 2012. Effect of growing media and organic nutrition on vegetative growth in Anthurium plants (*Anthurium andreanum* cv. Tropical). *Asian Journal of Horticulture* **7**(2): 354-358.
- Bell SA 2001. Growing cacti and other succulents in the garden. Guild Master Craftsman Publications, USA.
- Cabahug RA, Nam SY, Lim KB, Jeon JK and Hwang YJ 2018. Propagation techniques for ornamental succulents. *Flower Research Journal* **26**(3): 90-101.
- Dewir YH 2016. Cacti and succulent plant species as phytoplasma hosts: A review. *Phytopathogenic Mollicutes* **6**(1): 1-9.
- Dhanasekaran D, Ramya K and Sathappan CT 2020. Performance of spider plant (Chlorophytum comosum) in modular vertical green walls under various media and nutrients. *Annals of Plant and Soil Research* **22**(4):410-414.
- Edwards EJ and Ogburn RM 2013. Plant venation: from succulence to succulents. *Current Biology* 23: 340-341.
- ElShamy AM, ElHawary SS, Fahmy HA, Ali SA and Shahira ME 2018. Phytochemical investigation and antioxidant activities of certain *Haworthia* and *Gasteria* species. *International Research Journal of Pharmacy* **9**(9): 46-52.
- Franco JA, Banon S, Vicente MJ, Miralles J and Martinez-Sanchez J 2011. Root development in horticultural plants grown under abiotic stress conditions: A review. *The Journal of Horticultural Sciences and Biotechnology* 86(6): 543-556.
- Giuliani LM, Hallett PD and Loades KW 2024. Effects of soil structure complexity to root growth of plants with contrasting root architecture. *Soil and Tillage Research* **238**: 1-12.
- Gohil P, Gohil M, Rajatiya J, Halepotara F, Solanki M and Malam VR 2018. Role of potting media for ornamental pot plants. *International Journal of Pure Applied Biosciences* 6(1): 1219-1224.

- Grace OM 2019. Succulent plant diversity as natural capital. *Plants People Planet* 1: 336-345.
- Guddad M, Singh A, Shah HP, Chaudhari P and Ahlawat TR 2022. Effect of foliar application of chemicals on plant architecture in potted *Ixora chinensis* var. 'Mini Double'. *Current Journal of Applied Science and Technology* **41**(32): 9-15.
- Hartmann HT, Kester DE, Davies FT and Geneve RL 1997. *Plant propagation: principles and practices 7th Edition,* Prentice-Hall, Englewood Cliffs, N J, USA, p880.
- Herrera A 2009. Crassulacean acid metabolism and fitness under water deficit stress: If not for carbon gain, what is facultative CAM good for? *Annals of Botany* **103**: 645-653.
- Kapadiya DB, Singh A, Bhandari AJ, Patel AI and Patel K 2017. Development of in vivo plant propagation protocol in Euphorbia milii var. 'Pink Bold Beauty'. *International Journal of Current Microbiology and Applied Science* 6(12): 141-149.
- Kaur A, Dubey RK and Singh S 2015. Effect of different potting media on growth and flowering of kalanchoe (Kalanchoe blossfeldiana). *Indian Journal of Horticulture* **72**(3): 388-391.
- Kavipriya MV, Sankari A and Jegadeswari D 2019. Studies on the effect of alternate media on growth of Dracaena reflexa variegata. *International Journal of Current Microbiology and Applied Science* **8**(2): 3394-3400.
- Khayyat M, Nazari F and Salehi H 2007. Effects of different pot mixtures on Pothos (Epipremnum aureum Lindl. and 'Golden Pothos') growth and development. *American-Eurasian Journal* of Agricultural and Environmental Sciences **2**(4): 341-348.
- Larsen FE and Guse WE 1997. Propagating deciduous and evergreen shrubs, trees and vines with stem cuttings, A Pacific Northwest Cooperative Extension Publication, Washington, USA, p10.
- Lee JS 2010. Stomatal opening mechanism of CAM plants. *Journal* of *Plant Biology*. **53**:19-20.
- Nefzaoui A 2007. Cactus to improve livestock feeding and income

Received 11 June, 2024; Accepted 11 September, 2024

sources of the rural poor. Role of the FAO Cactusnet, pp 301-302. In: Priolo A, Biondi L, Ben Salem H, Mor and Fehr P (Eds). *Advanced nutrition and feeding strategies to improve sheep and goat.* Zaragoza, CIHEAM. Spain.

- Rashidha CK, Sankar M, Sreelatha U, Anupama TV and Prameela P 2021. Standardization of soilless growth media for raising potted ornamental foliage plants for export purpose. *Journal of Tropical Agriculture* **59** (1): 118-123.
- Sheoran OP, Tonk DS, Kaushik LS, Hasija RC and Pannu RS 1998. Statistical Software Package for Agricultural Research Workers, pp 139-143. In: Hooda DS and Hasija RC (Eds). Recent Advances in information theory, Statistics & Computer Applications. Department of Mathematics Statistics, CCS HAU, Hisar.
- Sindhuja M, Singh A, Bhandari AJ, Shah HP, Patel AI and Parekh VB 2022. Morphological characterization of different genotypes of adenium. *Indian Journal of Horticulture* **79**(3): 296-304.
- Singh A 2020. Indoor Gardening for Clean Air. *Nursery Today*, July-October: 39-42.
- Singh A, Ahlawat TR, Chavan S and Patel Al 2016. Wonder World of Adeniums. *Floriculture Today* 20(9): 44-49.
- Singh A, Bhandari AJ, Chavan S, Patel NB, Patel AI and Patel BN 2017. Evaluation of Adenium obesum for potted ornamentals under growing system. *International Journal of Current Microbiology and Applied Sciences* **10**(12): 2141-2146.
- Song CY, Lee SD, Park IT and Cho CH 2007. Effect of media and planting depth on growth of cacti and succulents in a pot. *Korean journal of horticultural science and technology* **25**(4): 429-435.
- Lodhi V, Bahadur V and Topno SE 2021. Influence of potting media on growth of succulents under shade net condition. *The Pharma Innovation Journal* SP-**10**(11): 101-104.
- Wazir MG, Amin NU, Khan I and Khan MI 2004. Effect of different potting mixtures and nitrogen source on the performance of Brassaia seedling-II. Sarhad Journal Agriculture 20(1): 25-31.



Manuscript Number: 4359 NAAS Rating: 5.38

# Assessment of Roadside Ornamental Trees Potential For Mitigating Heavy Metal Pollution in Ludhiana, Punjab

J. Verma, P. Singh and R. Singh

Department of Floriculture & Landscaping, Punjab Agriculture University, Ludhiana-141 004, India E-mail: jayothiverma@gmail.com

**Abstract:** This study examined heavy metal (HM) concentrations in roadside soil and ornamental tree leaves to identify trees with high bioaccumulation potential for mitigating HM pollution. Samples were collected from a highway site and PAU university campus, focusing on several heavy metals. The analysis revealed notable bioaccumulation factors (Bf) for chromium (Cr), nickel (Ni), copper (Cu), and lead (Pb) in the leaves of various tree species. Specifically, *Heterophragma adenophyllum* demonstrated strong affinity for zinc (Zn) and cadmium (Cd), while *Cassia siamea* showed significant uptake of Cu and Zn. These findings highlight the potential of certain tree species to help address heavy metal contamination in urban areas.

Keywords: Heavy metals (HM's), Ornamental trees, Bioaccumulation factor (Bf), Pollution, Affinity

Air pollution, primarily caused by industrialization and vehicular emissions, remains a significant global concern (Latif et al 2018). Among its components, heavy metals (HMs) pose direct and indirect risks to human health, exacerbated by anthropogenic activities such as industrial growth and increasing vehicular traffic in urban areas (Ugolini et al 2013, Hu et al 2014). Certain HMs, including arsenic, cadmium, chromium, mercury, and lead, are particularly hazardous to both plant and human health (Szyczewski et al 2009), while others like copper, selenium, and zinc are essential for plant growth at lower concentrations (Wuana and Okieimen 2011, Rehman et al 2018). The non-biodegradable nature of HMs leads to their accumulation in soil and the human body, posing long-term environmental and health risks (Nagajyoti et al 2010).

Vehicular emissions significantly contribute to elevated HM levels along urban roadside environments (Chen et al 2016, Wang et al 2022). Components of vehicles such as iron, copper, and zinc, present in alloys, tires, pipes, and wires, contribute to HM emissions through mechanical wear and tear over time (Ozaki et al 2004). This accumulation is concerning due to the persistence of HMs in the environment. Given the global urgency of this issue, numerous studies have investigated HM contamination along roadsides (Hu et al 2014, Mihailovic et al 2015). Plants, particularly trees, play a crucial role as bio-monitors by absorbing HMs from soil through their roots or from the air via stomata(Hosseini et al 2020, Kord et al 2010). Thereby, plants accumulate HM 's in their different body parts process known as phytoremediation, which is economical, sustainable, and eco-friendly method to protect environment (Hosseini et al 2020).Previous research has demonstrated the effectiveness of various plant species in monitoring HM impacts on their surrounding environments (Anjum et al 2021, Onder and Dursun 2006). Trees, with their extensive leaf and root surface areas, are particularly effective in this regard (Tomasevic et al 2008, Estrabou et al 2011, Li et al 2022). This study focuses on evaluating HM concentrations in selected roadside ornamental trees and soil, aiming to contribute to our understanding of HM distribution and bioaccumulation in urban environments.

#### MATERIAL AND METHODS

The study was conducted in Ludhiana, which experiences a semi-arid climate characterized by three distinct seasons: summer, monsoon, and winter. The city's average annual precipitation is 809.3 mm. The study area specifically focused on National Highway 5, and lies between the North latitude of 30°46'38" and the east longitude of 76°33'23". This highway, known for high vehicular traffic and associated emissions, was selected as the polluted site (study area). As a contrast, Punjab Agricultural University campus served as the control site. The polluted conditions of Ludhiana, stemming from intensive urbanization and industrialization, provided a suitable context for investigating pollution-related phenomena in this study.

**Sampling of plant and soil material:** The soil and leaf samples of selected tree species (*Acacia auriculiformis, Alstonia scholaris, Cassia fistula, Cassia siamea, Chukrasia tabularis, Dalbergia sissoo, Heterophragma adenophyllum, and Putranjiva roxburghii*) were collected from both sites (0–1 m from the road edge). Control soil and plant leaf



samples were collected from PAU, Ludhiana (100 m away from the road). Soil samples were collected 0, 10, 15, and 20 cm deep from each site along with both sites of road, and five representative samples of each site were prepared by the quadrant method (Rosa et al 2018). For accurate heavy metal analysis, leaf samples from five trees of each species, were harvested from each site during the mid-growing season, to ensure full leaf development (fully mature leaf) and reliable data on metal accumulation. The permissible limits for heavy metals in soil and plant leaves according to different standards, presented in Table 1.

Soil and plant samples analysis: The heavy metal content in soil samples was assessed using the method developed by Lindsay and Norvel (1978). Concentrations of heavy metals (Cr, Fe, Ni, Cu, Zn, Cd, and Pb) were determined using an ICP Spectrometer (ICP-MS 7700 Series). Similarly, heavy metal concentrations in leaf samples were analyzed using an ICP Spectrometer (Model iCAP 6000 Series). The heavy metals (Cr, Fe, Ni, Cu, Zn, Cd and Pb) were estimated by injecting filtered samples into Inductively Coupled Plasma Emission spectrophotometer and amount expressed in mg kg<sup>-1</sup>(Ammann 2007). Bioaccumulation factor (Bf) is the plant potential to take up HMs (or any other chemical) from the soil/growth media and it can be calculated as a leaf to soil ratio of the HM concentration (Alahabadiet al 2017). Bf was worked out to determine HM accumulation potential of an individual tree species.

**Statistical analysis:** The data were analyzed using a factorial randomized block design. and comparison was performed by Tukey's test at a 5% significant level using SAS software computer version 9.2.

#### **RESULTS AND DISCUSSION**

Heavy metal concentrations in soil: The concentrations of heavy metals (chromium, iron, nickel, copper, zinc, cadmium, and lead) showed significant differences between two locations (Table 2). The content of metals (Cr, Fe, Ni, Cu, Zn, Cd, and Pb) in the soil across two sites exhibited significant variability. All examined metals were elevated in roadside soils compared to the control site soil. HM concentrations between control and polluted site soil ranged from 1.98 to 18.74mg kg<sup>-1</sup> for Cr, 993.77 to 11947.86 mg kg<sup>-1</sup> for Fe, 1.54 to 14.66 mg kg<sup>-1</sup> for Ni, 3.09 to 19.61mg kg<sup>-1</sup> for Cu, 9.12 to 68.16mg kg<sup>-1</sup> for Zn, 0.49 to 0.55mg kg<sup>-1</sup> for Cdand 3.01 to 16.78mg kg<sup>-1</sup> for Pb. Levels of all metals were under permissible limits in both the locations. Despite being in within permissible limits, the elevated concentrations suggest that the polluted site is experiencing environmental stress, possibly due to industrial activities or pollution sources.

These findings are consistent with previous studies that have documented increasing HM (Fe, Ni, Cr, Zn, Cu, Cd and Pb) concentrations in polluted environments (He and Yang 2015). The high levels of heavy metals can have detrimental effects on soil health, plant growth, and overall ecosystem stability (Telo da Gama 2023). The highway site soil exhibited higher concentrations of heavy metals compared to the campus site, attributed to its high traffic volume (Elbagermi et al 2012, Wang and Zhang 2018). The detrimental effects of various heavy metals on both plants and humans, highlighting the need for effective management and mitigation strategies to address heavy metal contamination in Table 3.

Heavy metal concentrations in selected tree leaves: The concentration of heavy metals shows a significant increase of approximately 74.5% in polluted areas (106.52 mg kg<sup>-1</sup>) compared to control sites (61.04 mg kg<sup>-1</sup>) (Table 4). In comparing heavy metal concentrations between polluted and control sites, *Alstonia scholaris* shows the greatest increase at polluted site 119.43%, while *Cassia siamea* has the smallest increase at 39.45%, respectively. This trend indicates that all species significantly accumulate more heavy metals in polluted areas compared to control sites suggesting that the variations in HM levels are due to the impact of vehicular traffic (Ogutucu et al 2021). The percentage increases in heavy metal concentrations from

control to polluted sites, ranked from highest to lowest, highlight several significant trends. Cu shows the most dramatic rise, with increase of 5,875%, and Cr shows the smallest increase at 20.2%. . Levels of Zn, Ni and Pb surpass permissible limits, at both sites in all tree species suggesting notable pollution, conversely, Cu, Cd, and Cr are generally within acceptable ranges, though some species show elevated levels, particularly in polluted sites. Pb is a significant pollutant from auto exhaust (Muzychenko et al 2017). Ni in roadside plant may originate from engine oil, fuel, tire and brake wear, and the corrosion of nickel alloy components (Gupta 2020, Khalid 2018). Zn is commonly associated with roadside plants due to its extensive use in construction and transportation (Barber et al 2017). Overall roadside plants, being exposed to heightened anthropogenic activities, may exhibit increased accumulation of heavy metals in their leaves (Ogutucuet al 2021).

The study demonstrates that different tree species have varying capacities for accumulating heavy metals from the environment as it depends on leaf surface area quality, stomata size, and HM's particle size (Lei et al 2006, Ugolini et al 2013, Singh et al 2017). *Alstonia scholaris* is the most effective heavy metal accumulator, making it ideal for phytoremediation, while *Cassia fistula* and *Cassia siamea* show the least affinity for heavy metals. The study also highlighted differential affinity of selected roadside trees towards these heavy metals, suggesting their potential as bio-indicators. *Alstonia scholaris* exhibit highest

 Table 2. Heavy metal contents (mg kg<sup>-1</sup>) in soil samples from control and polluted locations

Heavy metal	Control site	Polluted site	Percent increase in polluted location (%)
Chromium (Cr)	1.98⁵	18.74ª	846.46
Iron (Fe)	993.77 <sup>b</sup>	11947.86ª	1102.28
Nickel (Ni)	1.48 <sup>♭</sup>	14.66ª	890.54
Copper (Cu)	3.09 <sup>b</sup>	19.61ª	534.63
Zinc (Zn)	9.12⁵	68.16ª	647.37
Cadmium (Cd)	0.49 <sup>b</sup>	0.55ª	12.24
Lead (Pb)	3.01 <sup>⁵</sup>	16.78ª	457.48
Overall location (Mean)	144.71 <sup>₅</sup>	1726.62ª	-

\* Different letters in each row are significantly different at P≤0.05 by Turkey's Test

Table 1. Acceptable limits of heavy metals (HM's) in soil and plant leaf samples

Sample	Standards	Cd	Cu	Pb	Zn	Ni	Cr
Soil (mg kg <sup>-1</sup> )	Indian Standard (Awashthi 2000)	3-6	135-270	250-500	300-600	75-150	-
	WHO(2006)	-	140	84	-	107	-
	European Union Standards (EU 2006)	3.0	100	300	300	50	100
Plant (mg kg <sup>-1</sup> )	Indian Standard (Awashthi 2000)	1.5	30.0	2.5	50.0	1.5	20.0
	WHO/FAO (2007)	0.2	40.0	5.0	60.0	-	-
	European Union Standards (EU 2006)	0.2	-	0.30	-	-	-

Heavy metal	Effects on plants	Effects on humans	References
Pb	<ul> <li>Reduces growth and chlorophyll content</li> <li>Causes leaf and root damage</li> <li>Interferes with nutrient uptake</li> </ul>	<ul> <li>Neurotoxicity, especially in children</li> <li>Kidney damage</li> <li>Developmental and cognitive impairments</li> </ul>	Alloway 2013, Tchounwou et al 2012, Alengebawy2021
Cd	<ul> <li>Inhibits seed germination and growth</li> <li>Causes leaf chlorosis and necrosis</li> <li>Interferes with enzyme activity</li> </ul>	<ul> <li>Causes kidney damage and osteoporosis</li> <li>Carcinogenic effects</li> <li>Impaired respiratory function</li> </ul>	Alloway 2013, Jarup 2003, Satarug et al 2011, Haider et al 2021
Cr	<ul> <li>Affects photosynthesis and growth</li> <li>Causes root and shoot damage</li> <li>Alters enzyme activity</li> </ul>	<ul> <li>Causes respiratory issues and skin ulcers</li> <li>Impairment of liver and kidney function</li> <li>Carcinogenic</li> </ul>	Kapoor et al 2022
Ni	<ul> <li>Causes reduced seed germination</li> <li>Inhibits root growth and development</li> <li>Alters photosynthetic activity</li> </ul>	<ul> <li>Respiratory issues and dermatitis</li> <li>Nausea, vomiting, and diarrhoea</li> <li>Potentially carcinogenic</li> </ul>	Hassan et al 2019, Genchietal2020
Cu	<ul> <li>Causes leaf necrosis and chlorosis</li> <li>Inhibits root growth</li> <li>Impairs nutrient absorption</li> </ul>	<ul> <li>Causes gastrointestinal distress</li> <li>Liver and kidney damage</li> <li>Neurological problems</li> </ul>	Balali-Mood et al 2021,Miretal2021
Zn	<ul> <li>Leads to leaf chlorosis and stunted growth</li> <li>Affects root and shoot development</li> <li>Interferes with enzyme functions</li> </ul>	<ul> <li>h Causes gastrointestinal issues</li> <li>Impaired immune function</li> <li>Possible disruption of growth and development</li> </ul>	Ali et al 2013 t

Table 4. Metal concentration (mg kg<sup>-1</sup>) in the leaf of selected roadside tree species growing at the control and polluted locations

Tree species	Location	Heavy metals (mg kg <sup>-1</sup> )					Mean	Mean		
		Fe	Cu	Zn	Cd	Cr	Ni	Pb	(Location)	(Specie)
Acacia	Control	340.11 <sup>m</sup>	1.62 <sup>°</sup>	21.23 <sup>°</sup>	0.05 <sup>g</sup>	16.30 <sup>i</sup>	5.93 <sup>i</sup>	5.55 <sup>ef</sup>	55.83 <sup>⊳</sup>	78.96°
auriculiformis	Polluted	639.83 <sup>ª</sup>	6.36 <sup>f</sup>	37.93 <sup>f</sup>	0.14°	17.62°	6.41°	6.37°	102.09ª	
Alstonia scholaris	Control	422.70 <sup>i</sup>	4.93 <sup>i</sup>	28.26 <sup>1</sup>	$0.10^{\text{de}}$	17.22 <sup>h</sup>	6.22 <sup>h</sup>	5.78 <sup>de</sup>	69.32 <sup>b</sup>	110.74ª
	Polluted	968.80ª	10.88 <sup>₫</sup>	52.34 <sup>b</sup>	0.22ª	18.35 <sup>⊳</sup>	6.88 <sup>b</sup>	7.65°	152.16°	
Chukrasia	Control	336.23 <sup>n</sup>	2.65 <sup>m</sup>	23.45 <sup>m</sup>	0.08 <sup>ef</sup>	17.65°	6.34 <sup>f</sup>	5.31 <sup>f</sup>	55.96 <sup>₅</sup>	75.85 <sup>f</sup>
tabularis	Polluted	585.70 <sup>f</sup>	5.86 <sup>°</sup>	47.79°	0.09 <sup>ef</sup>	17.76₫	6.67°	6.27°	95.73ª	
Cassia fistula	Control	321.60°	4.64 <sup>i</sup>	18.81 <sup>°</sup>	0.09 <sup>e</sup>	17.30 <sup>g</sup>	5.97 <sup>i</sup>	4.80 <sup>g</sup>	53.32⁵	73.21 <sup>°</sup>
	Polluted	573.83 <sup>9</sup>	10.14°	37.44 <sup>9</sup>	0.17 <sup>♭</sup>	17.49 <sup>f</sup>	6.42°	6.16d°	93.09ª	
Cassia siamea	Control	319.47°	3.34	20.45°	0.09 <sup>e</sup>	17.30 <sup>g</sup>	6.27 <sup>9</sup>	5.33 <sup>f</sup>	53.18 <sup>₅</sup>	63.68 <sup>h</sup>
	Polluted	437.73 <sup>h</sup>	14.22 <sup>⊳</sup>	36.33 <sup>h</sup>	$0.12^{\text{dc}}$	17.83⁴	6.85 <sup>b</sup>	6.20 <sup>dc</sup>	74.18ª	
Dalbergia sissoo	Control	375.51	0.24°	29.67 <sup>×</sup>	$0.10^{\text{de}}$	17.34 <sup>9</sup>	6.25 <sup>gh</sup>	5.81 <sup>de</sup>	62.13 <sup>⁵</sup>	81.08 <sup>d</sup>
	Polluted	607.58°	14.33ª	46.49°	0.17 <sup>⁵</sup>	18.05°	6.49 <sup>d</sup>	7.15⁵	100.04ª	
Heterophragma	Control	426.57	3.94 <sup>k</sup>	35.09 <sup>i</sup>	0.08 <sup>ef</sup>	17.61 <sup>e</sup>	6.29 <sup>g</sup>	5.50 <sup>ef</sup>	70.73 <sup>b</sup>	97.68 <sup>b</sup>
adenophyllum	Polluted	762.61 <sup>⁵</sup>	11.14°	66.31ª	0.29ª	18.34 <sup>b</sup>	6.67°	7.10⁵	124.64ª	
Putranjiva	Control	409.19 <sup>k</sup>	5.10 <sup>h</sup>	31.15 <sup>i</sup>	0.06 <sup>fg</sup>	17.60°	6.28 <sup>9</sup>	5.81 <sup>de</sup>	67.88 <sup>b</sup>	89.04°
roxburghii	Polluted	675.38°	14.32ª	46.70 <sup>d</sup>	0.12 <sup>de</sup>	19.61ª	7.56ª	7.61ª	110.19ª	
Overall Mean	Control								61.04 <sup>b</sup>	
	Polluted								106.52ª	

accumulation of Fe and Pb as compare to other species under study. Similarly, Dalbergia sissoo for (Cu), Heterophragma adenophyllum for (Zn & Cd) and Putranjiva roxburghii showed amassing for (Cr & Ni). The absorption ability of a plant species changes with the leaf properties of the plant as well as with plant species (Shahid et al 2017). These findings can guide the selection of tree species for environmental cleanup and management of heavy metal pollution. The bioaccumulation factor of the HMs in the selected roadside tree leaves varies with site and tree species (Patel et al 2015, Greksa et al 2019). Among the tree species studied, the overall Bioaccumulation Factor (Bf) for trees, follow the order: Putranjiva roxburghii (0.438) >Alstonia scholaris (0.433) >Heterophragma adenophyllum (0.396) >Acacia auriculiformis (0.362) >Cassia siamea (0.337) = Cassia fistula(0.334) > Chukrasia tabularis (0.316)>Dalbergia sissoo (0.272). Species with higher Bf values like Putranjiva roxburghii, Alstonia scholaris and Heterophragma adenophyllum are more adept at accumulating heavy metals and are therefore preferable for phytoremediation in highly contaminated soils.

## CONCLUSION

The study highlights the variable capacity of different tree species to accumulate heavy metals. Alstonia scholaris

emerged as the most effective accumulator, particularly in polluted areas, along with others such as Dalbergia sissoo, Heterophragm aadenophyllum, and Putranjiva roxburghii, which demonstrates significant potential for use in phytoremediation efforts. The capacity for heavy metal accumulation varies with leaf properties and species characteristics, supporting the need for tailored approaches in environmental management. Mostly HMs under consideration was within permissible limits, but the substantial exceedance in Zn, Ni and Pb levels in plant leaves underscore specific contamination concerns. HM accumulation is a major health concern globally. In Ludhiana, high traffic and industrial activities contribute to Pb buildup in roadside plants, raising health risks and highlighting a pressing issue in India. Continued monitoring, along with the use of appropriate tree species for phytoremediation, will be essential for mitigating the impacts of heavy metal pollution and improving soil and plant health.

#### REFERENCES

- Alahabadi A, Ehrampoush MH, Miri M, Aval HE, Yousefzadeh S, Ghaffari HR Ahmadi E, Talebi P, Fathabadi ZA, Babai F and Nikoonahad A 2017. A comparative study on the capability of different tree species in accumulating heavy metals from soil and ambient air. *Chemosphere* **172**: 459-467.
- Alengebawy A, Abdelkhalek ST, Qureshi SR and Wang MQ 2021. Heavy metals and pesticides toxicity in agricultural soil and

plants: Ecological risks and human health implications. Toxics  ${f 9}(3)$ : 42.

- Ali H, Khan E and Sajad MA 2013. Phytoremediation of heavy metals-concepts and applications. *Chemosphere* 91(7): 869-881.
- Alloway BJ 2013. *Heavy metals in soils: Trace metals and metalloids in soils and their bioavailability* (3rd ed.), Dordrecht: Springer Netherlands.
- Ammann AA 2007. Inductively coupled plasma mass spectrometry (ICP MS): A versatile tool. *Journal of Mass Spectrometry* 42(4): 419-27.
- Anjum S, Hussain M, Hameed M and Ahmad R 2021. Physiological, biochemical and defense system responses of roadside vegetation to auto-exhaust pollution. *Bulletin of Environmental Contamination and Toxicology* **107**(5): 946-954.
- Awashthi SK 2000. *Prevention of Food AdulterationAct no* 37 of 1954. Central and State Rules as Amended for 1999, Ashoka Law House, New Delhi.
- Balali-Mood M, Naseri K, Tahergorabi Z, Khazdair MR and Sadeghi M 2021. Toxic mechanisms of five heavy metals: mercury, lead, chromium, cadmium, and arsenic. *Frontiers in Pharmacology* 12: 643972.
- Chen JP, Wang LK, Wang MHS, Hung YT and Shammas NK 2016. *Remediation of Heavy Metals in the Environment* New York: CRC Press, Taylor & Francis Inc, p 131.
- Elbagermi MA, Edwards HGM and Alajtal AI 2012. Monitoring of heavy metal content in fruits and vegetables collected from production and market sites in the Misurata area of Libya. *International Scholarly Research Notices* **2012**(1): 827645.
- Estrabou C, Filippini E, Soria JP, Schelotto G and Rodriguez JM 2011. Air quality monitoring system using lichens as bioindicators in Central Argentina. *Environmental Monitoring and Assessment* **82**: 375-383.
- European Union 2006. Commission Regulation (EC) No. 1881/2006 of 20 December 2006 setting maximum levels for certain contaminants in soil and foodstuffs. *Official Journal of the European Union L* **364**: 5-24.
- Genchi G, Carocci A, Lauria G, Sinicropi MS and Catalano A 2020. Nickel: Human health and environmental toxicology. International Journal of Environmental Research and Public Health **17**(3): 679.
- Greksa A, Ljevnaic-Masic B, Grabic P, Benka V, Radonic B, Blagojevic B and Sekulic M 2019. Potential of urban trees for mitigating heavy metal pollution in the city of Novi Sad, Serbia. *Environmental Monitoring and Assessment* **191**(10): 636.
- Gupta V 2020 Vehicle-generated heavy metal pollution in an urban environment and its distribution into various environmental components. In *Environmental Concerns and Sustainable Development* (Vol. 1), *Air, Water and Energy Resources*, pp 113-127.
- Haider FU, Liqun C, Coulter JA, Cheema SA, Wu J, Zhang R and Farooq M 2021. Cadmium toxicity in plants: Impacts and remediation strategies. *Ecotoxicology and Environmental Safety* 2(11): 111887.
- Hassan MU, Chattha MU, Khan I, Chattha MB, Aamer M, Nawaz M and Khan TA 2019. Nickel toxicity in plants: reasons, toxic effects, tolerance mechanisms, and remediation possibilities: A review. *Environmental Science and Pollution Research* **26**: 12673-688.
- He Z and Yang X 2015. Heavy metal contamination of soils: Sources, indicators, and assessment. *Journal of Environmental Indicators* **9**:17-19.
- Hosseini NS, Sobhanardakani S and Cheraghi M 2020. Heavy metal concentrations in roadside plants (*Achillea wilhelmsii* and *Cardaria draba*) and soils along some highways in Hamedan, west of Iran. *Environmental Science and Pollution Research* **27**:13301-13014.
- Hu Y, Wang D, Wei L, Zhang X and Song B 2014. Bioaccumulation of

heavy metals in plant leaves from Yan'an city of the Loess Plateau, China. *Ecotoxicology and Environmental Safety* **110**: 82-88.

- Jarup L 2003. Hazards of heavy metal contamination. *British Medical* Bulletin 68(1): 167-182.
- Kapoor RT, Mfarrej MFB, Alam P, Rinklebe J and Ahmad P 2022. Accumulation of chromium in plants and its repercussion in animals and humans. *Environmental Pollution* **301**: 119044.
- Khalid N et al 2018. Effects of road proximity on heavy metal concentrations in soils and common roadside plants in SouthernCalifornia. *Environmental Science and Pollution Research* **25**: 35257-35265.
- Barber LB, Paschke SS, Battaglin WA, Douville C, Fitzgerald KC, Keefe SH, Roth DA and Vajda AM 2017. Effects of an extreme flood on trace elements in River water from urban stream to major River Basin. *Environmental Science & Technology* **51**(18): 10344-10356.
- Kord B, Mataji A and Babaie S 2010. Pine (*Pinus eldarica* Medw.) needles as indicators for heavy metals pollution. *International Journal of Environmental Science and Technology* 7(1): 79-84.
- Latif MT, Othman M, Idris N, Juneng L, Abdullah AM, Hamzah WP and Jaafar AB 2018. Impact of regional haze towards air quality in Malaysia: Areview. *Atmospheric Environment* **177**: 28-44.
- Lei WANG, Liu LY, Gao SY, Eerdun HASI and Zhi WANG 2006. Physicochemical characteristics of ambient particles settling upon leaf surfaces of urban plants in Beijing. *Journal of Environmental Sciences* 18(5): 921-926.
- Li Y, Xu Z, Ren H, Wang D, Wang J, Wu Z and Cai P 2022. Spatial distribution and source apportionment of heavy metals in the Topsoil of Weifang City, East China. *Frontiers in Environmental Science* **10**: 893938.
- Lindsay WL and Norvell WA 1978. Development of DTPA soil test for zinc, iron, manganese and copper. *Soil Science Society of America Journal* **42**:421-428.
- Mihailovic A, Budinski-Petkovic L, Popov S, Ninkov J, Vasin J, Ralevic NM and Vucinic-Vasic M 2015. Spatial distribution of metals in urban soil of Novi Sad, Serbia: GIS-based approach. *Journal of Geochemical Exploration* **150**: 104-114.
- Mir AR, Pichtel J and Hayat S 2021. Copper: uptake, toxicity and tolerance in plants and management of Cu-contaminated soil. *Biometals* 34(4): 737-59.
- Muzychenko I, Jamalova G, Mussina U, Kazulis V and Blumberga D 2017. Case study of lead pollution in the roads of Almaty. *Energy Procedia* **113**: 369-376.
- Nagajyoti PC, Lee KD and Sreekanth TVM 2010. Heavy metals, occurrence and toxicity for plants: A review. *Environmental Chemistry Letters* **8**(3): 199-216.
- Ogutucu G, Ozdemir G, Acararıcın Z and Aydın A 2021. Trend analysis of lead content in roadside plant and soil samples in Turkey. *Turkish Journal of Pharmaceutical Science* **18**(5): 581-588.
- Onder S and Dursun S 2006. Air borne heavy metal pollution of *Cedruslibani* (A. Rich.) in city center of Konya (Turkey). *Atmospheric Environment* **40**(6): 1122-1133.
- Ozaki H, Watanabe I and Kuno K 2004. Investigation of the heavy metal sources in relation to automobiles. *Water, Air, and Soil Pollution* **157**: 209-223.
- Patel K, Sharma R, Dahariya N, Yadav A, Blazhev B, Matini L and Hoinkis J 2015. Heavy metal contamination of tree leaves. *American Journal of Analytical Chemistry* **6**:687-93.
- Rehman K, Fatima F, Waheed I and Akash MSH 2018. Prevalence of exposure of heavy metals and their impact on health consequences. *Journal of Cellular Biochemistry* **119**(1): 157-184.
- Rosa DD, Tres G, Broti-Rissato B, Lorenzetti E, Guimaraes VF and Feiden A 2018. Rooting Platanus (*Platanus acerifolia* (Aiton) Willd.) cuttings in Marechal Cândido Rondon-PR, razil: Influence

of lesions at cutting bases and depth of planting. Acta Agronomica 67(1): 109-113.

- Satarug S, Garrett SH, Sens MA and Sens DA 2011. Cadmium, environmental exposure, and health outcomes. *Ciencia &Saudecoletiva* **16**(5): 2587-2602.
- Shahid M, Dumat C, Khalid S, Schreck E, Xiong T and Niazi NK 2017. Foliar heavy metal uptake, toxicity and detoxification in plants: A comparison of foliar and root metal uptake. *Journal of Hazardous Materials* **325**: 36-58.
- Singh H, Savita, Sharma R, Sinha S, Kumar M, Kumar P, Verma A and Sharma SK 2017. Physiological functioning of *Lagerstroemia speciosa* L. under heavy roadside traffic: An approach to screen potential species for abatement of urban air pollution. *Biotechonology* 7(61): 1-10.
- Szyczewski P, Siepak J, Niedzielski P and Sobczynski T 2009. Research on heavy metals in Poland. *Polish Journal of Environmental Studies* **18**(5): 755-768.
- Tchounwou PB, Yedjou CG, Patlolla AK and Sutton DJ 2012. Heavy metal toxicity and the environment. Molecular, clinical and environmental toxicology. *Environmental Toxicology* **3**: 133-164.
- Telo da Gama J 2023. The Role of Soils in Sustainability, Climate Change, and Ecosystem Services: Challenges and Opportunities. *Ecologies* **4**: 552-567.

Tomasevic M, Vukmirovic Z, Rajsic S, Tasic M and Stevanovic B

Received 08 June, 2024; Accepted 21 September, 2024

2008. Contribution to biomonitoring of some trace metals by deciduous tree leaves in urban areas. *Environmental Monitoring and Assessment* **137**: 393-401.

- Ugolini F, Tognetti R, Raschi A and Bacci L 2013. Quercus ilex L. as bio-accumulator for heavy metals in urban areas: effectiveness of leaf washing with distilled water and considerations on the tree's distance from traffic. *Urban Forestry & Urban Greening* **12**(4): 576-584.
- Wang A, Guo Y, Fang Y and Lu K 2022. Research on the horizontal reduction effect of urban roadside green belt on atmospheric particulate matter in a semi-arid area. *Urban Forestry & Urban Greening* **68**: 127449.
- Wang M and Zhang H 2018. Accumulation of heavy metals in roadside soil in urban area and the related impacting factors. International Journal of Environmental Research and Public Health **15**(6): 1064.
- WHO/FAO 2007. Joint FAO/WHO Food Standard Programme Codex Alimentarius Commission 13th Session Report of the Thirty-Eight Session of the Codex Committee onFood Hygiene. Houston, United States of America, ALINORM 07/30/13.
- Wuana RA and Okieimen FE 2011. Heavy metals in contaminated soils: A review of sources, chemistry, risks and best available strategies for remediation. *International Scholarly Research Notices* 2011(1):402647.



# Aquaponic Production of Ornamental Koi Carp (*Cyprinus carpio* Linn.) and Lettuce (*Lactuca sativa*) in Comparison with Traditional Fish Culture and Hydroponics System

Khushvir Singh, Vaneet Inder Kaur<sup>1</sup>, Meera D. Ansal, Dilpreet Talwar<sup>1</sup>, Abhishek Srivastava and Kulbir Singh<sup>1</sup>

Department of Aquaculture, College of Fisheries, Guru Angad Dev Veterinary and Animal Sciences University (GADVASU), Ludhiana-141 004, India <sup>1</sup>Department of Vegetable Science, College of Horticulture and Forestry Punjab Agriculture University, Ludhiana-141 004, India \*E-mail: vaneetinderkaur@gadvasu.in

**Abstract:** The study was conducted for comparative analysis of the production of ornamental fish, koi carp (*Cyprinus carpio* Linn.) and lettuce (*Lactuca sativa*) in aquaponics and traditional culture system. The experimental fish were stocked at three stocking densities, i.e.,  $20 \text{ m}^3$  (SD1),  $30 \text{ m}^3$  (SD2) and  $40 \text{ m}^3$  (SD3) in circular cemented tanks ( $10\text{ m}^3$ ) of the aquaculture unit and lettuce were planted (plant to plant spacing of 12 cm) in the hydroponics unit (HM) of automated aquaponics system. For comparison with the traditional system, koi carp were stocked in cemented tank of equal size ( $10\text{m}^3$ ) at a standard stocking density of  $2 \text{ m}^3$  and lettuce was planted as per standard protocol i.e. plant to plant space of 30 cm in outdoor conditions in soil bed traditional system (SM) The water quality parameters in both aquaponics and traditional tanks remained in optimum range for fish culture along with >95% of nutrient removal by lettuce. The survival (%) of fish was maximum in control (93.33%) followed by SD3, SD1 and SD2. The final total body length and body weight was significantly higher in control, but the average fish yield (kg m<sup>-3</sup>) was significantly higher in all aquaponic tanks, with the maximum yield of 3.73 kg m<sup>-3</sup> (9.56 times) in SD3 ( $40 \text{ m}^3$ ). The lettuce yield (32.2 q acre<sup>-1</sup>) was 1.15 times higher in the hydroponic unit, with significantly higher pigment content and insignificant differences for nutrient content except fibre.

## Keywords: Aquaponics, Koi carp, Lettuce, Hydroponics, Leafy vegetables

The depletion of water resources and increased water pollution has become a major threat throughout the world, especially in developing countries, due to rapidly progressing agricultural, aquaculture and industrial activities. In addition to this, lack of adequate wastewater treatment further acts as a limiting factor for the development of all these sectors (Morrison et al 2009). Since the last two decades, aquaculture has made enormous contribution towards livelihood/employment generation and food/nutritional security around the world. The country's total fisheries production was 17.545 million metric tonnes (MMT) during 2022-23 with a contribution to the world fish production (Handbook on Fisheries Statistics 2023). With this growth, the aquaculture sector is being looked forward to fill the gap, as the marine resources are already being overexploited and are facing many ecological imbalances.

For taking up traditional aquaculture, a large volume of water and land is required, and scarcity/limited availability of these two resources are the major challenges (FAO 2022) for expansion of inland aquaculture. Additionally, surplus organic waste in the form of ammonia, nitrate and phosphate is being added in aquaculture waste water and finally in

natural water bodies leading to water pollution. Therefore, it is essential to manage the heavy nutrient load of aquaculture waste waters, so that receiving water bodies can be protected from eutrophication for potential reuse of water (Endut et al 2009). Further, intensification of aquaculture farming with more advent techniques along with diversification of economically important species is considered one of the best way to achieve the goal of sustainable development of aquaculture sector (FAO 2022). Among intensive aquaculture technologies, aquaponics, is a closed loop system, in which the wastewater from fish farming is purified by the plants and returned to the fish production system. Thus, the cost of purification of water is reduced and profit margin increases significantly as the excreta from farming fish and other aquatic organisms is used as fertilizer for plants in a hydroponic system (Sas-Paszt et al 2023). Moreover, this synergestic combination of two systems, i.e., aquaculture and hydroponics (soilless agriculture system) is a sustainable arrangement with limited water and land (10-12% as compared to traditional practices) requirement (Shete et al 2016).

Punjab, one of the progressive state in North-Western

India, produced about 1.85 lakh tonnes of fish production (major contribution from freshwater carps) during 2022-23 (Handbook on Fisheries Statistics 2023). New initiatives has been taken w.r.to standardization of intensive aquaculture technologies like re-circulatory aquaculture system (RAS), biofloc aquaculture system and aquaponics in agroclimatic conditions of the region. Preliminary trials were conducted during 2020-21 to explore the possibility of combining fish with different crops/vegetables in the said system. In line with this, the present study was executed to optimize the stocking density of koi carp, one of the widely cultured ornamental fish throughout the world including India, in combination with lettuce during the winter season.

### MATERIAL AND METHODS

**Experimental layout:** The experimental study (3 months) was conducted in an automated aquaponics unit (175 m<sup>2</sup>) with aquaculture unit having 3 tanks (75 m<sup>2</sup>) and hydroponic unit (100 m<sup>2</sup>) at the Instructional cum Research Farm, College of Fisheries, GADVASU, Ludhiana. Koi carp fingerlings were stocked at three stocking densities in aquaculture unit and lettuce was planted in hydroponics (HM) unit. For comparison with the traditional system, koi carp fingerlings were stocked in cemented tanks (triplicate) at a standard stocking rate and lettuce was planted in outdoor conditions in a traditional soil bed (SM) system in completely randomized design (Table 1).

Water quality: Water quality parameters viz.; Temperature, pH, Dissolved Oxygen (DO), total alkalinity (TA), total

hardness (TH), ammonia (NH<sub>3</sub>-N), nitrite (NO<sub>2</sub><sup>-</sup>-N), nitrate (NO<sub>3</sub><sup>-</sup>-N) and ortho-phosphate (PO<sub>4</sub><sup>3-</sup>-P) were analyzed as per standard methods of APHA (2012) every fortnight throughout the experimental period in both control and experimental tanks.

**Nutrient removal (%):** The nutrient removal (%) of water in terms of  $NH_3$ -N,  $NO_3^{-}$ -N and  $PO_4^{-3}$ -P in water collected from hydroponics unit was studied at fortnightly intervals.

**Fish growth:** Fish growth was observed at monthly intervals in terms of total body length, body weight and growth parameters in terms of average total length gain (TLG), average net weight gain (NWG), yield (Kg m<sup>3</sup>), specific growth rate (SGR), condition factor (K), feed conversion ratio (FCR) and survival (%) were calculated after the completion of experiment.

Lettuce growth, pigment analysis and nutrient composition: Lettuce growth was assessed in terms of total plant height and root length at monthly interval. The plant height increase (PHI), root length increase (RLI), yield (Kg m<sup>-2</sup>) and pigment content (chlorophyll a, chlorophyll b, total chlorophyll and carotenoids) calculated/analyzed after completion of experiment. Nutrient composition (%) in terms of protein, fat, ash, carbohydrates and fiber was analyzed (AOAC 2007).

*Statistical analysis:* The data was analyzed by software statistical package CPCS-1 (Cheema and Singh 1991).

# **RESULTS AND DISCUSSION**

**Water quality:** The water quality parameters viz. water temperature pH, DO, TA, TH,  $NH_3$ -N,  $NO_2^-$ -N,  $NO_3^-$ -N) and  $PO_4^{3}$ -P of control and experimental tanks remained in

**Table 1.** Details of experimental layout & design

Particulars	Details
Preparation of experimental & control tanks	• Before stocking of fish, experimental and control tanks were completely sun dried (10 days) for disinfection, which were further disinfected with limestone applied @ 300kg/ha. Tube well water was used for filling and maintaining the water in the experimental and control tanks. After one week of liming and filling of water, the water quality parameters of all the tanks were analyzed before stocking the fish.
Tank capacity & water volume	<ul> <li>Aquaponics Tanks Circular cemented &amp; Circular) – 11m<sup>3</sup>, with water volume of 10,000 liters</li> <li>Control tanks (Rectangular cemented &amp; rectangular – 10m<sup>3</sup>)</li> </ul>
Fish acclimatization	• 10-15 days under indoor conditions (fed Ad libitum with commercial diet)
Fish stocking density (SD)	<ul> <li>Experimental stocking densities (SD) – (3) i.e. 20m<sup>-3</sup> (SD1), 30m<sup>-3</sup> (SD2), 40m<sup>-3</sup> (SD3)</li> <li>Control (C) Stocking density - 2m<sup>-3</sup></li> <li>Average total body length 3 39+0.06 cm</li> </ul>
Fish Size	<ul> <li>Average total body length 3.32±0.06 g</li> <li>Av. body weight 3.32±0.06 g</li> </ul>
Fish feeding	• 5% of fish body weight, twice daily (10:00 and 16:00 h). Amount of feed was adjusted after each sampling (monthly) according to increase in fish weight
Crop plantation	<ul> <li>Plantation of Lettuce (<i>Lactuca sativa</i>) procured from local nursery of Ludhiana following completely randomized design.</li> <li>Plant to plant space - 12 cm, planted in eight stands [9 pipes having 29 cups each (24 cups planted) in one stand] in hydroponics (HM) unit - (Vertical Farming). Water from aquaculture units was circulated into the hydroponic unit at fixed intervals through a biological filter, resulting in nutrient uptake by the lettuce plants</li> <li>Plant to plant space - 30 cm, planted in 100 m<sup>2</sup> plot area with 8 beds in traditional soil bed (SM) system - (horizontal farming). Standard management practices were followed</li> </ul>

optimum range (Table 2) for koi carp (*C. carpio*) with significant higher pH and significant lower values for NH<sub>3</sub>-N, NO<sub>3</sub><sup>-</sup>-N and PO<sub>4</sub><sup>3</sup>-P in control as compared to all the treatments. The values for water quality parameters remained in optimum range for fish culture and are in agreement with the previous studies by different researchers (Da Silva Cerozi and Fitzsimmons 2016, Nuwansi et al 2017 and Nuwansi et al 2020).

**Nutrient removal (%):** During experimental period, the nutrient removal (%) in terms of  $NH_3$ -N,  $NO_3$ -N and  $PO_4^3$ -P (Table 3) revealed that with the progress of experiment, nutrient removal (%) increased with maximum removal of  $NH_3$  N (98.22%) followed by  $NO_3$ -N (97.53%) and  $PO_4^3$ -P (96.41%). Lin et al (2002) reported 86-98% removal of ammonium nitrogen ( $NH_4^+$ -N) from constructed wetlands system receiving aquaculture waste water. Endut et al (2009) found total phosphorus removal rates of 43 to 53% in aquaponic systems, lesser than observed in present experiment.

**Fish survival and growth:** Fish survival (%) in C, SD1, SD2 and SD3 after completion of experiment was 93.33, 82.00, 78.33 and 83.25, respectively. Significantly higher fish survival (%) was observed in C as compared to all the treatments (Table 4).

At the completion of the experiment, treatment with low stocking density (C) resulted in the higher fish growth. Furthermore, there was a negative correlation between the

 
 Table 2. Mean water quality parameters in control and treatments during the experimental period

Parameters	С	SD1	SD2	SD3
Temperature (°C)	18.93ª	18.61ª	18.59ª	18.58ª
рН	8.45ª	7.75⁵	7.73⁵	7.75⁵
D.O. (mg l <sup>-1</sup> )	8.52⁵	9.26 <sup>ab</sup>	9.32ª	9.38ª
TA (CaCO₃ mgl⁻¹)	172.57ª	173.43°	174.28ª	174.48ª
TH (CaCO₃ mgl⁻¹)	187.71ª	185.33ª	184.62ª	184.95°
NH₃-N (mgl⁻¹)	0.044 <sup>b</sup>	0.194ª	0.197ª	0.184ª
NO <sub>2</sub> <sup>-</sup> -N (mgl <sup>-1</sup> )	0.022ª	0.044ª	0.044ª	0.044ª
NO₃ <sup>-</sup> -N (mgl <sup>-1</sup> )	0.331 <sup>b</sup>	2.035ª	2.037ª	2.030ª
PO <sub>4</sub> <sup>3</sup> -P (mgl <sup>-1</sup> )	0.047 <sup>b</sup>	0.86ª	0.861ª	0.862ª

Values with same superscript in row do not differ significantly (p≤0.05)

 Table 3. Nutrient removal in water collected from hydroponics unit (%)

Nutrient removal (%)	Number of days					
	15	45	75	90		
Ammonia-N	86.2	91.72	97.23	98.22		
Nitrate-N	65.45	86.07	96.34	97.53		
Ortho-phosphate	66.38	79.99	94.04	96.41		

stocking density and length and/or weight increase of the fish. Similar findings were observed in terms of impact of increasing stocking density on the growth and survival rate of different fish species in different aquaculture systems (Wang et al 2017, Haridas et al 2017, Nuwansi et al 2021) by previous researchers. The biomass of Koi carp significantly increased with increasing stocking density of fish. At the end of experimental period, the average yield (kgm<sup>-3</sup>) was significantly higher in all the treatments, with the maximum yield (9.417 times) recorded in SD3 followed by SD2, SD1 and C. The results in terms of total fish biomass/yield coincide with stocking density of fish. The findings of the study are consistent with those of Nuwansi et al. (2021) for Koi carp (C. carpio var. koi) in aquaponic system. Feed efficiency in terms of FCR was significantly higher (in all aquaponic treatments with the maximum 4.24 in SD2 followed by SD1, SD3 and control (2.85). Previous studies also revealed higher values for FCR (Shete et al 2013, Hussain et al 2014, Nuwansi et al 2020, Nuwansi et al 2021) for Koi carp in aquaponic systems.

Lettuce growth: Lettuce growth in terms of plant height (cm) and plant root length (cm) was significantly higher in HM as compared to SM (Table 5). Plant height and root length increase (%) in HM was 1.42 and 1.28 times higher as compared to SM. Similarly, significantly (P≤0.05) higher (1.15 times) total yield/biomass (32.20 Qt acre<sup>-1</sup>) was in HM as compared to SM (27.90 Qt acre<sup>-1</sup>). Sabwa et al (2022) also revealed similar results in terms of biomass of lettuce and root length in aquaponics system. Purwandari et al (2017) reported that romaine lettuce grown in hydroponic system was characterized by better growth compared to plants grown in an aquaponic system fed only with wastewater coming from giant gourami (Osphronemus goramy) culture system. Hairani et al (2022), observed that among three plants i.e. kailan (Brassica oleracea L.), lettuce (Lactuca sativa L.) and pakcoy (Brassica rapa L.), lettuce showed best growth in combination with striped catfish (Pangasianodon hypophthalmus) in aquaponic system, due to presence of maximum number of leaves with wider leaf width. However, contrary to this, Rana et al (2018) revealed 1.14 times higher lettuce growth in traditional soil beds as compared to aquaponics grown crop along with other growth parameters like plant height, leaf number, and plant and root weight. The aquaponics grown lettuce showed significantly higher yield compared to soil based system, and is attributed to availability of nutrients from aquaponic fish tanks through water recirculation and excessive water nutrients uptake by the lettuce in the hydroponic unit.

**Pigment analysis:** The significantly higher pigment content i.e. chlorophyll a, chlorophyll b, total chlorophyll and carotenoid content (mg g<sup>-1</sup>) was observed in lettuce grown in

Parameters	С	SD1	SD2	SD3
Average initial body length (cm)	3.35°	3.42ª	3.43ª	3.35°
Average final body length (cm)	11.35ª	9.54°	9.75°	10.45 <sup>b</sup>
TLG (cm)	7.99ª	6.12°	6.32°	<b>7</b> .11⁵
Average initial body weight (g)	3.27ª	3.33ª	3.33ª	3.33°
Average final body weight (g)	21.20 <sup>a</sup>	10.53⁵	10.80 <sup>⊳</sup>	11.20 <sup>⊳</sup>
NWG (g)	17.93ª	7.20 <sup>b</sup>	7.47 <sup>b</sup>	7.87⁵
Fish biomass (kg m <sup>-3</sup> ) (Av. Yield)	0.396 <sup>d</sup>	1.727°	2.538⁵	3.729ª
SGR (%)	2.08ª	<b>1.28</b> <sup>♭</sup>	1.31 <sup>⊳</sup>	1.35⁵
Condition factor (K)	1.46ª	<b>1.22</b> <sup>♭</sup>	1.16 <sup>bc</sup>	0.98°
FCR	2.85°	4.10ª	4.24ª	3.87⁵
Survival (%)	93.33ª	82.00 <sup>b</sup>	78.33°	83.25 <sup>b</sup>

Table 4. Comparative growth of koi carp in control and experimental tanks

\* SD = Stocking Density, C = 2 m<sup>3</sup>, SD1= 20 m<sup>3</sup>, SD2 = 30 m<sup>3</sup>, SD3= 40 m<sup>3</sup>

Values with same superscript (a, b,....d) in row do not differ significantly (p≤0.05)

TLG-Total Length Gain; NWG- Net Weight Gain; SGR- Specific Growth Rate; FCR- Feed Conversion Ratio

Table 5.	Comparative growth and pigment content of lettuce
	in SM and HM

In Ownand I IW			
Growth parameters	SM	HM	CD <sup>*</sup> (=0.05)
IPH (cm)	7.51	7.32	NS <sup>∺</sup>
PH at 30 DAT (cm)	13.22	19.45	3.80
PH at 60 DAT (cm)	16.8	20.37	1.96
PH at 90 DAT (cm)	17.4	24.17 (1.39)	2.39
PHI (%)	2.32	3.30 (1.42)	-
IRL (cm)	3.36	3.46	NS
FRL (cm)	10.39	32.77 (3.15)	3.38
RLI (%)	20.13	25.85 (1.28)	-
Yield (Qt acre <sup>-1</sup> ) <sup>#</sup>	27.90	32.20 (1.15)	NS
Yield (cup <sup>-1</sup> or plant <sup>-1</sup> )	485.27	632.78 (1.30)	79.53
Chlorophyll a (mg g <sup>-1</sup> )	4.64	5.34	0.39
Chlorophyll b (mg g <sup>-1</sup> )	2.97	3.62	0.30
Total chlorophyll (mg g <sup>-1</sup> )	2.60	3.14	0.11
Carotenoids (mg g <sup>-1</sup> )	3.53	3.89	0.29

Values in parentheses indicate change over control

<sup>#</sup>three cuttings of lettuce during 3 month experimental period in all SM and HM IPH/FPH – Initial/Final Plant Height, PHI – Plant Height Increase, IRL/FRL – Initial/Final Root Length, RLI – Root Length Increase, DAT –Days After Transplanting

HM as compared to SM (1.15, 1.22, 1.21 and 1.10 times higher, respectively) (Table 5). Rambe (2013) mentioned that leaf width is conducive to photosynthesis due to the presence of chlorophyll, because of which plants can independently prepare their own food for their growth. Therefore, more the number of leaves, more space for photosynthesis and better

Table	6.	Quality	of	lettuce	in	different	treatments	after
completion of experiment (%)								

Parameter (%)	Lett	P value	
	SM	HM	
Protein	1.39±0.23	1.41±0.17	0.94
Fat	0.30±0.02	0.26±0.00	1.12
Carbohydrate	1.35±0.41	1.08±0.04	0.54
Ash	1.63±0.03	1.51±0.20	0.59
Moisture	93.33±0.29	94.00±0.12	0.10
Fiber	1.98±0.03	1.72±0.03	0.00

p-value 0.05 or less indicate significant difference

plant growth. Higher pigment content in HM is indicative of extra leaf width and more number of leaves. Coronel et al (2008), also observed that total chlorophyll content was 1.09 times greater in the lettuce grown hydroponically as compared to traditionally grown lettuce. Sabwa et al. (2022) too reported higher total chlorophyll content (0.019-0.56 ugl<sup>-1</sup>) in aquaponics grown lettuce.

**Nutrient composition of lettuce:** After completion of experiment, quality (%) of lettuce in terms of proteins, fat, carbohydrates, ash, fiber and moisture, was estimated for each treatment and control. The differences for nutrient composition among treatments and control were insignificant except fiber (Table 6).

## CONCLUSION

The overall results indicated the potential of aquaponic technology with higher fish productivity (over 9 times) and lettuce biomass (15%) with improved pigment content. The synergistic combination of Koi carp and lettuce in aquaponics system with encouraging results is attributed to improved

water quality maintained in the aquaculture tanks of aquaponic system for fish through water recirculation and availability of copius amount/uptake of nutrients by the lettuce in the hydroponic unit, as compared to traditional soil bed system.

#### ACKNOWLEDGEMENTS

The financial support provided by Centre for Development of Advanced Computing (C-DAC), Mohali of Ministry of Electronics and Information Technology (MeitY) for establishing automated aquaponic unit at College of Fisheries, GADVASU, Ludhiana is highly acknowledged.

## AUTHOR CONTRIBUTION

Khushvir Singh – Conducted the experiment, analyzed the data and manuscript writing, Vaneet Inder Kaur – Conceived the idea and critical evaluation of data and editing of manuscript, Meera D. Ansal – Designing of experiment and evaluation of results, Dilpreet Talwar – Helped in maintenance of vegetable crops in hydroponic unit, Abhishek Srivastava - Helped in maintenance of fish in aquaponics unit, Kulbir Singh – Helped in overall maintenance of hydroponic unit.

#### REFERENCES

- APHA 2012. Standards methods for the examination of water and wastewater. 22nd ed. Washington, D. C. USA.
- AOAC 2007. Official Methods of Analysis. Association of Official Analytical Chemists International, 18<sup>th</sup> Edition. Gaithersburg. MD (USA). 18th Edition.
- Cheema H and Singh B 1991. *Software statistical package CPCS-1*. Department of statistics, Punjab Agric University, Ludhiana, India.
- Coronel G, Chang M and Rodríguez-Delfín A 2008. Nitrate reductase activity and chlorophyll content in lettuce plants grown hydroponically and organically. *In International Symposium on Soilless Culture and Hydroponics* 843: 137-144.
- Da Silva Cerozi B and Fitzsimmons K 2016. Use of *Bacillus spp*. to enhance phosphorus availability and serve as a plant growth promoter in aquaponics systems. *Scientia Horticulturae* **211**: 277-282.
- Endut A, Jusoh A, Ali N, WanNik WNS and Hassan A 2009. Effect of flow rate on water quality parameters and plant growth of water spinach (*Ipomoea aquatica*) in an aquaponic recirculating system. *Desalination and Water Treatment* **5**: 19-28.
- FAO 2022. The State of World Fisheries and Aquaculture 2022. Towards Blue Transformation. Rome, FAO. pp 1-266. https://doi.org/10.4060/cc0461en
- Hairani D, Dewi N and Rahardja BS 2022. The use of kailan (*Brassica oleracea L.*), lettuce (*Lactuca sativa L.*) and pakcoy (*Brassica rapa L.*) in the cultivation of striped catfish (*Pangasianodon hypophthalmus*) aquaponic system on blood glucose levels and oxygen consumption levels. IOP Conference Series: *Earth and Environmental Science*. 1036. 012073. 10.1088/1755-1315/1036/1/012073.
- Handbook on Fisheries Statistics 2023. *The State of World Fisheries* and Aquaculture: Sustainability in action. Department of Fisheries, Ministry of Fisheries, Animal Husbandry & Dairying, Government of India, New Delhi. Pp. 4-8.

- Hussain T, Verma AK, Tiwari VK, Prakash C, Rathore G, Shete AP and Nuwansi KKT 2014. Optimizing koi carp, *Cyprinus carpio* var. koi (Linnaeus, 1758), stocking density and nutrient recycling with spinach in an aquaponic system. *Journal of the World Aquaculture Society* **45**(6): 652-661.
- Haridas H, Verma AK, Rathore G, Prakash C, Sawant PB and Babitha Rani AM 2017. Enhanced growth and immunephysiological response of Genetically Improved Farmed Tilapia in indoor biofloc units at different stocking densities. *Aquaculture Research* **48**(8): 4346-4355.
- Lin YF, Jing SR, Lee DY and Wang TW 2002. Nutrient removal from aquaculture wastewater using a constructed wetlands system. *Aquaculture* **209**(1-4): 169-184.
- Morrison J, Morikawa M, Murphy M and Schulte P 2009. Water Scarcity and climate change. Growing risks for business and investors, Pacific Institute, Oakland, California, p 1-60.
- Nuwansi KKT, Verma AK, Chandrakant MH, Prabhath GPWA and Peter RM 2021. Optimization of stocking density of koi carp (*Cyprinus carpio* var. koi) with gotukola (*Centella asiatica*) in an aquaponic system using phytoremediated aquaculture wastewater. *Aquaculture* **532**: 735993.
- Nuwansi KKT, Verma AK, Rathore G, Chandrakant MH, Prabhath GPWA and Peter RM 2020. Effect of hydraulic loading rate on the growth of koi carp (*Cyprinus carpio* var. koi.) and Gotukola (*Centella asiatica* (L.) using phytoremediated aquaculture waste water in aquaponics. *Aquaculture International* **28**: 639-652.
- Nuwansi KKT, Verma AK, Tiwari VK, Prakash C and Chandrakant MH 2017. Standardization of the stocking density ratios of koi carp (*Cyprinus carpio* var. koi): Goldfish (*Carassius auratus*) in polyculture aquaponic recirculating system. *Turkish Journal of Fisheries and Aquatic Sciences* **17**(6): 1271-1278.
- Purwandari Y, Effendi H and Wardiatno Y 2017. The use of gouramy (Osphronemus goramy) rearing wastewater for growing romaine lettuce (Lactuca sativa I. var. longifolia) in aquaponic system. Asian Journal of Microbiology, Biotechnology and Environmental Sciences 19: 121-128.
- Rambe MY 2013. Penggunaan Pupuk Kandang Ayam dan Pupuk Urea Terhadap Pertumbuhan dan Hasil Tanaman Selada (*Lactuca sativa L.*) di Media Gambut (Pekanbaru: Fakultas Pertanian Universitas Islam Negeri Sultan Syarif Kasim).
- Rana KMS, Jahan M, Ferdous Z and Salam MA 2018. Production performance of lettuce (*Lactuca sativa*): aquaponics versus traditional soil. *Asian Journal of Medical and Biological Research* **4**(2): 149-156.
- Sabwa JA, Manyala JO, Masese FO, Fitzsimmons K, Achieng AO and Munguti JM 2022. Effects of stocking density on the performance of lettuce (*Lactuca sativa*) in small-scale lettuce-Nile tilapia (*Oreochromis niloticus* L.) aquaponic system. *Aquaculture, Fish and Fisheries* 2(6): 458-469.
- Sas-Paszt Lidia, Trzcinski Pawel, Lisek Anna, Gluszek Slawomir, Matysiak Bożena and Kaniszewski Stanislaw 2023. The influence of consortia of beneficial microorganisms on the growth and yield of aquaponically grown romaine lettuce. *Agronomy* **13**: 546.
- Shete AP, Verma AK, Chadha NK, Prakash C, Peter RM and Ahmad I 2016. Optimizing of hydraulic loading rate in aquaponic system with Common carp (*Cyprinus carpio*) and Mint (*Mentha arvensis*). Aquacultural Engineering **72**: 53-57.
- Shete AP, Verma AK, Tandel RS, Prakash C, Tiwari VK and Hussain T 2013. Optimization of water circulation period for the culture of goldfish with spinach in aquaponic system. *Journal of Agricultural Science* 5(4): 26-30.
- Wang C, Chang C, Dahms HU and Lai H 2017. Effects of stocking density of tilapia on the performance of a membrane filtrationrecirculating aquaponic system. *Desalination and Water Treatment* 96: 22-32.

Received 05 July, 2024; Accepted 11 September, 2024



Manuscript Number: 4361 NAAS Rating: 5.38

# Nest Site Selection and Habitat Preference of Baya Weaver *Ploceus philippinus* in Agricultural Landscape

Sukhpreet Kaur Sidhu, Gurkirat Singh Sekhon and Tejdeep Kaur Kler

Department of Zoology, College of Basic Sciences and Humanities, Punjab Agricultural University, Ludhiana-141 004, India E-mail: gurkiratsekhon@gmail.com

**Abstract:** Study on nest site selection and habitat preferences of Baya Weaver (*Ploceus philippinus*) was carried out in the breeding season of 2018 to 2020 in three districts namely Ludhiana, Rupnagar and Ferozepur. Three sites per district were selected and a total of nine nesting colonies were studied. A total of 298, 312 and 240 nests were recorded at selected locations during breeding season of 2018, 2019 and 2020 respectively. Maximum number of nesting colonies were 77.77% on date palm (*Phoenix dactylifera*) followed by 11.11% each on fishtail palm (*Caryota Urens*) and royal palm (*Roystonea regia*). The 66.66% of them were located near human habitations, 44.44 % within the houses/building premises and 22.22% were within the villages. The 33.3 and 77.73% were located in the agricultural fields and in vicinity of water bodies respectively. Positive correlation of distance from agricultural fields, water body and different species of palm trees with number of nests as well as with population number showed the significance of these habitat features for nest site selection of Baya Weaver.

Keywords: Agricultural landscape, Baya Weaver, Habitat, Nest site selection, Palm, Water bodies

India is a home to 1263 bird species including four weaver species belonging to family Ploceidae, out of the 124 weaver species recorded worldwide (Hoyo et al 2016, Parveen et al 2016). Baya Weaver (Ploceus philippinus Linnaeus 1766) is familiar throughout the Indian subcontinent and the adjoining countries of Pakistan, Bangladesh, Thailand Malaysia and Sri Lanka (Ali and Ripley 1983). Baya Weaver is a colony nester bird; workers had recorded the nesting colonies of Baya Weaver on thorny plants (Raju 2009). Different workers had recorded baya weaver preferring palm trees as their nesting sites on the west coast of India. Borges et al (2002) observed diverse types of nesting sites including eucalyptus trees were recorded. Raju (2009) had noted the use of leaves of Cycas sphaerica for nest construction by baya weaver near Tirumala Hills. Sohi and Kler (2017) had observed natural and man-made materials used by different avian species including baya weaver for nesting in Punjab. There was lack of information on the population and preferred nesting sites of baya weaver in Punjab from last 40 years. Keeping that lacuna in consideration, work on nest site selection and habitat preferences for nesting of Baya Weaver was undertaken.

# MATERIAL AND METHODS

The present study was carried out during the breeding season of 2018 to 2020 in three districts of Punjab State namely Ludhiana, Rupnagar and Ferozepur termed. Data was taken on nest site selection and habitat preferences of baya weaver (*Ploceus philippinus*) at nine selected sites in Baranhara (nesting colony I), Raghunath enclave (nesting colony II) and Rattan (nesting colony III) falling in district Ludhiana; Fatehpur (nesting colony IV), Mukarabpur (nesting colony V) and Manjitpur (nesting colony VI) in district Rupnagar; village Haraj (nesting colony VI), Wan (nesting colony VIII) and Toot (nesting colony IX) in district Ferozepur. Point count method was followed to observe the Baya Weaver's population and habitat preferences for nesting colonies (Verner 1985). Observations were made on weekly basis in morning between 6:00 am to 8:00 am in summers and between 7:00 am to 9:00 am in winters. For statistical analysis, correlation was carried out using SPSS v 16.0.

# **RESULTS AND DISCUSSION**

Pre-survey was carried out in 16 villages of three districts as mentioned before. Nesting colony I, II and III were located on a Fishtail Palm *Caryota Urens* (9.75 m tree height) in residential house, Date Palm *Phoenix dactylifera* (7.62 m tree height) in residential house and on Royal Palm Roystonea regia (8.53 m of tree height) situated in religious place. Distance to the crop fields and water body was highest for colony I followed by II. Distance to crop fields was least in case of colony III (Table 3). Nesting colonies IV, V and VI were observed on the Date Palm of tree heights ranging from 10.36 to 13.10m (Fig. 1, Table 1). Nesting colonies VII, VIII and IX of baya weaver were located on date palm of varying heights. Nesting colonies of baya weaver were highest on date palm followed by fishtail palm and royal palm. In absence of preferred date palm at Baranhara and Rattan, colonies were noted on fishtail palm and royal palm (Table 1). Six colonies were present near human habitation and seven colonies were within the vicinity of water body (Fig. 1). Trees selected for nesting colony I, VII, VIII and IX were solitary trees and nesting colony III was found on trees planted in a row. The nesting colonies IV and V were found on tree groups.

In breeding season of 2018, 2019 and 2020, a total of 298, 312 and 240 nests were recorded. In breeding season 2018, 136 nests were recorded out of which 58 were noted in nesting colony I, 43 in nesting colony II and 35 in nesting colony III during breeding season of 2018. At LI, 46, 30 and 26 nests were recorded active out of 58, 43 and 35 nests in nesting colony I, II and III. Total number of nests 28, 38 and 30 were observed in the nesting colony IV, V and VI. Out of total 96 nests, 71 were recorded active while 25 were recorded abandoned. During the breeding season of 2018, the nesting colonies were extended to adjoining Date Palm trees. A number of 21, 18 and 27 nests (66 in total) were recorded in nesting colony VII, VIII and IX during breeding season of 2018. Active nests 53 and abandoned nests 13 out of total 66 nests were noted (Table 1). Maximum population number of 340 was recorded at nesting colony I and minimum population number of 111 was recorded at nesting colony VII. During present study, nesting colonies were observed on the trees with height ranging from 25-43 ft. Selection of higher tree height seemed to make nests inaccessible to predators thus giving survival advantage.

In 2019, maximum of 72 nests were noted in nesting colony I wherein 59 nests were active and 13 nests were abandoned. Thirty-seven nests were noted in nesting colony

Il out of which 28 nests were active and 9 were abandoned. A total of 32 active nests were recorded out of 46 nests in nesting colony III. Thirty nests were observed on main nesting tree whereas 2-10 nests were extended to other palm trees present in the row adjoining to the main nesting tree during the breeding season of 2019 at III. During breeding season of 2019, 17 nests were recorded in nesting colony IV. A total of 33 and 19 nests were recorded in nesting colony V and VI in which colonies were extended to adjoining date palm trees. A total of 80 nests were located during breeding season of 2019, out of which 19 and 41 nests were noted in nesting colony VII and IX. In nesting colony VIII, 28 nests were recorded in which eight nests were extended to the adjoining Dhek tree *Melia azedarach* (Table 1).

In 2020, there were 51 active nests out of total 64 nests recorded in nesting colony I. Thirty-nine nests were observed in nesting colony II out of which 29 nests were active. A total of 27 active nests out of total of 43 nests were recorded in nesting colony III. A total of 19 nests were noted and active nests were 14 in nesting colony IV. A total of 30 and 20 nests were recorded in nesting colony V and VI. A total of 79 nests were located during breeding season of 2020, out of which 16, 26 and 37 nests were noted in nesting colonies VII, IX and IX. In nesting colony VII, there were 16 total nests having13 active and 3 abandoned nests. In nesting colony VIII and IX, 19 and 28 active nests out of total 26 and 37 nests were recorded respectively. Baya weaver was observed using palm fronds at nesting colonies constructed at different palm trees. Broad strand leaves of sorghum bicolor crop were utilized in nesting colonies III, IV, V, VI, VII, VIII and IX. Leaves of pearl millet Pennisetum glaucum were also observed in construction of nesting colonies V, VI and VII. Thin strands from grasses and the dry strands from the heaps of husk

**Table 1.** Nesting tree types, active nests and population number of Baya Weaver Birds

Nesting colonies Nesting tree		Tree height	Number of nests during different breeding seasons							Population number in		
		(m) -	2018		2019		2020		2018	2019	2020	
		_	Total	Active	Total	Active	Total	Active				
Baranhara (I)	Fishtail Palm	9.75	58	46	72	59	64	51	340	354	349	
Raghunath enclave (II)	Date Palm	7.62	43	30	37	28	39	29	244	180	205	
Rattan (III)	Royal Palm	8.53	35	26	46	32	43	27	214	275	223	
Fatehpur (IV)	Date palm	10.36	28	22	17	13	19	14	173	139	142	
Mukarabpur (V)	Date palm	13.10	38	24	33	25	30	21	273	256	238	
Manjitpur (VI)	Date palm	12.80	30	25	19	16	20	17	195	154	157	
Haraj (VII)	Date palm	13.10	21	17	19	17	16	13	111	168	141	
Wan (VIII)	Date palm	11.27	18	15	28	20	26	19	261	193	206	
Toot (IX)	Date palm	12.19	27	21	41	30	37	28	128	254	248	

were also observed used in the nest formation at nesting colonies I, IV, VI, VII and VIII. Males predominantly formed the nests; multiple trips from crop fields to nests were carried out. On an average, 2-4 minutes were spent in weaving nests between consecutive trips. Parent birds were noted using mud blebs as nesting material at five colonies while it was not noted at colonies II and V might be due to the presence of cemented border walls at water bodies (Table 2). Proximity of



Fig. 1. Distance of nesting colonies to agricultural field, water body, main road and residential area (in meters)

 Table 2. Nesting materials used at different nesting colonies

 Nesting material types Tree/crop used
 Nesting Colonies

Nestingmaterial type:	s nee/crop used	Nesting Colonies
Broad strand leaves	Fishtail Palm	I
	Royal Palm	III
	Sorghum	III, IV, V, VI, VII, VIII, IX
	Pearl Millet	V, VI, VII
	Date Palm	II, IV, V, VI, VIII
Thin strands	Grasses, Weeds	I, IV, VII, VIII
	Heaps of husk	IV, VI
Mud blebs	Soil from fields, pond sides	I, III, IV, VII, VIII
Feathers	-	I, II

colonies to water bodies and agricultural fields seemed to reduce not only energy expenditure but also time spent in hunting food for nestlings. Females were observed collecting and plastering mud blebs on the inner wall of brood chamber at nesting colonies I, III, IV, VII and VIII. Cushioning of brood chamber was done using feathers collected by females at nesting colonies I and II.

Dhindsa (1980) had recorded 45 colonies, out of which 20 colonies were present on small trees, bushes and other vegetation growing over abandoned wells. The most preferred trees were Babool *Acacia arabica* having 88 colonies followed by Indian date *Phoenix sylvestris* having 56 and Ber *Zizyphus jujube* having 47 colonies. Borges et al (2002) had stated affinity of Baya Weaver towards stout and tall trees because of strength attained by trees due to diameter at breast height (DBH) and safety of nestlings from predators. Workers had recorded the heights of trees having nests viz. seven to eight meter in Assam (Yashmita-Ulman et al 2017), 10m in Tamil Nadu (Ali 2009). The studied sites had experienced decline in the nests in heronries of Baya Weaver (Roshnath et al 2013, 2014).

Sohi and Kler (2017) found pendant nests of baya weaver on date palm, ber and pear trees; Baya weaver used grass blades, leaves, fibres, strips of palm fronds and cloth pieces as nesting materials. Kabir (2018) had the collection of nesting material by Baya Weaver from coconut trees and mature leaves of Palm trees. Pandian and Ahimaz (2018) had recorded 4273 nests of various stages in 52 villages in Viluppuram district of Tamil Nadu. Yasmita-Ulman et al (2017) had recorded *A. catechu*, *P. sylvasticus*, *C. nucifera* and *B. flabellifer* as potential host trees for the nesting of *P. philippinus*. Achegawe et al (2016) recorded the average tree height of nesting tree was 4.5m and nests were present on marginal parts of trees for their safety from predators. The site selection of colonial nesting can also reduce predation

Table 3. Habitat features including vegetation and sub habitat types around nesting colonies

Nesting colonies	Vegetation structure around nesting colonies			Habitat structure (distance to sub habitats)
	Number of trees	Number of crops	Number of weeds	
I	13	3	4	Agriculture fields> Water body> Main Road
П	12	-	3	Agriculture fields> Water body> Main Road
III	30	4	-	Residential area> Main Road> Agriculture fields> Water body
IV	9	2	4	Main road> Agriculture fields> Residential area> Water body
V	19	12	4	Agriculture fields> Water body> Main Road
VI	10	6	1	Main road> Agriculture fields
VII	9	4	4	Residential area> Main Road> Agriculture Water body
VIII	8	4	3	Water body> Agriculture fields> Main Road
IX	5	4	-	Residential area> Main Road



Fig. 2. Correlation of number of nests and population number of Baya Weaver with distance to the habitat structures during 2018



Fig. 3. Correlation of number of nests and population number of Baya Weaver with distance to the habitat structures during 2019



Fig. 4. Correlation of number of nests and population number of Baya Weaver with distance to the habitat structures during 2020

pressure and increase foraging efficiency (Kasprzykowski 2008). The positive correlation of avian diversity with rural ponds was observed in earlier studies (Sekhon et al 2023, Sidhu et al 2021). Statistical analysis had shown that distance from residential area is negatively correlated with population number during the year 2018 and positively correlated with population number during the year 2018 and positively correlated with population number during the year 2018 weaver within the vicinity of residential premises during the breeding season of 2019 (Fig. 2, 3). Distance from agricultural fields, waterbody and main road are positively correlated with number of nests and population number of Baya Weaver that showed the importance of these habitat structures for the

nesting and breeding of Baya Weaver (Fig. 3 and 4). Tree species abundance varied considerably (5-30), colony III having the highest number of trees in vicinity, while colony IX exhibited the least. Six colonies (III, V, VI, VII, VIII and IX) suggested potential dependence on crops for food and nesting materials. Four colonies (I, IV, V, VII) had substantial weed species in surrounding habitats indicating potential foraging grounds, while two colonies (III and IX) lacked it in their vicinity (Table 3).

#### CONCLUSION

The nest site selection by baya weaver showed preference for date palm followed by fishtail palm and royal palm. Habitat features included proximity to water bodies, agricultural fields and preferred tree heights ranging from 25-43 ft. Nest site selection seemed to be determined by factors such as safety from predators, energy expenditure during foraging trips and time saving during nest building and nestling rearing. Current investigation provides valuable information regarding significance of Palm tree diversity and small water bodies in agricultural landscape which might increase the range of habitat or nest site selection of Baya Weaver and help in sustaining stable population numbers.

#### REFERENCES

- Achegawe RM, Shivaji PC, Priyanka VP and Shashikant HT 2016. Nesting of Baya Weaver (*Ploceus philippinus*) in S.R.T.M. University, Nanded and fields along Asana River at Nanded, Maharashtra, India. *International Journal of Current Research* and Academic Review 4(6): 51-60.
- Ali AM 2009. Studies on nest site selection and prey delivery patterns to nestlings by the Baya Weaver *Ploceus philippinus* in Tamil Nadu, South India. *World Journal of Zoology* **4**(4): 308-312.
- Ali S and Ripley SD 1983. Handbook of the Birds of India and Pakistan. Oxford University Press Inc. Bombay, India, p 247.
- Borges S D, Desai M and Shanbhag A B 2002. Selection of nest platforms and the differential use of nest building fibres by the Baya weaver, *Ploceus philippinus* Linnaeus 1766. *Tropical Zoology* **15**(1):17-25.
- Dhindsa MS 1980. *Ecological studies on the Weaver Birds of the Punjab and their control*. Ph.D. Dissertation, Punjab Agricultural University, Ludhiana, India.
- Kasprzykowski Z 2008. Nest selection within the tree and breeding parameters of Rooks Corvus frugilegus. Bird Study 55: 59-65.
- Hoyo JD, Collar N and Christi DA 2016. *HBW and Bird Life International Illustrated Checklist of the Birds of the World.* Volume **2**. Lynx Edicions ISBN: 978-84-96553-98-9.
- Kabir MA 2018. Nesting of Baya Weaver *Ploceus philippinus* in Mohammadpur village under Rajshahi Division of Bangladesh. *International Journal of Research Studies in Science, Engineering and Technology* **5**(7): 21-25.
- Pandian M and Ahimaz P 2018. Nesting behaviour of the Baya Weaver, *Ploceus philippinus* (Linnaeus) (Passeriformes: Ploceide) in rural Tamil Nadu, India. *International Journal of Ecology and Environment Science* 44(1): 33-42
- Parveen J, Jaypal R and Pittie A 2016. A checklist of birds of India. *Indian BIRDS* **11**(5): 113-172.
- Raju AJS 2009. Nesting behaviour of the Baya Weaver, *Ploceus philippinus* (Ploceidae) and the life-cycle of the Plain Cupid butterfly, *Chilades pandava* (Lycaenidae) with red-listed *Cyas*

sphaerica and C. beddomei (Cycadaceae). Journal of Threatened Taxa 1(8): 429-433.

- Roshnath R, Ashokkumar M, Unni R, Jith S and Jose A 2013. Status of birds in Heronries of Kannur district, Kerala. *Malabar Trogon* **11**: 15-20.
- Roshnath R, Divakar N, Chandran K, Valsarajn D and Jose A 2014. Heronry census, 2014 in Kannur district, Kerala. *Malabar Trogon* **12**: 9-13.
- Sekhon GS, Aulakh RK and Kler TK 2023. Village ponds as unexplored habitation sites for resident migratory and migratory bird species in Punjab State, India. *Indian Journal of Ecology*. DOI: https://doi.org/10.55362/IJE/2023/0000

Sohi GK and Kler TK 2017. Adaptations in avian nesting behaviour

Received 08 June, 2024; Accepted 01 October, 2024

in relation to indigenous trees and housing structures in Punjab. *Journal of Entomology and Zoological Studies* **5**(5): 1045-1051.

- Sidhu SK, Sekhon GS, Aulakh RK and Kler TK 2021. Prioritizing Sustenance of village ponds for avian conservation: A case study from Punjab, India. *Pakistan Journal of Zoology* 1-8 DOI: https://dx.doi.org/10.17582/journal.pjz/20190918070927.
- Verner J 1985. Assessment of counting techniques. Current Ornithology 2: 247-302.
- Yashmita-Ulman, Kumar A and Sharma M 2017. Traditional home garden agroforestry system: Habitat for conservation of Baya Weaver *Ploceus Philippinus* (Passeriformes: Ploceidae) in Assam, India. *Journal of Threatened Taxa* **9**(4): 10076-10083.



Indian Journal of Ecology (2024) 51(5): 1109-1116 DOI: https://doi.org/10.55362/IJE/2024/4362 Manuscript Number: 4362 NAAS Rating: 5.38

# Status and Impact of Wooded Patches in Semi-Urban Landscape on Avian Community Structure in Aligarh, Uttar Pradesh, India

Aditya Rana and Jamal Ahmad Khan

Department of Wildlife Sciences, Aligarh Muslim University, Aligarh-202 002, India E-mail: ranaaditya1751@gmail.com

**Abstract:** This short-term study is an effort to estimate density and diversity of birds in wooded patches present at the outskirts of Aligarh city. Point count method was used to determine the diversity and abundance of birds. The total of 40 points were laid through systematic random sampling and monitored in the morning hours. In total, 140 species were recorded, with Passeriformes as the most dominating order. Seven near threatened and three threatened species were recorded. Bird density was 60 birds/ha and large grey babbler had the highest density (18.72 birds/ha). Density was significantly positively correlated with the shrub height. In addition to with resident species, many winter/summer migrants and passage migrant birds were found during the study period. These small and scattered patches are important for wildlife coping with urbanization; therefore better management of these wooded patches is required for effective conservation.

Keywords: Point count, Distance, Urbanization, Wooded patches, Density

Birds provide various ecosystem services and play important roles in our agroecosystems (Mariyappan et al . 2023). The flourishing bird population is essential for any stable ecosystem and their status indicates how well the ecosystems are functioning (Mariyappan et al 2023). Rapid urbanization is an emerging threat and it is having adverse effects on the diversity and abundance of birds and in turn, negatively affects ecosystem functioning. Urbanization has led to the conversion of natural habitats and farmlands into a series of different architectures which are heavily fragmented and have a high edge effect (Shochat et al 2006, Moller 2009). In this process, some wildlife species benefit and some are negatively affected (Devictor et al 2007, Akram et al 2015, Sahni & Kler 2023).

Urban green spaces such as parks and lawns etc. are vital for birds in urban areas but still, birds flourish in islets of natural habitats located in the vicinity of urban areas (Felappi et al 2020, Machar et al 2022). Most of the studies conducted in urban areas are usually focused on urban green spaces (Shochat et al 2006, Moller 2009, Prakash and Baldodia 2013, Rajashekara and Venkatesha 2015, Mukhopadhyay and Mazumdar 2019). These spaces only harbour those species that benefit from the urban environment and are better adapted to the changing environmental conditions (Moller 2009, Verma and Murmu 2015, Singh et al 2018).

The natural habitats located on the outskirts of cities provide more opportunities for those species that are not able to adapt to the urban environments and are more elusive, as these areas are comparatively less disturbed and less prone to the frequent changes that the city experiences (Rajashekara and Venkatesha 2015, Mukhopadhyay and Mazumdar 2019). These areas are important stopover points for locally migrating birds and provide shelter as well as variable food resources (Siddiqui et al 2019, Mazumdar and Khan 2020, Yashmita-Ulman 2022). Therefore, scientific information on the diversity and abundance of birds in these natural habitats located on the outskirts of cities is essential for the effective management of these areas.

Most of the natural vegetation of the Aligarh district is lost due to the conversion of natural areas into agricultural fields and human settlements. But few small isolated wooded patches of natural vegetation and plantations still existed in the district supporting various species of wildlife. Avian diversity of Aligarh is rich and varied and more than 150 species are reported from the district (Akram et al 2015, Siddiqui et al 2019, Mazumdar and Khan 2020). Each year many migratory birds visit the area as their wintering grounds and represent the ecological significance of the Aligarh region (Mazumder and Khan 2020). Therefore, this study is an attempt to find out avian density and diversity of these isolated wooded patches and to determine the vegetation parameters affecting density and diversity of birds.

#### MATERIAL AND METHODS

**Site location**: Aligarh district is located in the western part of Uttar Pradesh state of India and spread over an area of 3650 sq. km., from  $27.5714^{\circ} - 28.1798^{\circ}N$  to  $77.4756^{\circ} - 78.6077^{\circ}E$  (Fig. 1). A major part of the district is primarily under

agriculture (Ali 2013). There are sparsely distributed wooded patches in the district having variable sizes from 6 ha to > 200 ha surrounded by agricultural lands. Major tree species found in these patches are Vachellia nilotica, Prosopis cineraria, Azadirachta indica, along with various alien invasive species such as Prosopis juliflora, Lantana camara etc. The study was carried out from February to May 2021. Identification of probable wooded patches was conducted with the help of Google Earth Pro. A total of 14 patches were identified but after a quick reconnaissance survey and ground truthing, nine sites were selected for further study. The area of these patches varies from 19.5 ha to 256 ha (Table 1). The criteria for the selection of patches were that patches must have intact and continuous vegetation in a minimum of ~20 ha, not be owned by a private person and must not be under any agricultural practices (Fig. 1).

**Diversity and density**: To determine the diversity and density of birds, point count method was used. Systematic random sampling was used to lay the points in the study area. The first point in the patch had been chosen randomly and then successive points were laid at every 300 m in a fixed direction. A total of 40 points were laid for monitoring of birds.





Table 1. Wooded patches selected for the stud	ly
---	----

The number of points in a particular patch was laid in proportion to its area. Each point was monitored four times for 15 minutes, making a total effort of 160 points. As birds are most active in the morning hours, all the monitoring was conducted in the early mornings. Variable radius point count monitoring protocol was used following Verner (1985) and Javed and Kaul (2002). No flying bird had been considered for an observation. Birds sighted outside the point count stations were recorded on a separate sheet for preparation of the checklist.

To quantify the vegetation on the point, number of trees was counted in a 20-meter circular plot. Plants having >30 cm GBH were considered as trees. An ocular estimation of canopy cover was also taken at each point. Visual obstruction technique via Robel Pole was used to measure shrub layer height. A pole with black and white 10 cm bands is used. The pole was placed at a fixed distance from a point, and then the last visible band number was recorded to estimate relative heights of shrub layer in all the patches.

**Statistical analysis**: PAST (paleontological statistics) statistical software was used to compute diversity indices for each point and then further for each patch after the data had been pooled. The density of birds was calculated in DISTANCE software (v\_7.5). Multiple Covariate Distance Sampling (MCDS) was used using MCDS engine in Distance to estimate density after incorporating covariables such as tree density, canopy cover, shrub height, etc in the original dataset. Density estimation through MCDS engine gives more accurate results and better density figures (Marques et al 2007). Spearman's rank correlation was computed in PAST software to determine the correlation between bird density and various vegetation parameters.

#### **RESULTS AND DISCUSSION**

**Species occurrence:** Out of 140 bird species recorded during the study period, 71 species were recorded at point

Patch No.	Patch name	Area (ha)	Major tree species
1	Old Jawan Nursery	19.5	Cassia fistula, Holoptelea integrifolia, Magnifera indica & Vachellia nilotica
2	Jawan Patch (beside Hitachi Warehouse)	34	Prosopis juliflora
3	Jangalgarhi	23.2	Prosopis juliflora
4	Pala	41.8	Prosopis juliflora
5	Pala Sallu Blackbuck Community Reserve	33.9	Vachellia nilotica & Prosopis juliflora
6	Sikandrapur Chherat–- Manzurgarhi	75.56	Vachellia nilotica & Prosopis juliflora
7	Bilauthi	54.7	Prosopis juliflora
8	Akrabad	43	Vachellia nilotica & Prosopis juliflora
9	Gurushikharan	256.26	Vachellia nilotica & Prosopis juliflora
stations and a total of 1785 individuals were observed belonging to 18 orders and 56 families. Passeriformes with 68 species was the most dominating order (48.6%) in the study area, while only two species each from three orders (1.43%) have been recorded (Table 2). Birds are usually better adapted to urban establishments than any other faunal group (Luniak 2004). Wooded patches situated outside the city environments are being used more extensively by the negatively affected species as a result these species are seldomly seen in the cities (Siddiqui et al 2019). These patches provide refuge to these species as they are being driven away from the city due to various developmental processes.

The number of bird species (140 species) recorded is close to the species recorded by Mazumdar and Khan (2020) (146 species) and higher than the other studies Akram et al (2015) (92 species); Siddiqui et al (2019) (63). Birds are an indicator of the health of any ecosystem (Wade et al 2014). The rich diversity of birds in the study area highlights how important these wooded patches are for the sustainance of birds and in turn overall health of various ecosystems (Wade et al 2014, Mariyappan et al 2023).

Seven recorded species are near threatened, two are vulnerable (Indian Spotted Eagle Clanga hastata & Sarus Crane Grus antigone) and one (Egyptian Vulture Neophron percnopterus) is endangered. Eurasian Collared Dove had the highest relative frequency (8.77%) while 16 species had only 0.13% as the lowest relative frequency (Table 3).The seven near-threatened and three threatened species recorded in the study area need immediate conservation focus (IUCN 2023). Regular assessment focused on these species will provide insights for their better management and survival. Egyptian vulture, in particular, is an endangered species (IUCN 2023), but their frequent recordings from the study area, either in the form of a single individual or in large flocks (recorded 23 individuals in a single flock), may indicate yet another success story for the revival of an endangered species in the study area (Singh and Gibson 2011, Kierulff et al 2012). Grimmett et al (2011) observed that many bird species recorded in the study are winter and summer migrants. Furthermore, some species are passage migrants and utilise these patches as their stopover points. All of these species utilise these patches for varying duration in different seasons of the year; indicating the importance of these small and scattered wooded patches for them.

**Density and diversity of birds:** Based on minimum AIC value, the Hazard Rate- Simple Polynomial model was selected to calculate the density of birds. After considering all the covariables, terrestrial birds' density was 60.63 birds/ha (Table 4). Highest density was recorded in Old Jawan

Nursery patch with 235.21 birds/ha whereas lowest density in Jawan Patch (beside Hitachi warehouse) with 52.097 birds/ha. Bird density was is higher than Siddique et al (2019) with 36.82 birds/ha and Akram et al (2015) with 54.45 birds/ha but these studies have more focus on birds in urban areas of Aligarh district.

Further, the diversity of birds is more or less similar to the many studies conducted in nearby areas (Gupta et al 2009, Chopra et al 2012, Prakash and Baldodia 2013, Singh et al 2021). This indicates that the Aligarh region still holds importance for various species inhabiting these wooded patches and to the various migratory species that happen to visit this region annually. Density of birds and Shannon diversity index e of Old Jawan Nursery was highest in all the individual patches. This is because it holds the highest tree density and relatively high shrub height; these factors help in creating more available niches and hence greater density and diversity (Pigot et al 2016, Kaur and Kler 2019). Keeping this patch as a model, we can further enhance the carrying capacity of other patches, leading to greater avian density and diversity and also giving ideas for better management of such patches.

Large grey babbler have the highest density with 18.72 birds/ha, followed by Green Bee-eater (11.68 birds/ha) (Table 5) which was due to agroecosystems surrounding the

 Table 2. Number and percentage of species in their respective orders

Order	No. of families	Percentage of species
Accipitriformes	1	7.14
Anseriformes	1	2.14
Bucerotiformes	2	1.43
Caprimulgiformes	2	1.43
Charadriiformes	5	5.71
Ciconiiformes	1	2.14
Columbiformes	1	5.00
Coraciiformes	3	2.86
Cuculiformes	1	2.86
Galliformes	1	1.43
Gruiformes	2	2.86
Passeriformes	26	48.57
Pelecaniformes	2	6.43
Piciformes	2	2.14
Podicipediformes	1	0.71
Psittaciformes	1	2.14
Strigiformes	2	2.86
Suliformes	2	2.14

1	1	1	2

**Table 3.** Bird species listed on points with their frequency of occurrence and relative frequency

Species	Scientific name	Frequency of occurrence	Relative frequency %
Indian Peafowl	Pavo cristatus	19	2.38
Grey Francolin	Francolinus pondicerianus	28	3.51
Rock Pigeon	Columbia livia	2	0.25
Oriental Turtle Dove	Streptopelia orientalis	12	1.50
Eurasian Collar Dove	S. decaocto	70	8.77
Red Collar Dove	S. tranquebarica	4	0.50
Spotted Dove	Spilopelia suratensis	3	0.38
Laughing Dove	S. senegalensis	61	7.64
Yellow-footed Green Pigeon	Treron phoenicopterus	21	2.63
Greator Coucal	Centropus sinensis	8	1.00
Asian Koel	Eudynamys scolopaceus	13	1.63
Common Hawk Cuckoo	Hierococcyx varius	5	0.63
Indian Cuckoo	Cuculus micropterus	1	0.13
Indian Thick-knee	Burhinus indicus	10	1.25
Red Wattled Lapwing	Vanellus indicus	9	1.13
Indian Pond Heron	Ardeola grayii	3	0.38
Red Naped Ibis	Pseudibis papillosa	1	0.13
Egyptian Vulture	Neophron percnopterus	2	0.25
Oriental Honey Buzzard	Pernis ptilorhynchus	1	0.13
Crested Serpent Eagle	Spilornis cheela	3	0.38
Black Kite	Milvus migrans	10	1.25
Spotted Owlet	Athene brama	9	1.13
Grey Hornbill	Ocyceros birostris	7	0.88
White Throated Kingfisher	Halcyon smyrnensis	4	0.50
Asian Green Bee-eater	Merops orientalis	67	8.40
Brown Headed Barbet	Psilopogon zeylanicus	1	0.13
Lesser Goldenback Woodpecker	Dinopium benghalense	4	0.50
Alexandrine Parakeet	Psittacula eupatria	2	0.25
Rose Ringed Parakeet	P. krameri	22	2.76
Plum Headed Parakeet	P. cyanocephala	1	0.13
Black Drongo	Dicrurus macrocercus	20	2.51
Ashy Drongo	D. leucophaeus	1	0.13
Isabelline Shrike	Lanius isabellinus	1	0.13
Bay Back Shrike	L. vittatus	1	0.13
Rufous Treepie	Dendrocitta vagabunda	37	4.64
House Crow	Corvus splendens	1	0.13
Large Billed Crow	C. macrorhynchos	11	1.38
Ashy Crowned Sparrow Lark	Eremopterix griseus	1	0.13
Crested Lark	Galerida cristata	2	0.25
Common Tailorbird	Orthotomus sutorius	8	1.00
Jungle Prinia	Prinia sylvatica	7	0.88
Ashy Prinia	P. socialis	23	2.88
Plain Prinia	P. inornata	1	0.13

Species	Scientific name	Frequency of occurrence	Relative frequency %
Zitting Cisticola	Cisticola juncidis	1	0.13
Blyth's Reed Warbler	Acrocephalus dumetorum	17	2.13
Barn Swallow	Hirundo rustica	2	0.25
Red Vented Bulbul	Pycnonotus cafer	22	2.76
Red Whiskered Bulbul	P. jocosus	2	0.25
Hume's Leaf Warbler	Phylloscopus humei	1	0.13
Sulphur Bellied Warbler	P. griseolus	5	0.63
Common Chiffchaff	P. collybita	3	0.38
Greenish Warbler	P. trochiloides	1	0.13
Lesser White Throat	Sylvia curruca	30	3.76
Yellow Eyed Babbler	Chrysomma sinense	1	0.13
Indian White Eye	Zosterops palpebrosus	3	0.38
Jungle Babbler	Argya striata	24	3.01
Large Grey Babbler	A. malcolmi	45	5.64
Asian Pied Starling	Gracupica contra	13	1.63
Common Myna	Acridotheres tristis	12	1.50
Indian Robin	Saxicoloides fulicatus	11	1.38
Oriental Magpie Robin	Copsychus saularis	16	2.01
Red Breasted Flycatcher	Ficedula parva	7	0.88
Black Redstart	Phoenicurus ochruros	7	0.88
Pied Bushchat	Saxicola caprata	11	1.38
Purple Sunbird	Cinnyris asiaticus	21	2.63
Indian Silverbill	Euodice malabarica	5	0.63
Scaly Breasted Munia	Lonchura punctulata	1	0.13
Yellow Throated Sparrow	Gymnoris xanthocollis	7	0.88
Paddyfield Pipit	Anthus rufulus	4	0.50
Long Billed Pipit	A. similis	6	0.75
Tawny Pipit	A. campestris	3	0.38

Table 3. Bird species listed on points with their frequency of occurrence and relative frequency

## Table 4. Bird density, diversity indices and number of species in different wooded patches of Aligarh district

Patch name	Density (Birds/ha)	% coef. of variation	No. of species	Evenness	Shannon (H) diversity	Simpson diversity	Margalef richness
Old Jawan Nursery	235.21	24.14	34	0.757	2.843	0.925	2.834
Jawan Patch (beside Hitachi Warehouse)	48.55	31.07	8	0.696	1.147	0.596	1.147
Jangalgarhi	58.47	34.31	15	0.699	1.726	0.751	1.726
Pala	52.47	24.38	21	0.698	1.904	0.792	1.904
Pala Sallu Blackbuck Community Reserve	99.74	26.05	15	0.755	2.018	0.831	2.018
Sikandrapur Chherat- Manzurgarhi	92.36	11.24	39	0.694	2.545	0.888	2.545
Bilauthi	54.52	19.47	29	0.664	2.093	0.826	2.093
Akrabad	60.56	29.90	13	0.762	1.557	0.734	1.557
Gurushikharan	53.60	18.10	38	0.731	2.120	0.830	2.120
Overall	60.63	9.19	71	0.715	2.028	0.806	2.028

wooded patches and adding to the fact that they remain in flocks that can go >20 in number (Brraich et al 2023). Crop fields are abundant in food resources for most of the year and LGB being an omnivorous species gains advantage of its diet variability (Puckett et al 2009). LGB flocks can have up to 40 individuals at one time, giving them an advantage over their potential predators as they lookout for each other with greater strength in numbers (Brraich et al 2023). With better utilization of available resources, their populations flourish and hence they are found with the highest density in the study area.

**Correlation with vegetation parameters:** Highest tree density was recorded in Old Jawan Nursery patch (131.3 trees/ha) while lowest was in Bilauthi patch (3.18 trees/ha). Similarly, highest canopy cover was in Jangalgarhi patch and lowest in Bilauthi. Shrub height in Sikandrapur Chherat-Manzurgarhi patch was highest while in Jawan patch (beside Hitachi Warehouse) recorded lowest shrub height (Table 6). Shrub height was significantly positively correlated with bird density. Although a positive correlation was recorded in bird density with tree density and canopy cover but differences were non-significant.

These wooded patches are important for the sustenance and survival of various species that are negatively affected by urbanization. These patches should be given more concern on how to conserve them and should be viewed more than just as an infertile land. Thriving species like LGB, Green Bee-eater, and Eurasian Collared Dove as well as near threatened and threatened species residing here show the vitality of these patches. For shy migratory birds, these patches are an abode and without these patches, their migration will be affected. The size of these patches also matters and no further encroachment should be allowed over them. These patches need to remain intact without any further fragmentation. In addition to birds, these patches also have a rich diversity of mammalian species and 11 species of mammals (Blackbuck Antilope cervicapra, Nilgai Boselaphus tragocamelus, Wild Pig Sus scrofa, Golden Jackal Canis aureus, Rhesus Macaque Macaca mulatta, Indian Hare Lepus nigricollis, Jungle Cat Felis chaus, Small Indian Mongoose Herpestes auropunctatus, Grey Mongoose Herpestes edwardsii, Five Striped Palm Squirrel Funambulus pennantii and Indian Flying Fox Pteropus giganteus) were recorded during the study. It also indicates the importance of these wooded patches as they also utilise these wooded patches in toto. More emphasis should be given on how to connect nearby patches so that there will be fewer road kills of wildlife present in the area. Studies conducted over mammalian communities will enhance the knowledge of their association with avian communities in the study area (Cook et al 2020). Few studies have been done in the district on mammals but their association and effect on avian communities is yet to be explored (Khan and Khan 2016, Ahamad et al 2021tc.). These broad studies will further

Table 5. Density of bird species having statistically significant observations individually

Species	Density (Birds/ha)	Per cent coef. of variation	95% confidence interval
Large Grey Babbler	18.72	23.47	11.82 – 29.64
Green Bee-eater	11.68	21.68	7.62 – 17.90
Lesser White Throat	9.03	21.41	5.92 – 13.77
Rufous Tree Pie	7.02	28.39	4.03 – 12.23
Laughing Dove	4.27	23.4	2.70 - 6.74
Eurasian Collared Dove	3.89	19.21	2.67 – 5.67

Table 6. Patch-wise vegetation parameters and other co-variables

Patch name	Tree density (per ha)	Canopy cover (%)	Shrub height (m)
Jawan (Old Nursery)	131.3	45	3.7
Jawan Patch (beside Hitachi Warehouse)	34.48	18.33	0.33
Jangalgarhi	106.1	68.33	0.47
Pala	77.59	48.75	0.65
Pala Sallu Blackbuck Community Reserve	39.79	31.67	1.8
Sikandrapur Chherat-Manzurgarhi	35.81	22.5	4.5
Bilauthi	3.18	1	2.6
Akrabad	35.81	27.5	1.45
Gurushikharan	85.15	49.5	2.56

help in the effective management and conservation of avian populations in the district.

#### CONCLUSION

Wooded patches situated at the outskirts of urban establishments provide refuge to the faunal species coping with the speed of urbanisation. Conservation and management of these patches may prove critical in ensuring the long-term conservation of biodiversity and maintaining healthy urban ecosystems.

#### REFERENCES

- Ahamad M, Khan J A and Kumar S 2021. Blackbuck *Antilope cervicapra* (Mammalia: Cetartiodactyla: Bovidae) estimates in human-dominated landscape in Aligarh, Uttar Pradesh, India. *Journal of Threatened Taxa* **13**(9): 19232-19238.
- Akram F, Ilyas O and Prusty AK 2015. Impact of urbanization on bird community structure in and around Aligarh city, U.P., India. *International Journal of Engineering Technology Science and Research* 2(10): 1-11.
- Ali B 2013. Level of agriculture and rural development in Aligarh district. Journal of Humanities and Social Science 18(3): 61-67.
- Brraich OS, Singh J and Singh G 2023. Avifaunal diversity around urban and rural areas of district Patiala, Punjab, India. *Indian Journal of Ecology* **50**(5): 1536-1544.
- Chopra G, Tyor AK, Kumari S and Rai D 2012. Status and conservation of avian fauna of Sultanpur National Park Gurgaon, Haryana, India. *Journal of Applied and Natural Science* **4**(2):207-213.
- Cook RN, Ramirez-Parada T, Browne L, Ellis M and Karubian J 2020. Environmental correlates of richness, community composition and functional traits of terrestrial birds and mammals in a fragmented tropical landscape. *Landscape Ecology* **35**: 2825-2841.
- Devictor V, Julliard R, Couvet D, Lee A and Jiguet F 2007. Functional homogenization effect of urbanization on bird communities. *Conservation Biology* **21**(3): 741-751.
- Felappi JF, Sommer JH, Falkenberg T, Terlau W and Kotter T 2020. Green infrastructure through the lens of "One Health": A systematic review and integrative framework uncovering synergies and trade-offs between mental health and wildlife support in cities. *Science of the Total Environment* **748**: e141589.
- Gupta SK, Kumar P and Malik MK 2009. Avifaunal diversity in the university campus of Kurukshetra, Haryana. *Journal of Threatened Taxa* **1**(12): 629-632.
- Grimmett R, Inskipp C and Inskipp T 2011. *Birds of the Indian Subcontinent*. 2<sup>nd</sup> Edn. India: Oxford University Press, p 528.
- IUCN 2023. The IUCN Red List of Threatened Species. 2023-1, DOI ://nc.iucnredlist.org/redlist/content/attachment\_files/2023-1\_RL\_Table\_1a.pdf
- Javed S and Kaul R 2002. Field methods for bird surveys. *Bombay Natural History Society*, p 66.
- Khan KA and Khan JA 2016. Status, abundance and population ecology of Nilgai (*Boselaphus tragocamelus* Pallas) in Aligarh district, Uttar Pradesh India. *Journal of Applied and Natural Science* 8(2): 1080-1086.
- Kierulff MCM, Ruiz-Miranda CR, Oliveira PPD, Beck BB, Martins A, Dietz JM, Rambaldi DM and Baker AJ 2012. The Golden lion tamarin *Leontopithecus rosalia*: A conservation success story. *International Zoo Yearbook* 46: 36-45.
- Luniak M 2004. Synurbization-adaptation of animal wildlife to urban development, pp.50-55. In: WW Shaw, LK Harris and L VanDruff

(eds), Proceedings of 4<sup>th</sup>International Urban Wildlife Symposium, July, 2004, University of Arizona, Tucson, USA.

- Machar I, Simek P, Schlossarek M, Pechanec V, Petrovic F, Brus J, Spinlerova Z and Sejak J 2022. Comparison of bird diversity between temperate floodplain forests and urban parks. *Urban Forestry & Urban Greening* **67**: e127427.
- Mariyappan M, Rajendran M, Velu S, Johnson AD, Dinesh GK, Solaimuthu K, Kaliyappan M and Sankar M 2023. Ecological role and ecosystem services of birds: A review. *International Journal* of Environment and Climate Change **13**(6): 76-87.
- Marques TA, Thomas L, Fancy ST and Buckland ST 2007. Improving estimates of bird density using multiple-covariate distance sampling. *The Auk* **124**(4): 1229-1243.
- Mazumder S and Khan A 2020. A checklist of birds in agricultural landscapes of Aligarh, Uttar Pradesh, India. *International Journal of Fauna and Biological Studies* **7**(2): 16-23.
- Moller AP 2009. Successful city dwellers: A Comparative study of the ecological characteristics of urban birds in the Western Palearctic. *Oecologica* **159**(4): 849-858.
- Mukhopadhyay S and Mazumdar S 2019. Habitat-wise composition and foraging guilds of avian community in a suburban landscape of lower Gangetic plains, West Bengal, India. *Biologia* **74**: 1001-1010.
- Nath PC, Arunachalam A, Khan ML, Arunachalam K and Barbhuiya AR 2005. Vegetation analysis and tree population structure of tropical wet evergreen forests in and around Namdapha National Park, northeast India. *Biodiversity Conservation* 14(9): 2019-2036.
- Pigot AL, Trisos CH and Tobias JA 2016. Functional traits reveal the expansion and packaging of ecological niche space underlying an elevational diversity gradient in passerine birds. *Proceedings of the Royal Society B* 283(1822): 20152013.
- Prakash S and Baldodia M 2013. Urban avifaunal diversity: an indicator of anthropogenic pressures in southern ridge of Delhi. *Advances in Bioresearch* **4**(2): 135-144.
- Puckett HL, Brandle JR, Johnson RJ and Blankenship EE 2009. Avian foraging pattern in crop field edges adjacent to woody habitat. *Agriculture, Ecosystems & Environment* **131**(1-2): 9-15.
- Rajashekara S and Venkatesha M G 2015. Temporal and spatial avian community composition in Urban Landscapes of the Bengaluru region, India. *Journal of Environmental Biology* **36**: 607-616.
- Raza M and Ilyas O 2018. Preliminary study on status and ecology of *Pteropus giganteus* in Aligarh city, Uttar Pradesh. *Indian Forester* **144**(10): 986-991.0
- Sahni NK and Kler TK 2023. Impact of urban landscape on relative abundance of invasive bird species. *Indian Journal of Ecology* **50**(5): 1801-1805.
- Shochat E, Warren PS, Faeth SH, McIntyre NE and Hope D 2006. From patterns to emerging processes in mechanistic urban ecology. *Trends in Ecology and Evolution* **21**(4): 186-191.
- Siddiqui A, Ahmed T and Khan A 2019. Avifaunal Assemblage along Rural-Urban Gradients in Aligarh, Uttar Pradesh, India. *Notulae Scientia Biologicae* **11**(4): 421-427.
- Singh HS and Gibson L 2011. A conservation success story in otherwise dire megafauna extinction crisis: The Asiatic Lion (*Panthera leo persica*) of Gir forest. *Biological Conservation* **144**: 1753-1757.
- Singh K, Maheshwari A and Dwivedi SV 2018. Studies on avian diversity of Banda university of agriculture and technology campus, Banda, Uttar Pradesh, India. *International Journal of Avian & Wildlife Biology* 3(2): 184-187.
- Singh KP, Riyaz M, Singh G and Syed S 2021. Avifaunal diversity of Jodhpur Jhal wetland Mathura, UP, India: a preliminary survey. *Journal on New Biological Reports* **10**(2): 95-108.
- Verma SK and Murmu TD 2015. Impacts of environmental and

disturbance variables on avian community structure along a gradient of urbanization in Jamshedpur, India. *PLoS ONE* **10**(7): e0133383.

Verner J 1985. Assessment of counting techniques, pp 247-302. In: Johnston R F (ed). *Current Ornithology*, Vol. 2. Springer, New York, USA.

Received 07 August, 2024; Accepted 30 September, 2024

- Wade ASI, Barov B, Burfield IJ, Gregory RD, Norris K, Vorisek P, Wu T and Butler SJ 2014. A niche-based framework to assess current monitoring of European forest birds and guide indicator species' selection. *PLoS ONE* **9**(5): e97217.
- Yashmita-Ulman 2022. Birds in agricultural fields of Ayodhya District, Uttar Pradesh. Indian Journal of Ecology **49**(5): 1647-1653.



Manuscript Number: 4363 NAAS Rating: 5.38

# Assessment of Hazardous Effects of Lead on Growth Parameters of Soybean *(Glycine max* (L.) Merr.)

Siddhi Gupta and M.K. Meena<sup>1</sup>

Department of Botany, Govt, College, Sardarshahar-331 403, India <sup>1</sup>Department of Botany, University of Rajasthan, Jaipur-302 004, India E-mail: siddhigupta24@gmail.com

**Abstract:** This study was designed to assessment the effects of different concentrations of heavy metal lead acetate (0, 200, 400, 600, 800, 1000 mg/kg) on growth attributes such as root shoot length, root shoot dry weight at pre, peak and post flowering stages of soybean (*Glycine max* (L.) Merr.) There was clear dose-response relationship between lead acetate concentration and the growth attributes of soybean plants. At 200 mg/kg there is no significant reduction but gradually with increased concentrations of lead acetate, there was a significant reduction in root and shoot length, as well as root and shoot dry weight, compared to the control group at all stages. The most pronounced effects were observed at 1000 mg/kg, indicating a dose-dependent toxicity of lead acetate on soybean plants. These findings highlight the importance of mitigating lead emissions and contamination to ensure agricultural productivity, environmental sustainability and public health.

Keywords: Heavy metal Pb, Glycine max (L.) Merr.), Soyabean, Growth parameter

Environmental contaminants due to speedup of anthropogenic activities affect nearly every aspect of the ecosystem. Among the contaminant, heavy metals (HMs) is the new emerging concern. Because the persistence bioaccumulative and harmfulness of its nature affect the plants and significantly diminish yield. (Hussain et al 2023). The noxious heavy metals in different valence states include zinc (Zn), arsenic (As), chromium (Cr), cadmium (Cd), mercury (Hg), copper (Cu), nickel (Ni) and lead (Pb), of these lead (Pb) is more pronounced at higher concentrations and durations. Exposure cause oxidative cell damage likes lipids, proteins and nucleic acids and their ultimate result is impaired host metabolism by generating reactive oxygen species (ROS). ROS such as superoxide radical (O<sub>2</sub>), singlet oxygen  $({}_{1}O^{2})$ , hydrogen peroxide  $(H_{2}O_{2})$  and the hydroxyl radical (OH) (Qadir et al 2023). Visible symptoms include chlorotic spots, necrotic lesions etc. in leaf surface, senescence of leaf and stunted growth (Gupta et al 2016). Soybean is largest source of vegetable oil and protein in the worldwide but its yield and nutritional quality adversely affected by heavy metals. There is an urgent need to increase crop productivity for the increasing population (Gupta 2019). Hence, the present study was carried out with an objective to assessment of the hazardous effects of lead on growth of Glycine max (L.) Merr. (soybean).

#### MATERIAL AND METHODS

The experiments were conducted in the greenhouse at University of Rajasthan, Jaipur, during April under natural outdoor conditions with a photoperiod of 12 hours and an average temperature of 30°C. Pots filled with four kilograms of soil at 30 cm height and 25 cm diameter were used, with five lead concentrations (200, 400, 600, 800, 1000 mg/kg) applied as lead acetate. No other supplement nutrients were applied. Pots without added heavy metals constituted were taken as controls. Soybean seeds were surface sterilized with 0.1% mercuric chloride (HgCl<sub>2</sub>) for two minutes and thoroughly washed with distilled water (DW). Ten sterilized seeds of soybean were sown equidistant at 2cm deep in each pot. Three replicates were used for each concentration. Watering was done on alternate days.

To assess plant height, plant root and shoot system was preserved while measuring the length from the soil surface to the highest leaf or flowering axis for shoot length, and from the root tip surface for root length, with measurements recorded in centimeters. Root and shoot dry weights were determined by carefully extracting plants from pots, washing roots to remove soil particles, separating roots and shoots, and subsequently drying them in an oven at 80°C for 48 hours until a constant dry weight was achieved. Statistical analysis employed SPSS ver. 25.0 and Microsoft Office Excel 2016.

#### **RESULTS AND DISCUSSION**

The plant vegetative growth indicators were affected by treatments at pre, peak and post-flowering stages (Table 1). In heavy metal lead treatment at highest concentration i.e. 1000 mg/kg resulted in the highest reduction of all the measure growth parameter at all mentioned stages. It

indicates significant effect on vegetative growth parameters plant height, biomass which significantly decreased from the control plants. The highest reduction in root length (10.41cm), shoot length (21.27cm), root dry weight (0.218gm) and shoot dry weight (1.056gm) compared to untreated control plants root length (28.58cm), shoot length (47.46cm), root dry weight (0.462gm) and shoot dry weight was at pre flowering stage. All studied (2.695gm) parameters of soybean had much higher reduction due to lead treatments in comparison to control (Table 1) The reduction in root length varied from 12 to 63%, shoot length from 8 to 55%, RDW from 11 to 52 % and SDW from 19 to 68% in comparison to control (Table 2). The highest reduction at peak flowering stage in root length (15.80 cm), shoot length (28.02 cm), root dry weight (0.291 gm) and shoot dry weight (1.920 gm) was at 1000 mg/kg as compared to untreated control plants root length (36.02 cm), shoot length (55.94 cm), root dry weight (0.724 gm) and shoot dry weight (4.476 gm). The reduction in root length varied from 8 to 56%, shoot length varied from 7 to 50%, RDW varied from 8 to 59 % and SDW varied from 11 to 57% in comparison to control. At post-flowering stage result found that which highest reduction in root length (21.14 cm), shoot length (37.21 cm), root dry weight (0.375 gm) and shoot dry weight (2.345 gm) was at 1000 mg/kg compared to untreated control plants root length (48.18 cm), shoot length (67.02 cm), root dry weight (0.902 gm) and shoot dry weight (4.741 gm). The reduction in root length varied from 5 to 57%, shoot length varied from 9 to 44%, RDW varied from 21 to 58 % and SDW varied from 8 to 50% in comparison to control.

The growth parameters are important factors used to determine growth performance and crop productivity. This study revealed that under different lead concentrations, growth attributes such as root shoot length and root shoot dry weight were decreased with increasing concentrations of lead as compared to the control at pre, peak and post flowering stages of soybean (*Glycine max* (L.) Merr.) plants. Highest reduction found in these parameters at 1000mg/kg level. Similar finding presented by Li et al (2020) in the lead exposed *Lactuca sativa*. It was demonstrated that radicle growth of lettuce seed was easier inhibited by Pb at sever levels. Pb caused inhibition of cell division in plant's roots.

Table 1. Effect of lead on growth parameters of Glycine max (L.) Merr. at pre, peak and post-flowering stages

Parameter	Stage			Treatments (mg/kg)				Mean	CD (p=0.05)
		0	200	400	600	800	1000		
Root length (cm)	Pre-flowering	28.58	25.23	21.15	17.72	13.67	10.41	19.46	1.686
	Peak-flowering	36.02	33.05	28.39	24.31	19.29	15.80	26.14	
	Post-flowering	48.18	45.71	41.2	34.67	26.33	21.14	36.205	
	Mean	37.59	34.66	30.25	25.57	19.76	15.78	27.26	
	Mean difference between treatment	0	2.93	4.41	4.68	5.81	5.78		
Shoot length (cm)	Pre-flowering	47.46	43.24	36.36	31.21	26.18	21.27	34.29	0.865
	Peak-flowering	55.94	51.67	44.72	40.31	33.19	28.02	42.31	
	Post-flowering	67.02	61.04	58.16	51.32	44.17	37.21	53.15	
	Mean	56.81	51.98	46.41	40.94	34.51	28.83	43.24	
	Mean difference between treatment	0	4.83	5.57	5.47	6.43	5.68		
Root dry weight (gm)	Pre-flowering	0.462	0.410	0.359	0.290	0.258	0.218	0.333	0.0427
	Peak-flowering	0.724	0.662	0.59	0.454	0.373	0.291	0.516	
	Post-flowering	0.902	0.712	0.658	0.512	0.468	0.375	0.604	
	Mean	0.696	0.595	0.536	0.419	0.367	0.295	0.4843	
	Mean difference between treatment	0	0.101	0.059	0.117	0.052	0.072		
Shoot dry weight (gm)	Pre-flowering	2.695	2.179	1.938	1.705	1.331	1.056	1.817	0.1977
	Peak-flowering	4.476	3.987	3.612	3.343	2.204	1.920	3.257	
	Post-flowering	4.741	4.328	4.202	3.523	2.723	2.345	3.644	
	Mean	3.971	3.498	3.251	2.857	2.086	1.771	2.906	
	Mean difference between treatment	0	0.473	0.247	0.394	0.771	0.315		

Parameter	Stage	Treatments (mg/kg)					
		200	400	600	800	1000	
Root length (%)	Pre-flowering	11.72	25.99	37.99	52.17	63.58	
	Peak-flowering	8.02	21.11	32.51	46.45	56.14	
	Post-flowering	5.13	14.87	28.04	45.35	56.12	
Shoot length (%)	Pre-flowering	8.89	23.39	34.24	44.84	55.18	
	Peak-flowering	7.63	20.05	27.94	40.67	49.91	
	Post-flowering	8.92	13.22	23.42	34.09	44.48	
Root dry weight (%)	Pre-flowering	11.25	22.29	37.23	44.15	52.81	
	Peak-flowering	8.56	18.5	37.29	48.48	59.8	
	Post-flowering	21.06	27.05	43.23	48.11	58.42	
Shoot dry weight (%)	Pre-flowering	19.14	28.09	36.73	50.61	60.81	
	Peak-flowering	10.92	19.3	25.31	50.76	57.1	
	Post-flowering	8.71	11.37	25.69	42.56	50.53	

Table 2. Effect of lead on growth parameters (Percent decrease over the control) of *Glycine max* (L.) Merr. var. JS 95-60 at pre, peak and post-flowering stages

Rapid inhibition of root growth occurs probably due to the inhibition of cell division in the root tip or decreased cell expansion in the elongation zone or both of them. Lead toxicity is reported to inhibit growth of several plant species, including *Triticum aestivum* (Kaur et al 2013), Cassia species (Khan MR, 2013), Oryza sativa (Srivastava et al 2014; Ashraf et al 2015) and *Carthamus tinctoriu* (Li et al 2015). It also seems that lead had a toxic effect on shoots elongation and overall development of shoots system as was shown in the current work and confirmed by Sedzik et al (2015), *Hordeum vulgare* (Al-Ghzawi et al 2019). The decrease in root shoot dry weights under Pb toxicity has been reported in sunflower (Saleem et al 2018), *Capsicum annum (*Kaya et al 2019), *Coriandrum sativum* (Fattahi et al 2021) and *Triticum aestivum* (Perveen et al 2022).

#### CONCLUSIONS

The study reveals that lead, significant heavy metal pollutant, exhibits acute toxicity to plants. Increasing lead concentrations adversely affect growth parameters like root length, shoot length, root and shoot dry weight in soybean seedlings. Particularly, the highest lead concentration of 1000 mg/kg demonstrates the most detrimental impact on these parameters. This underscores the importance of informing farmers about soil contamination by heavy metals.

#### REFERENCES

- Ashraf U, Kanu AS, Hussain S, Anjum SA, Khan I, Abbas RN and Tang X 2015. Lead toxicity in rice: Effects, mechanisms, and mitigation strategies. *Environmental Science and Pollution Research* 22(23): 18318-18332.
- Al-Ghzawi ALA, Al Khateeb W, Rjoub A, Al-Tawaha ARM, Musallam I and Al Sane KO 2019. Lead toxicity affects growth and

biochemical content in various genotypes of barley (*Hordeum vulgare* L.). *Bulgarian Journal of Agricultural Science* **25**(1): 55-61.

- Fattahi B, Arzani K, Souri MK and Barzegar M 2021. Morphophysiological and phytochemical responses to cadmium and lead stress in coriander (*Coriandrum sativum* L.). *Industrial Crops and Products* **171**: 113979, https://doi.org/10.1016/j .indcrop.2021.113979
- Gupta S 2019. Antioxidant enzyme activities in Glycine max (l.) Merr. exposed to selective heavy metals stress. Ph.D. Thesis, University of Rajasthan, Jaipur, Rajasthan, India.
- Gupta S, Meena MK and Datta S 2016. Effect of selected heavy metals (lead and Zinc) on seedling growth of soybean *Glycine* max (L.) Merr. International Journal of pharmacy and Pharmaceutical Science 8(8): 302-305.
- Hussain A, Shah M, Hamayun M, Iqbal A, Qadir M, Alataway A, Dewidar AZ, Elansary HO and Lee IJ 2023. Phytohormones producing rhizobacteria alleviate heavy metals stress in soybean through multilayered response. *Microbiological Research* 266: 127237, DOI: 10.1016/j.micres.2022.127237.
- Kaya C, Akram NA, Sürücü A and Ashraf M 2019. Alleviating effect of nitric oxide on oxidative stress and antioxidant defence system in pepper (*Capsicum annuum* L.) plants exposed to cadmium and lead toxicity applied separately or in combination. *Scientia Horticulturae* 255: 52-60.
- Kaur G, Singh HP, Batish DR and Kohli RK 2013. Lead (Pb)-induced biochemical and ultrastructural changes in wheat (*Triticum aestivum*) roots. *Protoplasma* 1: 53-62.
- Khan MR 2013. Effect of heavy metals on seeds germination of some Cassia species. Weekly Science Research Journal 1(13): 1-6.
- Li S, Zhang G, Gao W, Zhao X, Deng C and Lu L 2015. Plant growth, development and change in GSH level in safflower (*Carthamus tinctorius* L.) exposed to copper and lead. *Archives of Biological Sciences* **67**(2): 385-396.
- Li J, Qiu Y, Zhao Q, Chen D, Wu Z, Peng AA, Niazi NK, Trakal L, Sakrabani R, Gao B and Wang H 2020. Lead and copperinduced hormetic effect and toxicity mechanisms in lettuce (*Lactuca sativa* L.) grown in a contaminated soil. *Science of The Total Environment* **741**: 140440, DOI: 10.1016/j.scitotenv. 2020.140440
- Perveen S, Parveen A, Saeed M, Arshad R and Zafar S 2022. Interactive effect of glycine, alanine, and calcium nitrate Ca (NO<sub>3</sub>) on wheat (*Triticum aestivum* L.) under lead (Pb) stress.

Environmental Science and Pollution Research **29**(25): 37954-37968.

- Qadir M, Hussain A, Shah M, Hamayun M and Iqbal A 2023. Enhancement of chromate phytoremediation and soil reclamation potential of *Brassica campestris* L. by *Aspergillus niger. Environmental Science and Pollution Research* **30**(4): 9471-9482.
- Saleem M, Asghar HN, Zahir ZA and Shahid M 2018. Impact of lead tolerant plant growth promoting rhizobacteria on growth, physiology, antioxidant activities, yield and lead content in

Received 10 June, 2024; Accepted 11 September, 2024

sunflower in lead contaminated soil. *Chemosphere* **195**: 606-614.

- Sędzik M, Smolik B and Krupa-Małkiewicz M 2015. Effect of lead on germination and some morphological and physiological parameters of 10-day-old seedlings of various plant species. *Environmental protection and natural resources* **26**(3): 22-27.
- Srivastava R K, Pandey P, Rajpoot R, Rani A and Dubey R S 2014. Cadmium and lead interactive effects on oxidative stress and antioxidative responses in rice seedlings. *Protoplasma* **251**: 1047-1065.



Manuscript Number: 4364 NAAS Rating: 5.38

# Temporal Variation in Water Quality of Dhansiri River, North East India

## Wati lemla, Maibam Romeo Singh and Khikeya Semy<sup>1</sup>

Department of Botany, Nagaland University, Lumami-798 627, India <sup>1</sup>Department of Botany, New Model College, Wakching-798 622, India E-mail: watilemm@gmail.com

**Abstract:** This study focused on understanding how various activities throughout the year affect water quality and contribute to seasonal fluctuations. Three sampling stations were selected for the study, and sixteen physicochemical parameters of water variables were investigated from February 2021 to January 2022. Turbidity in all seasons and TA in winter exceeded the World Health Organizations (WHO 2017) and Bureau of Indian Standards (BIS 2012) drinking water permissible limits. There was significant relationship existing among the sixteen parameters.

Keywords: Dhansiri river, Physicochemical parameters, Permissible limits, Anthropogenic activities, Variable, Correlations

Water is the most abundant natural resource and a prerequisite for all living beings. It is available as surface water (rivers, streams, ponds, and natural springs) and subsurface water (groundwater). Among all the sources, rivers play a crucial role in sustaining livelihood and maintaining various sectors of the economy by providing water supply for drinking and industrial purposes, irrigation, hydropower generation, tourism, infrastructure, and recreational activities (Venkatramanan et al 2014). However, these rivers are continuously being tainted by extensive growth of urbanization and industrial activities, generating large amounts of solid and liquid waste in urban areas and leading to physical, chemical and microbial contamination of river water. The Ganga River near Kanpur was reported by Trivedi et al (2009) to be quite turbid, and significant relationship between total alkalinity and Cl<sup>-</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, and TDS. In contrast, the Himalavan rivers were discovered to be degrading due to eutrophication, tourism, geogenic and anthropogenic factors (Seth et al 2016). Various workers, like Nandal et al (2020) and Rahman et al (2021), reported on water pollution load, stating that industrial discharge and toxicological substances have adversely affected the river. Sarmah et al (2020) concluded that anthropogenic activities are the cause of the decline in water quality. Thus, considering the importance of water, it is imperative to know the status of river water and the activities impacting their quality.

The Dhansiri, with a total catchment area of 1,220 square km is a major river of Chümoukedima and Dimapur district of Nagaland. It flows through 352 km from south to north before joining the Brahmaputra on its south bank. Indigenous populations living along the riverbank rely solely on this river for daily household needs. However, it is subjected to tremendous pressure and is susceptible to contamination by both natural and man-made factors. Furthermore, there are no records of seasonal studies on this river, necessitating water quality monitoring and assessment. Therefore, the objective of this research is to examine the spatiotemporal variation in the river's physicochemical qualities along the stretch of the river, as well as to determine the state of water quality and its usability for the community that relies on it.

#### MATERIAL AND METHODS

**Study area:** Following a thorough assessment of the study area, three sampling sites were chosen for the Dhansiri River water quality investigation. The activities along the riverbed and the pollutants entering the river were considered when selecting the research areas; GPS coordinates and characteristics of sampling points were distributed as follows (Table 2).

**Sampling and analysis:** Water samples were collected for a period of 12 months covering the four seasons: winter (December-February), spring (March-May), summer (June-august), and autumn (September-November) from 2021 to 2022. Approximate 10-20 cm depth was considered for the water sample collection. It was stored in a pre-cleaned plastic bottle, appropriately labelled, and taken to the laboratory in an ice box for subsequent analysis. The sampling, preservation, and transportation were done following standard sampling methods (APHA 1998). Physicochemical analysis of 16 water parameters was done using standard method (Trivedi and Goel 1986, APHA 2012). Pearson

correlation matrix among the water parameters was estimated using statistical software SPSS version 16.0.

#### **RESULTS AND DISCUSSION**

The seasonal values of the sixteen physicochemical parameters are depicted in Table 3.

**pH:** The average pH of the three sites ranged from 7.82-8.07 (winter), 7.65-7.76 (spring), 7.33-7.42 (summer), and 7.41-7.47 (autumn). The seasonal pH study shows a maximum in the winter at SS1, and minimum at SS2 in the summer. Hussain et al (2021) noticed the similar tendency, stating that alkaline pH is caused mostly by carbonate production, microbial respiration at high temperatures, and organic waste deposition during rainfall.

Water temperature: The temperature of river water is dependent on the interaction with environmental factors (Hussain et al 2021). During summer, due to a rise in the atmospheric temperature, the WT was at its maximum, ranging from 29.87°C (SS2) to 30.25°C (SS1) in summer. Meanwhile, with the change in the seasonal pattern of the region, the WT drops to 22.97°C (SS2) to 23.08 °C (SS1) in winter.

**Total dissolved solids:** TDS estimates the amount of organic and inorganic particles in the water sample and displays the quality attributes. The lowest TDS value was recorded in SS2 during winter and highest in summer at all stations; sand mining is common in all three stations, especially in summer. The elevated level of TDS is due to the weathering of rock and the increased flow rate of river water, which causes soil erosion and resuspension of riverbed sediments (Effendi et al 2015).

**Electrical conductivity:** EC measures the capacity of a solution to carry electrical current and indicates the concentration of dissolved ions present in the water. Seasonally, the EC value was recorded highest and lowest in summer (193.38  $\mu$ S/cm) and winter (124±11.66  $\mu$ S/cm) at SS2.

**Turbidity:** The observed mean value of turbidity in winter (8.82-22.62 NTU), spring (14.15-30.03 NTU), summer (29.54-44.67 NTU), and autumn (23.85-33.80 NTU) were within the permissible limit of BIS/WHO. The mixing of suspended particles and activities, including sand mining, recreational fishing, washing, and irrigation, are responsible for the high turbidity level in all the sites.

#### Table 1. Water parameters and its analytical methods

Parameters	Unit	Method
рН		Potentiometric portable digital meter (HM Digital pH- 80)
Water Temperature (WT)	°C	Thermometer
Total Dissolve Solids (TDS)	mg/l	Gravimetric
Electrical Conductivity (EC)	µS/cm	Electrometric method (Portable digital meter)
Turbidity	NTU	Nephelometric method
Chloride (Cl <sup>-</sup> )	mg/l	Titrimetric method (Argentometric method using silver nitrate)
Total Hardness (TH)	mg/l	Titrimetric method (standard solution of EDTA)
Total Alkalinity (TA)	mg/l	Titrimetric method (using indicator methyl orange and phenolphthalein
Dissolve Oxygen (DO)	mg/l	Winkler's titrimetric method
Biological Oxygen Demand (BOD)	mg/l	Winkler's titrimetric method (20°C 5 days incubation)
Potassium (K⁺)	mg/l	Flame photometer
Sulphate (SO₄²)	mg/l	Spectrophotometric (Turbidimetry)
Nitrate nitrogen (NO <sub>3</sub> )	mg/l	Spectrophotometric (Brucine)
Inorganic Phosphorus (PO <sub>4</sub> <sup>3-</sup> )	mg/l	Spectrophotometric (colorimetric)

APHA (2012) and Trivedi and Goel (1986)

Table 2. Desc	cription of th	e three	sampling	sites	and	coordinates

Sampling station	GPS Coordinates	Brief description of sampling points
Chümoukedima (SS1)	25º45′20"N 93º34′37"E	Recreational activities, plastic and food waste from picnics, chemicals fertilizers and pesticides used in agriculture and sand mining.
Dimapur (SS2)	25⁰55′14"N 93⁰44′54"E	Domestic sewage, drainage connections from hospitals and food factories, machinery activities, laundry, washing utensils, and bathing.
Karbi Anglong (SS3)	26º13′06"N 93º50′57"E	Suspended particles from cement factory and brick kilns, debris from religious ritual and oil leakage and urban effluents.

L
ive
Ē
ans
Ĝ
đ
ons
tati
g S
plir
am
se s
thre
he
oft
ies
<i>bert</i>
20 C
<u>g</u>
i U
che
<u>8</u>
hys
le p
금
n ii
iatic
vari
nal
ISSOI
Se
с.
e

Table 3. S	easonal va	rriation in t	he physicc	ochemical <sub>1</sub>	properties	of the thre	e sampling	g stations o	of Dhansiri	river					
Parameters		Winter			Spring			Summer			Autumn		Permi	issible limit	s
	S1	S2	S3	S1	S2	S3	S1	S2	S3	S1	S2	S3	Range	BIS	NНО
Нq	8.1±0.10	7.8±0.05	7 9±0 18	7.7±0.13	7 7±0 13	7 8±0 10	7 3±0 10	7 3±0 15	7 4±0 08	7.7±0.22	7.4±0.21	7 4±0 27	8.1-7.33	6.5-8.5	6.5-8.5
WT (°C)	23.1±1.43	22.9±1.52	23.1±1.73	27 1±0 69	27 1±0 7	27±0.76	30.2±0.48	29.9±0.57	29.7±0.48	28.3±1.21	28.7±0.99	28.1±1.09	22.9-30.2	NA	AN
TDS (mg/l)	72.78±8.7	71.60±6	81.9±3.99	82.8±11.5	82.8±11.5	84.7±6.90	109.1±11.2	119.4±5.54	124 9±10 3	108.6±9.8	96.3±12.23	117.2±9.7	71.6-124.9	500	600
EC (µS/cm)	126.8±10.9	124 6±11 7	164.6±7.8	147 9±14 7	147 9±14 7	164.4±10	178 5±11 4 ′	193 4±7 76	186±9.84	164.6±9.7	167 1±10 3	161.5±12	124.6-193.4	NA	AN
Turbidity (NTU)	8.8±0.58	22.6±1.33	22.4±1.32	14.1±1.77	14.1±1.77	30±2.26	29.5±4.74	42.1 <u>±</u> 4.21	44.7±4.40	23.8±5.29	31.1±3.94	33.8±6.03	8.8-44.7	5	5
Cl <sup>-</sup> (mg/l)	9.5±0.82	13.7±0.82	11.8±1.64	14±1.77	14±1.77	19 5±4 23	19.9±2.84	21.2±2.85	21 8±3 28	12.8±2.42	16.1±3.28	12.2±4.39	9.5-21.8	250	250
TH (mg/l)	111.7±7.51	106.1±9.87	109 1±7 87	86.5±8.98	86.5±8.98	82.9±10	52.9±6.73	57±2.85	60.7±8.54	73.6±13.2	65.7±1.37	76.9±8.52	52.9-109.1	200	200
TA (mg/l)	124.6±5.51	114 4±6 68	132.5±10.5	105.5±8.56	105.5±8.56	118.3±5.7	73.9±13.04	77 3±2 43	82.8±9.43	76 2±8 46	89.6±7.79	91.8±10.3	73.9-132.5	120	200
Ca <sup>2+</sup> (mg/l)	19.5±3.24	19.5±2.01	17.4±2.45	11.2±3.67	11.2±3.67	13.1±2.45	9.6±1.38	10.2±1.68	9.6±1.6	12.8±4.16	13.6±2.12	13.1±1.67	9.6-19.5	75	75
Mg <sup>2+</sup> (mg/l)	15.3±1.01	14.6±0.84	15.1±1.29	9.9±4.42	9.9±4.42	11.2±3	6.7±1.22	7.8±0.97	8.1±0.74	7 3±1 46	7.6±3.25	7.5±2.94	6.7-15.3	30	50
DO (mg/l)	7.9±0.61	8.6±1.52	7.1±0.60	6.4±1.06	6.4±1.06	6.9±1.29	5.6±1.13	5±0.35	5.3±0.42	8.3±0.61	7.6±1.06	7.4±0.23	5-8.59	5	AN
BOD (mg/l)	4.5±0.31	5.3±0.51	4.3±0.28	3.1±0.61	3.1±0.61	3.6±0.08	3.4±1.25	2.8±0.52	3.1±0.21	3.5±0.17	3.7±0.34	3.9±0.23	2.8-5.3	5	5
K⁺ (mg/l)	2.9±0.77	3.6±0.42	3.1±0.76	4±0.87	4±0.87	4.5±0.52	6.1±0.19	6.1±0.44	6.6±1.01	4.1±0.71	4±0.27	4.4±0.41	2.9-6.6	NA	AN
$SO_4^{2}$ (mg/l)	7.9±0.74	9.9±0.47	10.3±0.51	8.7±1.01	8.7±1.01	11.4±0.96	11.2±0.79	15.6±1.19	14.3±1.11	10.1±0.19	11.7±1.57	12.4±0.66	7.9-15.6	200	250
NO <sup>3</sup> (mg/l)	0.05±0.03	0.14±0.04	0.2±0.01	0.1±0.08	0.1±0.08	0.3±0.09	0.3±0.09	0.5±0.06	0.5±0.09	0.2±0.09	0.3±0.07	0.3±0.06	0.05-0.5	45	50
PO <sub>4</sub> <sup>2-</sup> (mg/l)	0.14±0.02	0.15±0.03	0.2±0.05	0.2±0.03	0.2±0.03	0.2±0.03	0.2±0.05	0.2±0.02	0.3±0.07	0.2±0.07	0.2±0.02	0.2±0.06	0.1-0.3	AN	ΝA

**Chloride:** Chloride is naturally available in all types of water, and when it is present in high concentrations, it is considered an indicator of organic, inorganic, and industrial pollution (Das and Semy 2023). The presence of high value of chloride in water is harmful to irrigation and aquatic life (Venkatesharaju et al 2010). Among all the seasons, CI<sup>-</sup> was highest in SS3 during summer (21.77 mg/l) and least at SS1 in winter (9.47 mg/l). During summer, the raised value of chloride in river water may be due to rainwater washing off municipal sewages and domestic waste into the river (Singh and Shrivastava 2015).

**Hardness:** Most freshwaters primarily contain calcium and magnesium ions (Kumar et al 2011). The observed values of TH in the three stations ranged from 52.88-111.46 mg/l. The seasonal  $Mg^{2^+}$  and  $Ca^{2^+}$  level in SS1, SS2 and SS3 ranged from 6.66-15.27 mg/l and 9.62-19.5 mg/l, respectively. The results of the seasonal analysis of the parameters TH,  $Ca^{2^+}$ , and  $Mg^{2^+}$  revealed that the highest levels were seen in the winter and the lowest during the summer, and that the values obtained were within the acceptable range of the BIS/WHO standard.

**Total alkalinity:** Alkalinity in water is due to carbonates, bicarbonates, or hydroxides from limestone, salts, dissolved rocks, and sediments (Kumar et al 2012). The average TA value obtained from all the sampling stations during the four seasons ranged from 73.95-132.50 mg/l. TA exceeded desirable limit of BIS/WHO during the winter. The value decreased in the summer and autumn, which according to Singh et al (2004) is caused due to excessive dilution by rain water.

**Dissolved oxygen:** It measures oxygen dissolved in water, and the concentration depends on its physical, chemical, and biological activities along the river (Bora and Goswami 2017). The maximum concentration of DO was observed in winter, 7.05 (SS3) - 8.59 (SS2), which may be attributed to the low temperature in winter resulting in increased concentration of DO. Meanwhile, the minimum DO concentration in summer (5.04 -5.63mg/l) could be attributed to high microbial activities due to organic waste in the river water (Venkatesharaju 2010).

**Biochemical oxygen demand:** The presence of a high level of BOD in the water indicates the presence of organic waste and active microbial respiration in the river water (Semy and Singh 2019). The BOD in Dhansiri River ranged from 3.06-5.3 mg/l, with the highest value in winter (SS2) and the lowest concentration during summer (SS3). The rise of BOD could be attributed to high microbial activities triggered by organic pollution from the discharge of domestic waste and runoff from farmland and residential and commercial areas.

Potassium: In winter, the lowest K<sup>+</sup> value was 2.97 mg/l at

SS1, while in summer, the highest value was 6.61 mg/l at SS3. Agricultural practices along the river stretch contribute to a rise in nutrient levels downstream. Semy and Singh (2019) have reported that the increase in  $K^+$  levels in summer corresponds to nutrient runoff from cultivated lands that reach river bodies.

**Sulphate:** Sulphate is naturally found in river water in minute concentration or it can also occur from anthropogenic activities like mining (Titilawo et al 2019). The average  $SO_4^{2^-}$  value in river water throughout the sampling stations varied from 7.94 mg/l (winter) to 15.58 mg/l (summer).

**Nitrate:** The seasonal NO<sub>3</sub> concentration fluctuated from 0.05- 0.52mg/l, which was under the permissible limit of WHO/BIS. As per observation in the sampling stations, NO<sub>3</sub> value increased from SS1 to SS3 throughout the seasons, presumably due to the influx of fertilizers from the agricultural land, which resonates with Meher et al (2015).

**Inorganic phosphorus:** The  $PO_4^{3-}$  concentration was maximum in autumn at SS3 (0.27 mg/l) and minimum in winter at SS1 (0.14 mg/l). The use of fertilizers and pesticides was observed at all sampling sites. Similar to the current findings, Semy and Singh (2021) stated that the extensive use of fertilizers and pesticides in the agricultural fields along the river banks increases  $PO_4^{3-}$  concentration during the premonsoon period.

Correlation analysis among water physicochemical parameters: pH is negatively correlated with WT, TDS, turbidity,  $K^*$ ,  $SO_4^{2-}$  and  $NO_3^{-}$ , while positively significant with TH, TA, Ca2+, Mg2+ and BOD indicating that an increase or decrease in the pH value will also exhibit change in increasing or decreasing of these water parameters (Table 4). The EC was positively significant with turbidity, K<sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, and NO<sub>3</sub><sup>-</sup> meanwhile, negatively significant (p<0.01) with TH, TA, Ca<sup>2+</sup>, Mg<sup>2+</sup>, DO, and BOD. A significant positive correlation is observed between Ca<sup>2+</sup> and TA, which was also reported by Hussain et al (2021) due to the presence of carbonates, bicarbonates, and hydroxide in river water. Likewise, turbidity level showed a negative correlation with TH, TA, Ca<sup>2+</sup> and Mg<sup>2+</sup>, DO and BOD but positively significant with Cl<sup>-</sup>, K<sup>+</sup>, SO<sub>4</sub><sup>2-</sup> and NO<sub>3</sub>. TH is positively significant at p<0.01 with TA,  $Ca^{2+}$ , Mg2+, and BOD. A similar positive correlation was also observed between TH, TA and BOD (Rahman et al 2021) and TH and Mg<sup>2+</sup> (Bhandari and Nayal 2008). DO shows a negative correlation with WT at p<0.05 indicates that oxygen decreases with increasing temperature. PO43- did not show any significant relationship with other parameters. Some significant negative correlations were noted between pH, TA, Ca<sup>2+</sup> Mg<sup>2+</sup>, DO and BOD, which was consonant with the observation of Titilawo et al (2019). The correlation analysis reveals inter-relationships existing among the physical and

Table 4. Pearson's correlation coefficient matrix for the physicochemical water quality parameters

	pН	WT	TDS	EC	Turb	Cl	TH	TA	Ca <sup>2+</sup>	$Mg^{2+}$	DO	BOD	K⁺	SO4 <sup>2-</sup>	$NO_3^-$	PO4 3-
pН	1															
WT	-0.902**	1														
TDS	-0.953	0.895**	1													
EC	-0.908**	0.894**	0.943	1												
Turb	-0.915	0.909**	0.947	0.977**	1											
Cl	-0.593 <sup>*</sup>	0.717**	0.567	0.661 <sup>*</sup>	0.723**	1										
тн	0.933**	-0.981**	-0.919 <sup>**</sup>	-0.938**	-0.941**	-0.752**	1									
TA	0.950	-0.959	-0.961	-0.927**	-0.933	-0.651 <sup>*</sup>	0.976**	1								
Ca	0.790**	-0.894**	-0.740"	-0.781**	-0.802**	-0.861**	0.920**	0.845**	1							
Mg	0.846**	-0.866**	-0.802 <sup>**</sup>	-0.814**	-0.839	-0.553	0.890**	0.891**	0.815	1						
DO	0.516	-0.628	-0.532	-0.747**	-0.710 <sup>***</sup>	-0.743**	0.690*	0.556	0.726**	0.548	1					
BOD	0.773	-0.847**	-0.702 <sup>*</sup>	-0.785**	-0.820	-0.818**	0.862**	0.798**	0.856**	0.771 <sup></sup>	-0.728**	1				
K⁺	-0.760	0.831 <sup>**</sup>	0.753 <sup>**</sup>	0.871 <sup>**</sup>	0.889	0.870	-0.864	-0.772 <sup>***</sup>	-0.870	-0.702	-0.853	-0.854	1			
SO4 2-	-0.859**	0.878**	0.893	0.966**	0.974**	0.777 <sup>**</sup>	-0.932**	-0.903**	-0.840**	-0.825	-0.804**	-0.856 <sup>**</sup>	0.913	1		
NO <sub>3</sub> <sup>-</sup>	-0.867**	0.867**	0.916 <sup>**</sup>	0.966**	0.990**	0.721 <sup>**</sup>	-0.906**	-0.902**	-0.752 <sup>***</sup>	-0.808**	-0.713 <sup>**</sup>	-0.804	0.871 <sup>***</sup>	0.975**	1	
PO <sub>4</sub> <sup>3-</sup>	-0.392	0.505	0.503	0.495	0.481	0.297	-0.486	-0.536	-0.452	-0.497	-0.334	-0.411	0.433	0.465	0.461	1

\*\*Correlation is significant at the 0.01 level (2-tailed) \*Correlation is significant at the 0.05 level (2-tailed)



Fig. 1. Drainage area map of the three-sampling location of Dhansiri river

chemical parameters of the Dhansiri river which has its impact on the function of the river system.

#### CONCLUSION

The physicochemical parameters of the three stations fluctuated seasonally, and the parameters WT, TDS, EC, turbidity, and Cl<sup>-</sup>were high during summer while the variables TH, TA, Ca<sup>2+</sup>, and Mg<sup>2+</sup>escalated during winter months. The anthropogenic activities, runoff waste debris, and other organic waste from the landfill areas have significantly impacted the water quality. The TA and turbidity of all three sampling locations were above the WHO/BIS permissible limit, indicating that the river water is unsuitable for use without proper treatment.

#### REFERENCES

- APHA 1998. Standard Methods for the Examination of Water and Wastewater. 20th Edition, American Public Health Association, Washington DC.
- APHA 2012. Standard Methods for Examination of Water and Wastewater. 22 Edition, American Public Health Association, Washington DC, USA.
- Bhandari NS and Nayal K 2008. Correlation study of physicochemical parameters and quality assessment of Kosi River water Uttarakhand. *E-Journal of Chemistry* **5**(2): 342-346.
- Bora M and Goswami DC 2017. Water quality assessment in terms of water quality index (WQI): Case study of the Kolong River, Assam, India. *Applied Water Science* **7**(6): 3125-3135.
- Das M and Semy K 2023. Monitoring the dynamics of acid mine drainage affected stream surface water hydrochemistry at Jaintia Hills, Meghalaya, India. *Environmental Science and Pollution Research* **30**(30): 75489-75499.

#### Wati lemla et al

- Effendi H, Romanto and Wardiatno Y 2015. Water quality status of Ciambulawung River, Banten Province, based on pollution index and NSF-WQI *Procedia Environmental Sciences* **24**: 228-237.
- Hussain M, Jamir L and Singh MR 2021. Assessment of physicochemical parameters and trace heavy metal elements from different sources of water in and around institutional campus of Lumami, Nagaland University, India. *Applied Water Science* **11**(4). DOI: 10.1007/s13201-021-01405-5.
- Kumar RN, Solanki R and Kumar N 2011. An assessment of seasonal variation and water quality index of Sabarmati River and Kharicut canal at Ahmedabad, Gujarat. *Electronic Journal of Environmental, Agricultural and Food Chemistry* **10**(5): 2248-2261.
- Kumar SK, Chandrasekar N, Seralathan P, Godson PS and Magesh NS 2012. Hydrogeochemical study of shallow carbonate aquifers, Rameswaram Island, India. *Environmental Monitoring* and Assessment **184**(7): 4127-4138.
- Meher PK, Sharma P, Gautam PY, Kumar A, Mishra KP, Nagarani N, Arumugam K, Kumaraguru VJ, Devi C and Archana D 2015. Evaluation of water quality of Ganges River using water quality index tool. *Environment Asia* 8(1): 124-132.
- Nandal A, Kaushik N, Yadav SS, Rao AS, Singh N and Gulia SS 2020. Water quality assessment of pond water of Kalanaur Block, Rohtak, Haryana. *Indian Journal of Ecology* 47(1): 1-6
- Rahman A, Jahanara I and Jolly YN 2021. Assessment of physicochemical properties of water and their seasonal variation in an urban river in Bangladesh. *Water Science and Engineering* **14**(2): 139-148.
- Sarmah R, Dutta R, Bhagabati SK, Nath D, Mudoi LP, Pokhrel H and Ahmed AM 2020 Seasonal variation of water quality parameters of river Dikhow in Nagaland and Assam. *International Journal of Chemical Studies* **8**(5): 1429-1434.
- Semy K and Singh MR 2019. Assessment on the water quality of Tsurang River, Nagaland affected by coal mining drainage.

Received 01 July, 2024; Accepted 01 October, 2024

Indian Journal of Ecology 46(4): 845-849.

- Semy K and Singh MR 2021. Quality assessment of Tsurang River water affected by coal mining along the Tsurangkong Range, Nagaland, India. *Applied Water Science* **11**(7).
- Seth R, Mohan M, Singh P, Singh R, Dobhal R, Singh KP and Gupta S 2016. Water quality evaluation of Himalayan Rivers of Kumaun region, Uttarakhand, India. *Applied Water Science* 6(2): 137-147.
- Singh KP, Malik A, Mohan D and Sinha S 2004. Multivariate statistical techniques for the evaluation of spatial and temporal variations in water quality of Gomti River (India): A case study. *Water Research* **38**(18): 3980-3992.
- Singh PK and Shrivastava P 2015. Analysis of water quality of river Narmada. *International Journal of Current Research* **7**(12): 24073-24076.
- Titilawo Y, Akintokun A, Shittu O, Adeniyi M, Olaitan J and Okoh A 2019. Physicochemical properties and total coliform distribution of selected rivers in Osun state, Southwestern Nigeria. *Polish Journal of Environmental Studies* 28(6): 4417–4428.
- Trivedy RK and Goel PK 1986. Chemical and Biological method for water pollution studies. *Environmental Publication* **6**: 10-12.
- Trivedi P, Bajpai A and Thareja S 2009. Evaluation of Water Quality: Physicochemical Characteristics of Ganga River at Kanpur by using Correlation Study. *Nature and Science* **1**(6). http://www.sciencepub.net
- Venkatesharaju K, Ravikumar P, Somashekar RK and Prakash KL 2010. Physicochemical and bacteriological investigation on the river Cauvery of Kollegal stretch in Karnataka. *Kathmandu* University Journal of Sciences, Engineering and Technology 6(1): 50-59.
- Venkatramanan S, Chung SY, Lee SY and Park N 2014. Assessment of river water quality via environmentric multivariate statistical tools and water quality index: A case study of Nakdong River Basin, Korea. *Carpathian Journal of Earth and Environmental Sciences* **9**(2): 125-132.



Manuscript Number: 4365 NAAS Rating: 5.38

## Premonsoon Groundwater Trends for Nanjangud Taluk, India: A Spatio-Temporal Analysis

### M.C. Manjunatha

DBT-BUILDER, JSS AHER, Sri Shivarathreeswara Nagar, Mysuru-570 015, India E-mail: mcmanju1@gmail.com

**Abstract:** Groundwater is a crucial natural resource that plays a vital role in providing drinking water and supporting various industries and ecosystems. The study investigates the trends in premonsoon groundwater levels in southern Karnataka state, focusing on understanding fluctuations before the monsoon season (Jan-May). The analysis reveals a predominant decreasing trend in groundwater levels, with varying rates of decline observed across monitoring wells. The spatial distribution of groundwater trends was analysed by statistical methods and IDW tool of GIS software. More than 46% of the studied region indicated a declining trend and falling from south-east to eastern parts between 2003 and 2019. The output results contribute to a comprehensive understanding of premonsoon groundwater dynamics and their implications for water resource management by analyzing decadal fluctuations in groundwater depths.

#### Keywords: Premonsoon, Groundwater trends, Nanjangud, GIS

Groundwater is essentially important for rural communities, where about 90% of drinking water is sourced from groundwater (Dubey et al 2022). It is estimated that as much as 50% of the global population relies on groundwater for their drinking water needs, with approximately 43% of all water used for irrigation coming from groundwater sources (Johnson et al 2022, Van Loon et al 2024). This hidden water source, stored in aquifers beneath the Earth's surface, makes up 99% of the Earth's liquid fresh water and significantly contributes to the water cycle, impacting rivers, lakes, wetlands, and other surface water bodies (Poeter et al 2020). Hot spots of groundwater depletion have been identified in arid and semiarid regions worldwide, with significant depletion has been observed in countries like India, the United States, China, Iran, Mexico, and Saudi Arabia, which collectively account for a substantial portion of global annual water extraction. This issue poses challenges such as reduced flows in streams and wetlands, land subsidence, water quality degradation (Jakeman et al 2016), and increased pumping costs, highlighting the urgent need for sustainable groundwater management practices on an international scale. Groundwater also plays a critical role in stabilizing water levels in rivers, lakes and wetlands during drier months, providing a sustainable water source for wildlife and plants in these environments.

The spatial and temporal analyses of rainfall and groundwater levels show a declining trend in the premonsoon groundwater levels (Halder et al 2021), with rates of decrease varying across wells in Nanjangud taluk. The decrease in pre-monsoon groundwater levels can have adverse effects on irrigation practices in the taluk, as it reduces the availability of water for agricultural purposes, impacting crop growth and productivity. Additionally, declining groundwater levels can affect domestic water supply, leading to water scarcity for households relying on groundwater sources for daily use. Sustainable management of groundwater resources is crucial to maintain a balance between water availability and demand for various purposes, especially in regions where groundwater serves as a primary source of water for agriculture, drinking, and industrial activity (Li et al 2021).

Research has shown that premonsoon water levels can vary significantly between years, impacting water availability for various uses such as agriculture, drinking water, and industry (Vaibhav Deoli and Deepak Kumar 2020). Monitoring these trends is essential for assessing changes in water availability, especially in regions heavily reliant on groundwater. Factors like rainfall patterns, land use changes, and groundwater pumping practices play a significant role in influencing premonsoon groundwater trends. Sustainable management of groundwater is paramount, especially with the expected increase in the global population by 2100, as overexploitation and pollution threaten this finite resource. Historically the groundwater was mostly used by agriculture and irrigational activities in the taluk earlier, but since last few decades its utilization was noticed by various industrial and factories observed next to Perennial River Kapila. This would immensely impact on groundwater resource management in the vicinity near future. The present aims to understand the dynamics of premonsoon groundwater levels using GIS tool

in Nanjangud taluk mainly focusing on the long-term groundwater sustainability.

#### MATERIAL AND METHODS

**Description of the Study area:** The taluk of Nanjangud is measured an area of 983.95 sq km and situated between 72° 26' and 76°56' E longitude and 11°51' and 12°13' N latitude (Fig. 1). The general altitude is 600-700 ft above MSL covering 184 villages (Manjunatha and Basavarajappa 2021). With the exception of a few scattered hillocks to the south and west, this forms practically a plain boundary. The highest point, 3111 ft above MSL, is located in the southwest corner and the overall slope is north to south, with a narrow but gradually expanding depression located in the Kapila river basin.

**Data used and analysis:** The taluk boundary was freely downloaded from K-GIS, Govt. of Karnataka website and the groundwater level data was acquired from District Groundwater Board, Mysuru, Govt. of Karnataka for a period of 16 years (2003-19) from 10 observation groundwater well points (Table 1). Spatio-temporal maps were generated by calculating depths, and drawing contours using methods like Inverse Distance Weighted (IDW) tool in ArcGIS software (Manjunatha et al 2023). IDW tool was processed on groundwater level analyses (meters, bgl) for pre-monsoon seasons (Jan-May) on four year fluctuation (2003-07, 2007-11, 2011-15, 2015-19) and long-term fluctuation study (2003-19). This study specifically focusing on premonsoon periods between Januarys to May to avoid seasonal recharge effects from rainfall.

#### **RESULTS AND DISCUSSION**

**Groundwater trend analysis:** The fluctuations of groundwater level yield a precise geological formation that increases the point value of recharge, which is computed from the rise in a well's water level. Over a 16-year period

(2003-2019), 10 representative observation bore well points were taken into consideration in order to analyse the fluctuations in groundwater levels from January to May (premonsoon season) (Table 1, Fig. 2) (Manjunatha et al 2020). The deeper water levels were recorded as 29.96 m at Hanumanapura observation well measured below ground level (bgl). Groundwater levels were showing declining patterns from the south-eastern (2003) to east (2019) (Table 3, Fig. 3).

Over-exploitation of subsurface water through pumping wells was noticed in eastern parts contributing to groundwater declining levels (Table 2, Fig. 2). Bar graphs of groundwater data revealed the groundwater trend analyses over a period of 16 years (2003-19) (Table 1, Fig. 4). Prominent lineament structures like dykes and fractures are noticed along the NS, NE-SW, and NW-SE directions that control regional subsurface water flow and help in natural recharge of groundwater along various streams, rivers and ponds. More than 46% (461.92 sq km) of the study area showed declined trend from 2003-2019 in groundwater depth (Table 4). Most parts of eastern Nanjangud showed higher demands of groundwater resources, especially for domestic and agricultural activities during summer months. Inclining trend of groundwater levels in all observation well points were observed due to flash floods and heavy rainfall occurred during August 2018 in Wayanad region of Kerala state (Manjunatha and Basavarajappa 2022). More water intensive crops such as paddy, cotton, sugarcane and others could be avoided in critical and over-exploited areas. Artificial recharge structures (ARS) are the best techniques to recharge aquifers within town limits for reducing the load on urban water supply systems. Roof top Rain Water Harvesting (RRWH) structures would be most suitable for larger industrial structures to overcome their demand of groundwater.

Premonsoon groundwater fluctuations in India are

 Table 1. Average pre-monsoon groundwater level meters (2003-2019)

J J J	5		<b>(</b>	- /			
Station name	Latitude	Longitude	2003	2007	2011	2015	2019
Deburu	12.11385	76.61656	5.24	5.37	4.81	4.52	4.14
Hanumanapura	12.06005	76.82569	0.4	13.21	21.28	29.96	27.27
Hullahalli	12.09455	76.55575	12.41	11.2	9.61	9.22	0.1
Hura	12.00258	76.54400	9.27	5.94	7.97	11.73	9.93
Kalale	12.07446	76.66335	9.26	7.84	6.86	7.7	5.31
Kothanahalli	11.90696	76.53156	6.03	6.05	7.02	7.07	4.45
Kowlande	12.03232	76.79744	17.33	19.36	18	13.09	13.97
Nanjangud	12.11626	76.67928	3.94	2.65	3.41	3.32	2.66
Sindhuvallipura	12.03095	76.63965	10.38	12.03	10.74	12.73	9.28
Thagaduru	12.09297	76.80928	0.7	17.95	7.22	17.16	23.3

Premonsoon Groundwater Trends



Fig. 1. Groundwater well points map of Nanjangud taluk



Fig. 2. Premonsoon Groundwater Depths maps for the years (a) 2003, (b) 2007, (c) 2011, (d) 2015 and (e) 2019



Fig. 3. Premonsoon Groundwater level fluctuation maps for various years (a) 2003-07, (b) 2007-11, (c) 2011-15, (d) 2015-19 and Long-term fluctuation for (e) 2003-19

influenced by various factors, including climate conditions, human activities, and geological characteristics (Vinod Kumar and Vipin Kumar 2020). Studies indicates that around 23% of areas in India experienced a decline in groundwater levels from 1996 to 2016 during the premonsoon period (Gopal Krishan et al 2015). Groundwater based study is a much necessary task in major towns of Karnataka due to many uprising issues of rapid increase of population, water supply & demand in various fields of major industries, factories, urbanization, mining areas and others (Urvashi Sharma et al 2020). Recent studies indicated that some northern Karnataka regions experienced a rise in groundwater levels, while southern regions often face declines. The depth of groundwater levels during the premonsoon season has been recorded between 7 to 22 mts bgl in various districts (Surender Kumar et al 2020). Insufficient premonsoon rainfall often leads to a decline in water levels due to increased evaporation and higher extraction for irrigation purposes. Agricultural practices play a

pivotal role in groundwater dynamics. The extensive use of tube wells for irrigation contributes to the depletion of groundwater resources, particularly in regions dependent on monsoon rains for crop cultivation. For instance, fluctuations can range dramatically from below 2-6 mts depending on local geological conditions and rainfall distribution.

The final output maps were generated in analyzing the pre-monsoon groundwater trend using a period of 16 years (2003-19) of observation bore-well point's data. The groundwater levels of Nanjangud taluk were heavily being withdrawn through pumping wells in eastern parts due to the existence of various major, medium and minor industries contributing to groundwater decline. The presence of numerous industries in the area, such as distilleries and other manufacturing units, lead to the discharge of effluents and sewage directly into the Kapila River, which flows through the industrial zones. This pollution contributing to an increase in nutrients in the sediments, affecting the overall water quality and ecosystem health (Mohammad Amin et al 2016).



Fig. 4. Bar graph depicting groundwater fluctuations for various years (2003-19)

	•		1.00				•	
Inhin 7	Aroa	undor	dittoront	aroundwotor	donth	70000 10	VOLIDIA	loarc
	AIEA	UTICIET	umerem	UTUTUVVALET	UEUIII	701165 111	VALIDITS	veals
	,	anaor		grounditation	aopai	201100 111	10110000	,

Depth of groundwater levels (m, bgl)	2003	2007	2011	2015	2019
0-5	169.76	20.72	21.46	18.89	181.93
5 – 10	719.73	514.81	657.96	403.13	422.58
10 - 15	87.84	333.69	203.83	344.58	130.87
15- 20	6.62	114.73	96.67	154.82	146.06
>20	Nil	Nil	4.03	62.53	102.51

Table 3. Area un	der different groun	dwater depth fluctu	ation in various years
	J		, <u> </u>

Fluctuation in depth to		Area	(km²)		Trends in
water levels (m, bgl)	2003 – 07	2007 – 11	2011 – 15	2015 – 19	fluctuation
<4.0	-	45.68	2.91	95.38	Inclination
0.0 - 4.0	365.47	637.07	61.47	658.13	
-4.0 - 0.0	282.71	289.38	732.33	216.29	Declination
-8.04.0	147.62	11.69	163.57	14.15	
> -8.0	188.15	0.13	23.67	-	

Fluctuation in depth to water level (m, bgl)	2003 - 19	Trend in fluctuation
<4.0	91.27	Inclination
0.0 - 4.0	430.76	
4.0 - 0.0	137.59	Declination
-8.04.0	78.97	
> -8.0	245.36	

Table 4. Area under long-term fluctuation from 2003 to 2019

Geospatial approach showed a better illustrations in continuous monitoring of groundwater levels using observation wells which is crucial for assessing trends over time. This has been very critical for successful analysis, prediction, validation and involves developing models that can guide environmental and socioeconomic planning related to groundwater resources.

#### CONCLUSIONS

Nanjangud taluk showed a wide variation of 29 mts in groundwater fluctuations at Hanumanapura observation well point due to natural and anthropogenic activities. Industrial activities in Nanjangud taluk showed a significant impact on groundwater quality and availability. Integrating GIS tools with statistical groundwater data would better portray the groundwater dynamics, availability and sustainability especially for the industrial town in India. Regular analysis provides valuable insights for policymakers to improve groundwater levels, aquifer-recharge potential, and regular monitoring of over-dependency regions of groundwater resources across the state. Implementing GIS based monitoring, mapping high-potential areas, community engagement and integrated water resource management can help in predicting water availability, and efficient water use practices to create a comprehensive groundwater management plan. The outcome of this study can help the farmers to take necessary steps in their crop rotation and cultivation who depend only on shallow groundwater levels. Utilizing GIS techniques in Nanjangud taluk offers a robust framework for understanding and managing premonsoon groundwater fluctuations. By focusing on aquifer characteristics, drainage patterns, and potential zones identified through GIS, stakeholders can make informed decisions to ensure sustainable groundwater use in the region.

#### REFERENCES

Dubey SK, Lal P, Choudhari P, Sharma A and Dubey AK 2022. Assessment of long-term groundwater variation in India using GLDAS reanalysis. In: Advances in Remediation Techniques for Polluted Soils and Groundwater Elsevier: 219-232.

Gopal Krishan, Singh RP and Tashi KS 2015. Water level fluctuation

Received 12 July, 2024; Accepted 20 September, 2024

as the sum of environmental and anthropogenic activities in Southeast, Punjab (India). Journal of Environmental & Analytical Toxicology 5(5): 1-7.

- Halder S, Lingaraj Dhal, and Madan KJ 2021. Investigating groundwater condition and seawater intrusion status in coastal aquifer systems of Eastern India. Water 13: 1-15.
- Jakeman AJ, Barreteau O, Hunt RJ, Rinaudo JD, Ross A, Muhammad Arshad, and Hamilton S 2016. Integrated groundwater management: An overview of concepts and challenges. In Integrated Groundwater Management, Cham: Springer International Publishing: 3-20.
- Johnson TD, Belitz K, Kauffman LJ, Watson E and Wilson JT 2022. Populations using public-supply groundwater in the conterminous U.S. 2010; Identifying the wells, hydrogeologic regions, and hydrogeologic mapping units. Science of the Total Environment 806(2): 150618.
- Li P, Karunanidhi D, Subramani T and Srinivasamoorthy K 2021. Sources and consequences of groundwater contamination. Archives of Environmental Contamination and Toxicology 80(1): 1 - 10.
- Manjunatha MC and Basavarajappa HT 2022. 2019-Flashflood impacts of Kapila River on Temple town of Nanjangud, Karnataka, India. International Advanced Research Journal in Science, Engineering and Technology 9(7): 359-368.
- Manjunatha MC and Basavarajappa HT 2021. Land classification analysis using geospatial approach in Nanjangud taluk of Karnataka state, India. International Advanced Research Journal in Science, Engineering and Technology 8(6): 629-638.
- Manjunatha MC, Maruthi NE, Siddaraju MS and Basavarajappa HT 2020. Pre-monsoon groundwater trend analyses in Mysuru Taluk of Karnataka State, India Using Geospatial Technology. Journal of Chemical, Biological, and Physical Sciences 11(1): 51-61
- Manjunatha MC, Siddaraju MS, Abrar Ahmed and Basavarajappa HT 2023. Application of geospatial mapping in the analyses of pre-monsoon groundwater fluctuation: A case study of Piriyapatna Taluk of Karnataka State, India. Journal of Global Ecology and Environment 17(1): 26-32.
- Mohammad Amin B, Grewal MS, Ramprakash, Rajpaul Sheeraz AW and Eajaz Ahmad D 2016. Assessment of Groundwater quality for irrigation purposes using Chemical indices, Indian Journal of Ecology 43(2): 574-579.
- Poeter E, Fan Y, Cherry J, Wood W and Mackay D 2020. Groundwater in Our Water Cycle: Getting to Know Earth's Most Important Fresh Water Source. The Groundwater Project. E-ISSN: 978-1-7770541-1-3.
- Surender Kumar, Mamta Rani, Anurag and Shubham Verma. 2020. Assessment of pre-monsoon and monsoon groundwater level and fluctuation at regional level in South of Haryana; reference to Rewari District, International Journal of Applied Science and Engineering 8(2): 123-129.
- Urvashi Sharma, Adeeba and Venkatesh Dutta 2020. Impact of declining groundwater levels on river flows in the Ganga Alluvial Plain: A case study of Gomti River, India. Indian Journal of Ecology 47(1): 40-48.
- Vaibhav Deoli and Deepak Kumar 2020. Analysis of groundwater fluctuation using GRACE satellite data, Indian Journal of Ecology 47(2): 299-302.
- Van Loon AF, Wanders N, Bloomfield JP, Fendeková M, Ngongondo C and Van Lanen HAJ 2023. Human influence. In: Hydrological Drought: Processes and Estimation Methods for Streamflow and Groundwater, 2<sup>nd</sup> Edition, Elsevier: 479-524.
- Vinod Kumar and Vipin Kumar 2020. Spatial analysis of groundwater fluctuation in Nathusari Chopta Block of Sirsa district (Harvana, India) using geospatial technology. Indian Journal of Ecology 47(2): 308-311.



# Spatiotemporal Variation in Litter Bacterial Community of Ayiramthengu Mangrove Ecosystem of Kerala Coast, India

Mintu Ann Varghese, Anit M Thomas<sup>1</sup> and R. Sunil Kumar

Post Graduate and Research Department of Zoology, Catholicate College (Affiliated to Mahatma Gandhi University, Kottayam, India), Pathanamthitta-689 645, India. <sup>1</sup>Department of Zoology, Baselius College (Affiliated to Mahatma Gandhi University, Kottayam-686 001, India. *E-mail: mintuvar@gmail.com* 

**Abstract:** The present study focusses on the spatiotemporal variation of litter bacterial composition isolated from Ayiramthengu mangrove ecosystem of Kerala coast. Monthly sampling was conducted from three different sampling stations of mangrove ecosystem for various physicochemical parameters and bacterial analysis. Total heterotrophic bacteria study revealed that there was significant difference in bacteria among different seasons as well as different stations. 18 genera of gram-negative bacteria and 4 genera of gram-positive bacteria were isolated. *Acinetobacter* sp. was the dominant bacteria. This indicate that, Ayiramthengu mangrove contains an abundant number of bacteria, which were directly influenced by the different physicochemical parameters in the ecosystem.

Keywords: Seasons, Spatiotemporal variation, Total Heterotrophic bacteria, Mangrove, Litter

Mangroves are unique ecosystem known for its beneficial impact on both ecological and economical aspect. They provide nutrients to the coastal water, preserve and stabilize coastal zones. Microbial community is considered as the most important population in this ecosystem. Multitude of microbial communities inhabit the mangrove ecosystem as a result of the constant supply of carbon and other nutrients, and these communities are able to adapt to the environment's fluctuations and mild salinity (Thatoi et al 2013). The accumulation of litter in the mangrove is regulated by abscission, withering, death, or other factors such as wind or birds . Mangrove forest litter plays a significant role in its ecosystem as a source of energy and nutrients for numerous decomposing organisms, leading to a significant amount of organic matter and nutrient storage or export to both marine and terrestrial environments through tidal water exchange and freshwater inputs (Feller et al, 2009). The decomposition of litter is influenced by various factors like climatic conditions (Imgraben and Dittmann 2008, Kristensen et al 2008, Arnaud et al 2020), soil properties, organic matter quality (Chomel et al 2016), and the characteristics of microbial communities (Hattenschwiler et al 2005, Gießelmann et al 2010). The leaves and wood of mangrove were mainly composed of lignocellulose, that are degradable by microorganisms. Major nutrient transformation within these bacteria sustains mangroves' ecological balance and nutritional well-being (Holguin et al 2006). The present study is to examine the spatiotemporal variation of litter bacteria in relation to various physicochemical factors of Ayiramthengu mangrove ecosystem.

#### MATERIAL AND METHODS

Ayiramthengu mangrove ecosystem (lat.  $9^{\circ}07' 30'' - 9^{\circ} 07'$ 40'' N and long. 76° 28' 40''-76° 28' 50''E) was selected as the study area and was divided into three sampling stations as Station 1, Station 2 and Station 3. Station 1 near to the land area and influenced by humans, Station 2 within the middle of the mangrove with thick dense mangrove trees and Station 3 situated near to the estuary with narrow strip of vegetation. Monthly sampling was carried out for a period of one year (February 2018 to January 2019) and represented seasonally as pre monsoon (February- May), monsoon (June-September) and post monsoon (October-January).

Litter samples were taken from decaying leaf litter surface aseptically after the removal of surface contaminants. They were transferred to sterile labelled containers with 25% glycerol and transported to laboratory. Samples were serially diluted and plated on ZoBell Marine Agar (Hi Media, India) using standard plate count method and Total Heterotrophic Bacteria (THB) were determined. The individual bacterial colonies were isolated and purified. They were identified up to generic level based on cell morphology and various standard biochemical reactions as per Bergey's Manual of Determinative Bacteriology (Holt et al 2000). Several water quality and sediment parameters were recorded. The determination of various physicochemical parameters of water like temperature, pH, salinity, dissolved oxygen, nitrate, phosphate and sulphate was done according to the standard methods (APHA, 1998). Sediment characteristics like temperature, pH, total organic carbon (TOC) content (Walkley and Black method, 1934), total organic matter and nutrients like nitrate, phosphate and sulphate (Grasshoff et al 1983) were estimated.

Statistical analysis: Statistical analysis was carried out for seasonal analysis of total heterotrophic bacteria and post hoc test was done. Based on generic composition of bacteria, the similarity among the seasons were analysed by hierarchical agglomerative cluster analysis and non-metricmultidimensional scaling (MDS) based on Bray-Curtis similarities and the results obtained were plotted as ordination graphs.

#### **RESULTS AND DISCUSSION**

All the studied parameters showed variations with change in season. High temperature, pH, conductivity and salinity was during pre-monsoon season, whereas high dissolved oxygen and nutrients found during monsoon season (Table 1). During pre-monsoon season, more sunlight could increase the temperature in mangrove as well as less freshwater influx could increase the conductivity. In spite of heavy rainfall during monsoon season coupled with high wind velocity resulted in freshwater mixing in mangrove ecosystem which could be the reason for high dissolved oxygen concentration. The fluctuation in nutrient content was controlled by natural sources like igneous rock, plant decay and animal debris as well as man-made sources like fertilizers, livestock, urban and agricultural runoff and wastewater discharges (Lee et al 2002, Jalali 2005, Lotfinasabasl et al 2013). High organic content could be due to the accumulated dead and decaying planktonic organisms and large amounts of humus transported by the rivers and terrestrial land drainages.

Seasonal analysis of total heterotrophic bacteria showed significant difference among various seasons observed (Fig. 1). The post hoc analysis suggested that pre monsoon and monsoon season, and both post monsoon and monsoon season showed significant difference in heterotrophic bacterial population whereas pre monsoon and post monsoon showed no significant difference. Station-wise litter heterotrophic bacterial density indicated significant difference among stations and on post hoc analysis Station 1 and Station 2 and Station 2 and Station 3 showed significant difference whereas Station 1 and Station 3 show no significant difference.

The heterotrophic bacteria were correlated with the water and soil parameters of the region (Table 2). The water parameters like dissolved oxygen, nitrate, phosphate and sulphate when corelated with the heterotrophic bacteria of litter showed positive results. The correlation of heterotrophic bacteria of litter with the soil parameters such as organic carbon, organic matter, nitrate, phosphate and sulphate was positive when compared to the other parameters. Cotano and Villate (2006) also observed that the increase in bacterial population was contributed by the release of effluents.

The litter bacterial composition appeared to be relatively heterogeneous with various stations. This was due to the

Table 1. Ph	vsicochemical	parameters of Av	viramthenau	u manarove	ecosvstem	(Mean ± SD	)
						•	

Parameters	Pre monsoon	Monsoon	Post monsoon
Water quality parameters			
Temperature (°C)	30.3 ± 0.57	27.5 ± 0.49	28.8 ± 0.87
рН	7.53 ± 0.10	$7.47 \pm 0.06$	7.51 ± 0.06
Conductivity (m/s)	23.92 ± 2.75	9.42 ± 1.24	22 ± 2.21
Dissolved Oxygen (mg/l)	$2.97 \pm 0.69$	$3.3 \pm 0.74$	$2.06 \pm 0.69$
Salinity	8.1 ± 2.48	$3.43 \pm 0.69$	4.65 ± 1.55
Nitrate (mg/l)	1.13 ± 0.2	2.47 ± 0.77	2.18 ± 0.8
Phosphate (mg/l)	1.38 ± 0.14	$2.28 \pm 0.26$	1.32 ± 0.31
Sulphate (mg/l)	75.08 ± 2.64	138.27 ± 17.35	115.1 ± 8.73
Sediment characteristics			
Temperature (°C)	$26.8 \pm 0.42$	$23.7 \pm 0.42$	25.1 ± 0.51
рН	6.83 ± 0.13	6.37 ± 0.15	6.38 ± 0.11
OC (%)	$0.40 \pm 0.09$	0.41 ± 0.09	0.61 ± 0.09
OM (%)	$0.65 \pm 0.15$	$0.75 \pm 0.32$	1.05 ± 0.16
Nitrate (mg/g)	$2.64 \pm 0.0.06$	$2.58 \pm 0.03$	$2.24 \pm 0.06$
Phosphate (mg/g)	$3.04 \pm 0.05$	$3.20 \pm 0.06$	$3.08 \pm 0.08$
Sulphate (mg/g)	82.02 ± 1.35	85.78 ± 1.42	81.50 ± 1.56



Fig. 1. Spatio-temporal variation in litter heterotrophic bacteria



Fig. 2. Spatiotemporal variation in the generic distribution of litter bacteria (%)



Fig. 3. Cluster diagram plotted based on similarity/dissimilarity of the various seasons in relation to the litter bacterial diversity

 
 Table 2. Correlation analysis of litter bacteria with water and soil parameters

Paramet	er Correlation	Parameters ® value)	Correlation coefficient (r)
Water	Positive	Dissolved	0.373
	correlation	Oxygen	0.466
		Nitrate	0.727
		Phosphate	0.558
		Sulphate	
	Negative	Temperature	-0.379
	correlation	рН	-0.167
		Conductivity	-0.619
		Salinity	-0.480
Soil	Positive	Organic carbon	0.018
	correlation	Organic matter	0.019
		Nitrate	0.172
		Phosphate	0.383
		Sulphate	0.264
	Negative	Temperature	-0.58
	correlation	рН	-0.462

difference in the rate of decomposition found in different stations. Besides that, all the stations had equal importance in the detritus decomposition and were thereby actively involved in biogeochemical cycles. But the litter heterotrophic bacteria indicated changes among different seasons. However, the physicochemical variations that occurred during different seasons affected the litter production rate (Juman 2005, Zafar et al 2012). It is well known that mangrove microbial communities were strongly influenced by the environmental parameters like temperature, pH, salinity, dissolved oxygen and nutrient concentrations (Tam 1998, Li and Gu 2013). During the study, heterotrophic bacterial load from litter revealed that gram negative bacteria were relatively abundant with 18 genera (82%), while gram positive bacteria contributed only 4 genera (18%). The comparatively higher percentage of gram-negative bacteria showed higher decomposition favoured by the litter substratum for the better adapted gram-negative bacteria.

Acinetobacter sp. (13%) was the dominant followed by Micrococcus sp., Bacillus sp., Pseudomonas sp. (Fig. 2). The cluster analysis suggested that the monsoon and post monsoon season (88%) showed more similarity than to pre monsoon season (Fig. 3). Acinetobacter involved in degradation pathways of various long-chain dicarboxylic acids and aromatic and hydroxylated aromatic compounds. Dorothy et al (2003) stated that Micrococcus were actively involved in leaf litter decomposition in mangrove forest. Bacillus sp. was known for its cadmium biodegradation, hydrocarbon degradation, emulsification, phytase production, plant growth promotion and protection of plant from microbes (Holguin et al 2001, Macrae et al 2001). Pseudomonas had a major role in the turnover of certain nutrients like nitrogen, carbon and phosphorous appeared in the leaf biomass (Mumby et al 2004, Romero et al 2005).

#### CONCLUSION

The heterotrophic bacterial population was influenced by various physico-chemical parameters of soil and water. Moreover, Gram negative bacteria was comparatively high than Gram positive bacteria with a dominance of *Acinetobacter* sp. Thus, it is concluded that Ayiramthengu mangrove ecosystem have a rich litter bacterial population holding a well-organized community structure at various seasons as well as stations.

#### REFERENCES

- Alongi DM 1994. The role of bacteria in nutrient recycling in tropical mangrove and other coastal benthic ecosystems. *Hydrobiologia* **285**(1-3): 19-32.
- APHA 1998. Standard Methods for the examination of water and wastewater. 20<sup>th</sup> Edition, American Public Health Association, American Water Works Association and Water Environmental Federation, Washington DC.
- Arnaud M, Baird AJ, Morris PJ, Dang TH and Nguyen TT 2020. Sensitivity of mangrove soil organic matter decay to warming and sea level change. *Global Change Biology* 26:1899-1907.
- Cotano U and Villate F 2006. Anthropogenic influence on the organic fraction of sediments in two contrasting estuaries: A biochemical approach. *Marine Pollution Bulletin* **52**(4): 404-414.
- Chomel M, Guittonny-Larchevêque M, Fernandez C, Gallet C, DesRochers A, Pare D, Jackson BG and Baldy V 2016. Plant secondary metabolites: A key driver of litter decomposition and soil nutrient cycling. *Journal of Ecology* **104**:1527-1541.
- Dorothy KP, Satyanarayana B, Kalavati C and Raman AV 2003. Protozoa associated with leaf litter degradation in Coringa mangrove forest, Kakinada Bay, east coast of India. *Indian Journal of Marine Sciences* **32**(1):45-51.
- Feller IC, Lovelock CE, Berger U, McKee KL, Joye SB and Ball MC 2009. Biocomplexity in mangrove ecosystems. *Annual Review of Marine Science* **2**: 395-417.
- Gießelmann UC, Martins KG, Brändle M, Schädler M, Marques R and Brandl R 2011. Lack of home-field advantage in the decomposition of leaf litter in the Atlantic Rainforest of Brazil. *Applied Soil Ecol*ogy **49**: 5-10.
- Grasshoff P 1983. *Methods of seawater analysis*. Verlag Chemie. *FRG*, *419*.
- Hättenschwiler S, Tiunov AV and Scheu S 2005. Biodiversity and litter decomposition in terrestrial ecosystems. *Annu Review of Ecology, Evolution, and Systematics* **36**: 191-218.
- Holguin G, Vazquez P and Bashan Y 2001. The role of sediment microorganisms in the productivity, conservation, and rehabilitation of mangrove ecosystems: An overview. *Biology* and Fertility of Soils 33(4): 265-278.
- Holguin G, Gonzalez-Zamorano P, de-Bashan LE, Mendoza R, Amador E and Bashan Y. 2006. Mangrove health in an arid environment encroached by urban development: A case study. *Science of the Total Environment* **363**(1-3): 260-274.
- Holt GH, Kreig NH, Sneath PHA, Staley JT and Williams 2000. Bergey's Manual of determinative bacteriology. Williams & Wilkins.
- Imgraben S and Dittmann S 2008. Leaf litter dynamics and litter consumption in two temperate South Australian mangrove forests. *Journal of Sea Research* **59**: 83-93.
- Jalali M 2005. Nitrates leaching from agricultural land in Hamadan, western Iran. Agriculture, Ecosystems and Environment **110**(3-4): 210-218.

Received 22 June, 2024; Accepted 11 September, 2024

- Juman RA 2005. Biomass, litter fall and decomposition rates for the fringed *Rhizophora mangle* forest lining the Bon Accord Lagoon, Tobago. *International Journal of Tropical Biology and Conservation* **53**: 207-217.
- Kristensen E, Bouillon S, Dittmar T and Marchand C 2008. Organic carbon dynamics in mangrove ecosystems: A review. Aquatic Botany 89: 201-219.
- Lotfinasabasl S, Gunale VR and Rajurkar NS 2013. Water quality assessment of Alibaug mangrove forest using multivariate statistical technique, Maharastra, India. *Indian Journal of Geo-Marine Sciences* **42**(7): 915-923.
- Lee SM, Min KD, Woo NC, Kim YJ and Ahn CH 2002. Statistical assessment of nitrate contamination in urban ground water using GIS. *Environmental Geology* **44**: 210-221.
- Li M and Gu JD 2013. Community structure and transcript responses of anammox bacteria, AOA, and AOB in mangrove sediment microcosms amended with ammonium and nitrite. *Applied Microbiology and Biotechnology* **97**(22): 9859-9874.
- Macrae A, Lucon CMM, Rimmer DL and O'Donnell AGO 2001. Sampling DNA from the rhizosphere of *Brassica napus* to investigate rhizobacterial community structure. *Plant and Soil* 233(2): 223-230.
- Mumby PJ, Edwards AJ, Arias-González JE, Lindeman KC, Blackwell PG, Gall A, Gorczynska MI, Harborne AR, Pescod CL, Renken H, Wabnitz CC and Llewellyn G 2004. Mangroves enhance the biomass of coral reef fish communities in the Caribbean. *Nature* 427(6974): 533-536.
- Nagarajaiah CS and Gupta TRC 1983. Physico-chemical characteristics of brackish water ponds along Nethravathi estuary, Mangalore. *Indian Journal of Marine Sciences* **12**: 81-84.
- Romero LM, Smith TJ and Fourqurean JW 2005. Changes in mass nutrient content of wood during decomposition in a South Florida Mangrove Forest. *Journal of Ecology* **93**(3): 618-631.
- Strickland JDH and Parsons TR 1972. A practical handbook of seawater analysis. Fisheries Research Board of Canada, Ottawa.
- Tam NFY 1998. Effects of wastewater discharge on microbial populations and enzyme activities in mangrove soils. *Environmental Pollution* **102**(2-3): 233-242.
- Thatoi H, Behera BC, Mishra RR and Dutta SK 2013. Biodiversity and biotechnological potential of microorganisms from mangrove ecosystems: A review. *Annals of Microbiology* **63**(1): 1-19.
- Tomlinson PB 1986. *Botany of mangroves*. Cambridge University Press.
- Zafar F, Seema S, Khawer LK, Amjad A, Pervaiz I and Pirzada JAS 2012. Assessment of litter production in semi-arid mangroves forests near Active Indus River Mouth (Hajambro Creek) and Karachi Backwaters, Pakistan. *Pakistan Journal of Botany* 44(5): 1763-1768.



## Abundance and Diversity of Mollusks and Related to Some Physicochemical Properties in Euphrates River in Al-Muthanna Province

Afrah Kadhim Helal, Ali Abdulhamza Al-Fanharawi<sup>\*</sup> and Ibtehal Aqeel Al-Taee

College of Science, Al-Muthanna University, Iraq E-mail: alialfanharawi@mu.edu.iq

**Abstract:** Study was conducted to identify the composition of mollusks in relation to some physical-chemical parameters of the Euphrates river during the period from November 2019 to October 2020. The water temperature ranged between 11.33-38.67 °C. The highest mean of electrical conductivity was 3026.6 $\mu$ S/cm, whereas the lowest was 1203.3  $\mu$ S/cm. The maximum TDS was 2766.6 mg/l . The water flow was ranged from 0.133 to 0.426 m/sec. The turbidity ranged between 11.59 and 134.63NTU. TSS recorded the highest mean of 36.66 mg/l and lowest of 21 mg/l. The pH values ranged between 6.63-8.60 and the concentrations of DO and BOD5 between 5.15-10.10 and 1-12.33 mg/l, respectively. TA, TH, Ca+2, and Mg<sup>+2</sup> were within 131.33-181.66, 423.33-1386.6, 125-320 mg. The NO2<sup>-2</sup>, NO3<sup>-2</sup> and PO4<sup>-2</sup> varied between (0.03-3.05, 0.053-6.890 and 0.06-0.77  $\mu$ g l<sup>-1</sup>, respectively). The TOC ranged between 0.055-3.12 % and type of texture was silty in all stations. The study revealed the occurrence of 6 species of Mollusks with total density of 242.84 Ind. /m<sup>2</sup>. The relative abundance index, showed that the species *M. nodosa* was dominant and *C. fluminalis* less abundant. The Shannon-Weiner and Simson diversity indices ranged between 0.39-2.95 and 0.1-0.93, respectively.

Keywords: Physicochemical properties, Biodiversity, Euphrates river, Mollusks, Samawa.

Freshwater constitutes 3% of the total water on earth and the rivers as a freshwater source are complex systems containing rich diversity of living species which are in close interdependence with their surrounding physical environment (Brachet 2015). Among the organisms in the rivers, mollusks are the major grazers and playing a pivotal role in aquatic food webs, significantly influence algal primary productivity, nutrient cycling, water purification, and provide valuable information by their presence, absence, and abundance regarding their surrounding habitat and can be used to assess the local environmental impact. Freshwater mollusks are perfect organisms for rapid biological survey due to their sensitivity to changes in the environment and water quality and capacity for water filtering (Cummings 2016). The deterioration of physical and chemical water quality is often caused by human influences gradually, and the invisible adaptations of aquatic ecosystems to these changes may not always be discovered until a major transformation in the state of the ecosystem occurs (Stark et al 2000). These systems should be continuously monitored by assessing the species of organisms or measuring some physical and chemical properties. The current study was designed to study monthly variation in physicochemical parameters, which affect the abundance and diversity of Mollusk in the study area.

#### MATERIAL AND METHODS

Study area: The study area included three sampling sites located on a Euphrates river at Al-Muthanna province. Al-Muthanna province is located about 270 km south of Baghdad, the capital of Iraq. The first sitelocated in Al-Hilal area(31º42'40.34"N- 45º45'11.16"E), while the second siteabout 34 km south of the first site(31°31'49.46"N-45º.29'72.79"E), in the city center of Samawa near Al-Shuhada Bridge, and the third sitelocated about 16 km south of the second site (31º.29'31.17"N - 45º.45'63.35"E) (Fig. 1). Sampling procedures: The study was conducted monthly from November 2019 to October 2020. Water samples were taken for physical and chemical parameter analysis. Water temperature (°C) was measured and water flow (m /sec) directly using the float method mentioned (US EPA (1997). Electrical conductivity (µS/cm) and pH were estimated by using a portable Multimeter (SM801). Alkalinity was measured by the titration method (Estefan et al 2013) and reactive phosphate, nitrate, and nitrite were estimated according to Nollet (2014). Total dissolved solids were measured by an evaporation method, total dissolved solids were measured by gravimetric method (APHA (2017). The total hardness, Ca hardness, Mg and dissolved oxygen were measured by using a titrimetric method (APHA 2017). Biological oxygen demand (BOD<sub>5</sub>) measured by sensor method (Al-Fanharawi et al 2019). The turbidity was measured in the field by turbidity meter, type Hanna (WTW-Turb-550). Sediment samples were collected for analysis the total organic carbon (Estefan et al 2013) and grain size analysis was carried out by using the standard sieving and pipet techniques (Folk 1974).

Mollusks were also sampled monthly using Vanveen Grab Sampler monthly (Al-Fanharawi 2010). Mollusks have been identified according toPlaziat and Younis, (2005) and Damborenea et al (2020). Mollusks were grouped by species and density determined (Ottensmann 2018) and expressed as individual/m<sup>2</sup>. Relative abundance was calculated (Baderan et al 2019). Shannon-Wiener diversity (H') counted according to Baderan et al (2019) and Simpson's diversity as mentioned by Vajravelu et al (2018).

#### **RESULTS AND DISCUSSION**

**Physical parameters:** The highest water temperature was 38.67 °C at St.3 in July and the lowest 1.33°C at St.1 in February (Table 1). The monthly fluctuation in the water temperature was due to a long day period in the summer and the short day period in winter. The spatial variations in water temperatures may be due to the difference in the time of collection of the sample at each station. The highest value of electrical conductivity (EC) was 3026.6  $\mu$ S/cm recorded in July at St.3, while lowest mean value 1203.3 ( $\mu$ S/cm) was in November at St.1 (Table 1). The highest values of electrical conductivity may be due to higher temperatures, which lead

to an increase in the solubility of the salts in addition to an increase in evaporation and an increase in the concentration of salts. Najafpoor et al (2007) indicated that highest electrical conductivity depends on the amount of total dissolved solids in water, which have the highest value of 2766.6 mg/l during July in St.2, while the lowest value 833.3 mg/l during November in St.1. The high concentrations of TDS may be due to higher temperature leads to high rates of evaporation that an increase of salt concentrations (Al-Fanharawi 2010). The highest mean water flow was 0.426 m/sec recorded in June at St.2, whereas the lowest value



Fig. 1. Sampling site on Euphrates River

#### Table 1. Physical and chemical parameters in Euphrates river (Mean ± standard deviation)

Stations/Parameters	St.1	St.2	St.3
Water temperature (C°)	25.06±8.88A	23.44±8.74A	21.92±8.53A
Electrical conductivity (µS/cm)	2032.7±548.7A	1988.8±492.8A	1810.5±486.2B
Total dissolved solids (mg/l)	1794±512.4A	1753.0±517.3A	1541.9±512.5B
water flow (m/sec)	0.236±0.065C	0.340±0.084A	0.266±0.073B
Turbidity (NTU)	40.17±29.33A	42.40±31.43A	29.74±19.27B
Total suspended solids (mg/l)	83.80±26.13A	8.16±25.63B	43.50±19.14C
pH	7.73±0.05A	7.76±0.05A	7.66±0.15A
dissolved oxygen (mg/l)	6.70±0.90B	7.23±0.99B	7.85±1.24A
biochemical oxygen demand (mg/l)	7.11±2.98A	6.19±2.93B	4.66±2.48C
Total alkalinity (mgCaCO₃/L)	153.63±12.57	153.00±14.72	154.47±16.59
Total hardness (mgCaCO₃/L)	869.44±241.1A	815.83±208.4AB	761.94±175.2B
Calcium hardness (mgCaCO <sub>3</sub> /L)	185.58±50.99	175.02±37.04	162.13±30.95
Magnesium (mg/l)	165.77±46.91A	156.85±42.77AB	145.76±35.76B
Nitrite (µg/I)	1.136±0.889A	1.121±0.883A	0.993±0.7A
Nitrate (µg/l)	4.152±2.226A	3.481±0.184B	3.017±1.816C
Reactive phosphate (µg/l)	0.340±0.211A	0.295±0.178B	0.238±0.142C

Different capital letters in row indicate significant difference in parameters between stations

0.133 m/sec recorded at St.1 (Table 1). The highest mean water flow in St.2 may be due to narrowing of the river at this station, and that the velocity of the river can change at various stations along the course of a river due to the shape, the lowest mean value in St.3 attributed to the presence of aquatic plants that delayed water flow. The highest turbidity and total suspended solids values were in October were 134.63 NTU and 136.66 mg/l, respectively .This may be due to the increased discharge of water into the Euphrates River, which flows with a high suspended matter content (Rabee et al 2011) and the lowest values were recorded in November at St.1 was 11.59 NTU and 21 mg/l respectively and may be due to the presence of aquatic plants (Noaman 2008).

Chemical parameters: The pH of water in the current ecosystem can be classified as slightly alkaline, the higher pH was in February at St.2 (6.63-8.6). This may be due to reduced microbial activity, improved algal productivity and the decreased decomposition (Sisodia and Chaturbhuj 2006). The low pH in July may be due to an increased rate of decomposition, leading to acidification (Ahipathy and Puttaiah 2006). The minimum of dissolved oxygen (5.15 mg/l) and higher levels of biological oxygen demand (12.33 mg/l) was in July (Table 1). The high BOD<sub>5</sub> may be due to the high decomposition processes of microbial organisms due to the height of the temperature Arimoro et al (2006) and these processes require the consumption of high concentrations of dissolved oxygen that lead to low DO. The highest alkalinity, total hardness, calcium hardness, and magnesium were in July (181.66, 1386.6, 320 mg CaCO3/L and 259.2 mg/l) respectively. The high alkalinity attributed to the relationship between total alkalinity and total hardness, through common ions formed in aquatic systems, carbonate, and bicarbonate are the principal cations responsible for hardness are ions associated with alkalinity (Burton Jr and Pitt 2002). The higher total hardness, calcium hardness, and magnesium, maybe due to low water level and high rate of evaporation and decomposition that concentrating the salts (Mossa 2006).

The highest mean nitrite  $3.05 \ \mu g/l$  was at St.2 in December, while the lowest  $0.03 \ \mu g/l$  was at St.3 in July Low levels of nitrite concentration may be related to a complex of biological processes, including the oxidation of nitrite as an unstable form of nitrogen to nitrate by nitrifying bacteria (Abdo and El-Nasharty, 2010). The highest mean nitrate was 6.890  $\mu g/l$  in July and this may be due to deep ground waters with high nitrate concentrations which is important source of nitrate in surface waters (Tesoriero et al 2013). The lowest mean was 0.053  $\mu g/l$  in Januarydue to the high water level of the river and the dilution factor that effect on concentrations (Al-Fanharawi 2010). The highest reactive phosphate was

0.77  $\mu$ g/l in July at St.3 due to the increasing water temperature, which causing increasing the degradation of organic matter containing phosphate compounds, including algal cells, or may be due to phosphate fertilizers used to fertilize agricultural land and discharged to the river and this agree with Mokaya et al (2004), while lowest mean of reactive phosphate was at St.1 (0.060  $\mu$ g/l) and may be due to location of station more distant from the effluent discharge point.

**Sediment samples:** Sediment analysis in the present study showed that the lowest value of TOC % in sediments (0.055 %) was in June, while the highest (3.12 %) was at St. 3 in October (Fig. 2). The highest value may be due to contamination of this station with organic pollutants due to the direct discharge of domestic wastewater to the river (Zhibo et al 2009). Grain size analysis of sediment showed that the sediment texture of the three indicates silt, clay and sand as 90, 5 and 5%. According to the soil textural triangle, the sediments of the Euphrates River on sites can be considered as silty.

Identification and population mollusks: In the current study, from 577 individuals of Mollusks, identified 6 taxa belonging to 5 families, 3 orders, and 2 classes (Gastropoda and Bivalvia) in three sites on Euphrates river. Bivalvia class (3 species) comprised the orders Unionida (*U. tigridis*), and Veneroida (*C. fluminea, and C. fluminalis*), whereas the class Gastropoda (3 species) comprised one order Mesogastropoda were *M. tuberculata, M. nodosa, and V. bengalensis.* 

**Density of Mollusks:** The highest mean of mollusks density was in St.2 was 390.4 Ind./m<sup>2</sup> and this may be due to the



Fig. 2. Monthly variation in total organic carbon values in Euphrates river during study periods

effect of Al- Shuhada Bridge. Blettler and Marchese (2005) also showed the effect of bridges on the benthic invertebrates. The mollusks and found that the species density was highest under the bridges than upstream and downstream, while the lowest mean recorded in St.1was 127.52 Ind./m<sup>2</sup>(Table 2).

**Ecological indices:** The relative abundance of species varied between study stations, St.2 was the most abundant among while St.1 was less abundant (Fig 3-A). The species *M. nodosa* was the most abundant species (75.79%), followed by *C. fluminea* (8.07%), *V. bengalensis* (6.81%), and *M. tuberculata* (6.37%), *U. tigridis* (2.59%), whereas *C. fluminalis* (0.37%) was the less abundant species. Shannon-Wiener diversity index indicate lowest monthly diversity of

 
 Table 2. Total density of mollusk species in Euphrates river (Ind./m<sup>2</sup>)

Stations/ Months	St.1	St.2	St.3
Nov. 2019	30.3	75.75	75.75
Dec.	30.3	45.45	60.6
Jan.2020	45.45	30.3	45.45
Feb.	75.75	15.15	121.21
Mar.	90.9	60.6	75.75
Apr.	90.9	136.36	121.21
Мау	151.5	121.21	121.21
Jun.	90.9	212.12	136.36
Jul.	90.9	1015.15	454.54
Aug.	106.06	1045.45	45.45
Sep.	345.45	1163.63	651.51
Oct.	381.81	763.63	618.18
Means	127.52	390.4	210.6

mollusks was 0.39 in August (Fig. 3A). This may be due to a combination of abiotic factors, including, low water flows and temperature increase, while the highest value of 2.95 was in October, may be due to the rapid migration of these organisms. Bishop et al (2009) reported that the rapid migration of mollusks reflects the behavioral response from them or may be due to the effect of temperature, the presence of vegetation cover nutrition as key factors for the success and the development of certain species. Badsi et al (2010) observed that highest organic carbon in sediment was in October. Sharmin et al (2019) reported that the lowest species diversity associated with low sediment organic carbon and low sediment organic matter. The highest value of Simpson index was 0.93 in October and may be due to the higher TOC recorded in this month, or may be due to the appropriate temperature and present the plant cover, while the lowest value was 0.1 in April (Fig. 3A). Two indices (Shannon-Wiener diversity and Simpson diversity) agreed on recording the same highest and lowest in the same stations St.1 and St.2 respectively (Fig. 3B). The lowest diversity in St.2 may be due to lack of evenness of species and decreases in the species richness, this due to the dominance of a few species with high densities, while that the opposite of what was on he St.1 was have the highest richness of species. Kimbro (2006) reported that diversity depended on two major components, richness, and evenness of species.

#### CONCLUSION

1. The highest mean of most physicochemical parameters (electrical conductivity, total suspended solids, biological oxygen demand, total alkalinity, total hardness, calcium hardness, nutrients, and total organic carbon) was recorded at outside city center St.3.



Fig. 3. Simpson and Shannon-Weiner diversity indices values of Mollusks species in stations in Euphrates river A- Monthly variation; B: Variations between stations

- The values of density, relative abundance, and diversity (Shannon-Weiner and Simpson diversity indices) of mollusks, were impacted by total organic carbon, water flow, and temperatures.
- The current study showed the species *M. nodosa* was the most abundant species in all stations during the study periods, while the species *C. fluminalis* was the less abundant species.
- 4. When more indices are used to assess the diversity, which gives a more accurate and clear picture, the two diversity indices (Shannon-Weiner and Simpson indices) agreed that the highest and lowest values were recorded in the same stations.
- Biological diversity index showed that Euphrates river in the study area was low diversity, according to the Shannon-Weiner diversity index <1.</li>

#### REFERENCES

- Abdo MH and El-Nasharty SM 2010. Physico-chemical evaluations and trace metals distribution in water-surficial sediment of Ismailia canal. Egypt. *Nature and Science* **8**(5): 198-206.
- Ahipathi MV and Puttaiah ET 2006. Ecological characteristic of Vrishabhavathy River in Bangalor (India). *Environmental Geology* 49(8): 1217-1222.
- Al-Fanharawi A 2010. *Distribution and diversity of the benthic macroinvertebrates in sediments of Shatt Al-Hilla/Iraq*. Master Thesis. Science College, Babylon University, Iraq.
- Al-Fanharawi AA, Rabee AM and Al-Mamoori AM 2019. Multibiomarker responses after exposure to organophosphates chlorpyrifos in the freshwater mussels Unio tigridis and snails Viviparous benglensis. Human and Ecological Risk Assessment: An International Journal 25(5): 1137-1156.
- APHA (American public Health Association) 2017. Standard methods for the examination of water and waste water. 23rd', Washington DC, USA.
- Arimoro FO, Ikomi RB and Osalor EC 2006.. The impact of Sawmill wood wastes on the water quality and fish communities of Benin River, Niger Delta area. *Nigeria World Journal* 1(2): 94-102.
- Baderan DWK, Hamidun MS, Utina R, Rahim S and Dali R 2019. The abundance and diversity of Mollusks in mangrove ecosystem at coastal area of North Sulawesi, Indonesia. *Biodiversitas Journal* of *Biological Diversity* 20(4): 987-993.
- Badsi H, Ali HO, Loudiki M, ElHafa M, Chakli R and Aamiri A2010. Ecological factors affecting distribution of zooplankton community in Massa Lagoon (southern Morocco). African Journal of Environmental Science and Technology 4(11): 751-762.
- Bishop MJ, Morgan T, Coleman MA, Kelaher BP, Hardstaff LK and Evenden RW 2009. Facilitation of molluscan assemblages in mangroves by fucalean alga *Hormosira banksii*. *Marine Ecology Progress Series* **392**: 111-122.
- Blettler MCM and Marchese Garello MR 2005. Effects of bridges construction on the benthic invertebrates structure of the Delta Paraná River. *Interciencia* **30**(2): 60-66.
- Brachet C 2015. Handbook for Management and Restoration of Aquatic Ecosystems in river and lake basins. Anne-Marie Harper. ISBN: 978-91-87823-15-2.
- Burton Jr GA and Pitt R 2002. Stormwater Effects Handbook: A Toolbox for Watershed. CRC Press LLC, Washington, D.C, 0-87371-924-7.
- Cummings KS, Jones HA and Lopes-Lima M 2016. Rapid

bioassessment methods for freshwater molluscs. Core Standardized Methods: p. 186.

- Damborenea C, Rogers DC and Thorp JH (Eds.) 2020. *Thorp and Covich's Freshwater Invertebrates:* Volume 5: Keys to Neotropical and Antarctic Fauna. Academic Press.
- Estefan G, Sommer R and Ryan J 2013. Methods of soil, plant, and water analysis. *A manual for the West Asia and North Africa Region* **3**: 65-119.
- Mokaya SK, Mathook JM and leichtfried M 2004. Influence of anthropogenic activities on water quality of a tropical stream ecosystem. *African Journal of Ecology* **42**(4): 281-288.
- Matsukura K, Tsumuki H, Izumi Y and Wada T 2009. Physiological response to low temperature in freshwater apple snail, *Pomacea canaliculata* (Gastropoda: Ampullariidae). *Journal of Experimental Biology* **212**(16): 2558-2563.
- Mohammad MK 2014. Ecology of freshwater snail *Melanopsis* buccinoidea (Olivier 1801) in Ain Al-Tamur, Kerbala Province. International Journal of Current Microbiology and Applied Sciences **3**(2): 390-394.
- Mossa ZJ 2006. Study the Environmental Changes of the Phenols in the Shatt Al-Arab and Its Branches and Its Impact on the Internal Density of Algae. Master's Thesis, University of Basra, Basrah, Iraq.
- Najafpoor AA, Vojoudi Z, Dehgani MH, Changani F and Alidadi H 2007. Quality assessment of the Kashaf River in north east of Iran in 1996-2005. *Journal of Applied Sciences* 7(2): 253-257.
- Noaman MM 2008. Effect of Industrial influent on water quality of Tigris river and upon the performance treatment plant within sector Baiji-Tikrit. M.Sc. thesis, College. of Engn., Tikrit University. 199p.
- Nollet LM and De Gelder PS 2014. Hand Book of Water analysis. 3th ed. CPC Press, Taylor and Francis Group, LLC.
- Ottensmann JR 2018. On population-weighted density. Available at SSRN 3119965.
- Plaziat JC and Younis WR 2005. *The modern environments of Molluscs in southern Mesopotamia*, Iraq: A guide to palaeogeographical reconstructions of Quaternary fluvial, palustrine and marine deposits. CG. Notebooks on Geology-A01:18.
- Rabee AM, Abdul-Kareem BM and Al-Dhamin AS 2011. Seasonal variations of some ecological parameters in Tigris River water at Baghdad Region, Iraq. *Journal of Water Resource and Protection* **3**(4): 262.
- Sharmin S, Rahman SH and Naser MN 2019. Distribution and diversity of molluscs at migratory bird visiting and non-visiting lakes of Jahngirnagar University, Savar. *Bangladesh Journal of Zoology* 47(2): 355-366.
- Sisodia R and Chaturbhuj M 2006. Assessment of the water quality index of wetland Kalakho Lake, Rajasthan, India. *Journal Environmental* Hydrology **14**(23): 1-11.
- Stark J, Hanson P, Goldstein R, Fallon J, Fong A, Lee K, Kroening S and Andrews W 2000. Water Quality in the Upper Mississippi River Basin, Minnesota, Wisconsin, South Dakota, Iowa, and North Dakota, 1995-98. United States Geological Survey, Circular 1211, Reston, Virginia, 36 pp.
- Tesoriero AJ, Duff JH, Saad DA, Spahr NE and Wolock DM 2013. Vulnerability of streams to legacy nitrate sources. *Environmental science and technology* **47**(8): 3623-3629.
- Vajravelu M, Martin Y, Ayyappan S and Mayakrishnan M 2018. Seasonal influence of physico-chemical parameters on phytoplankton diversity, community structure and abundance at Parangipettai coastal waters, Bay of Bengal, South East Coast of India. *Oceanologia* **60**(2): 114-127.
- Zhibo LU, Lian Z, Ning L and Yue W 2009. Analysis of pollution status of water quality in Bailianjing River and countermeasures. *International Conference on Energy and Environment Technology*.pp: 423-426.

Received 01 June, 2024; Accepted 21 September, 2024

# CONTENTS

4350	Diversity and Phosphate Solubilization Potential of Rhizospheric Fungi from different Land-use of Mokokchung district, Nagaland, India Imlimenla Jamir, Wati Temjen and Tali Ajungla	1049
4351	Studies on Four Species of Genus <i>Hypolampurus</i> Hampson (Lepidoptera: Thyrididae) from Western Ghats, India <i>Amit Katewa and P.C. Pathania</i>	1054
4352	Potential of Twin Key Management Practices: Higher Colony Strength and Lower Honey Extraction Frequency in Improving Honey Quality of <i>Apis mellifera</i> <i>Sumit Saini, O.P. Chaudhary, Vadde Anoosha and Lalita</i>	1060
4353	Morphological and Molecular Confirmation of Thrips palmi, Karny 1925 (Thripidae) and <i>Haplothrips tenuipennis</i> Bagnall, 1918 (Phlaeothripidae) of Order Thysanoptera in Muthalamada Mangoes of Kerala, India	1065
	Syed Mohamed Ibrahim S., Malini Nilamudeen, Pratheesh P. Gopinath and Linitha Nair	
4354	Life Table of Pod Borer, <i>Helicoverpa armigera</i> (Hubner) on Pigeonpea B.C. Patel, Bindu Panickar and M.R. Dabhi	1070
4355	Phenology, Productivity and Profitability with Phosphate and Zinc Solubilizing Microbes in Lentil ( <i>Lens culinaris</i> L.) under tarai region of Uttarakhand Monica, Yaying Anil Shukla, Supriya, Sudarshan S., Shobhana Singh and Gunashekhar H. and Chandra Bhushan	1076
4356	Morphological and Chemical Profile of <i>Pulicaria undulata</i> L. (Compositae) in Iraq <i>Khansaa R. Al-Joboury</i>	1081
4357	Impact of Western Disturbances on Stone Fruits in Mid Hill Zone of Himachal Pradesh Prakriti Dadial, M.S. Jangra, S.K. Bhardwaj, Purnima Mehta and Akanksha Klate	1085
4358	Standardization of Growing Media for Haworthia Pot Plant Alka Singh, G.D. Patel, H.P. Shah, A.J. Bhandari, R.A. Gurjar and S.T. Bhatt	1089
4359	Assessment of Roadside Ornamental Trees Potential For Mitigating Heavy Metal Pollution in Ludhiana, Punjab <i>J. Verma, P. Singh and R. Singh</i>	1093
4360	Aquaponic Production of Ornamental Koi Carp ( <i>Cyprinus carpio</i> Linn.) and Lettuce ( <i>Lactuca sativa</i> ) in Comparison with Traditional Fish Culture and Hydroponics System <i>Khushvir Singh, Vaneet Inder Kaur, Meera D. Ansal, Dilpreet Talwar, Abhishek Srivastava</i> <i>and Kulbir Singh</i>	1099
4361	Nest Site Selection and Habitat Preference of Baya Weaver <i>Ploceus philippinus</i> in Agricultural Landscape	1104
	Suknpreet Kaur Sidnu, Gurkirat Singh Seknon and Tejdeep Kaur Kier	
4362	Status and Impact of Wooded Patches in Semi-Urban Landscape on Avian Community Structure in Aligarh, Uttar Pradesh, India <i>Aditya Rana and Jamal Ahmad Khan</i>	1109
4363	Assessment of Hazardous Effects of Lead on Growth Parameters of Soybean ( <i>Glycine max</i> (L.) Merr.) Siddhi Gupta and M.K. Meena	1117
4364	Temporal Variation in Water Quality of Dhansiri River, North East India Wati lemla, Maibam Romeo Singh and Khikeya Semy	1121
4365	Premonsoon Groundwater Trends for Nanjangud Taluk, India: A Spatio-Temporal Analysis <i>M.C. Manjunatha</i>	1127
4366	Spatiotemporal Variation in Litter Bacterial Community of Ayiramthengu Mangrove Ecosystem of Kerala Coast, India <i>Mintu Ann Varghese, Anit M Thomas and R. Sunil Kumar</i>	1132
4367	Abundance and Diversity of Mollusks and Related to Some Physicochemical Properties in Euphrates River in Al-Muthanna Province Afrah Kadhim Helal, Ali Abdulhamza Al-Fanharawi and Ibtehal Aqeel Al-Taee	1137



Indian Journal of Ecology (2024) 51(5)

## CONTENTS

4333	Trends of Forest Phenology Studies in India-Challenges and Opportunity for Satellite Remote Sensing and Near Surface Sensors Dhruvi Sedha, Chandra Prakash Singh, Hitesh Solanki, Jincy Rachel Mathew, Mehul R. Pandya, Bimal K. Bhattacharya, C. Jeganathan and Siddhartha Khare	933
4334	Carbon Sequestration Potential of Woody Species in Thiagarajar College Campus, Madurai, India <i>K. Saraswathi and P. Sneha</i>	948
4335	Ethnobotanical Exploration of Polypetalous Riparian Flora along Beas River in Himachal Pradesh Utilized by Traditional Practitioners in the Management of Hypertension Neha Thakur and Nitesh Kumar	954
4336	Carbon Stock Quantities of <i>Shorea robusta</i> Gaertn. along Altitudinal Gradient in Shivalik Hills of Western Himalaya <i>Himshikha Gusain</i>	961
4337	Variation in Fruit and Seed Morphology of <i>Pyrus pashia</i> (Buch-Ham ex D. Don) in Alaknanda Valley of Garhwal Himalaya, India <i>Amreen, A.K. Negi and Himshikha Gusain</i>	966
4338	Wood-rotting Macrofungi of Kikruma Community Forest, Phek, Nagaland <i>Kuno Chuzho and M.S. Dkhar</i>	971
4339	Genetic Analysis of Morpho-Physiological Traits in Relation to Heat Tolerance in Barley ( <i>Hordeum vulgare</i> L.) <i>Ashok, Yogender Kumar and Amit</i>	977
4340	Grain Quality, Soil Fertility Status and Nutrient Uptake Pattern of Emmer Wheat ( <i>Triticum dicoccum</i> L.) under System of Wheat Intensification Nagesh Rathod, Kumar D. Lamani, Milid Potdar and Uday G. Reddy	985
4341	Nutrient Release Rate of <i>Crotolaria juncea</i> Degradation with Incorporation of Ligno-cellulolytic Bioinoculant under Dry and Flooded Conditions <i>Jaspreet Kaur, S.K. Gosal, S.S. Walia, Jupinder Kaur and Neha Khipla</i>	997
4342	Productivity and Biological Efficiency Indices of Sesamum + Cowpea Intercropping System in Response to Row Ratio and Nutrient Management Sabitha B., Shalini Pillai P, Usha C. Thomas Sheeja K. Raj and Chitra N.	1005
4343	Influence of Potassium and Phosphate Solubilizing Bacteria on Growth and Development of Davana ( <i>Artemisia Pallens</i> Bees) ( <i>Artemisia Pallens</i> Bees) Charul Khatri, Priyanka Mehra, Arul Prakash T., K.M. Prakhyath, N.D. Yogendra, B.S. Dattesh and V.S. Pragadheesh	1010
4344	Productivity and Economic Assessment of Diverse Rice ( <i>Oryza sativa</i> L.) Varieties with Varied Nitrogen Levels in Eastern Indo-Gangetic Plains Mohammad Hashim, Man Mohan Deo and Sanjeev Kumar	1016
4345	Source- Sink Regulation in Red Gram through Foliar Nutrition of Potassium and Growth Promoters in the Warm Humid Tropics of Kerala <i>Madiki Aashiq and Sheeba Rebecca Isaac</i>	1022
4346	Impact of Intercropping with Black Gram on Incidence of Stem Fly, <i>Melanagromyza sojae</i> (Zehntner) during the <i>Kharif</i> Season in Gujarat <i>N.P. Pathan, D.B. Sisodiya and R.D. Dodiya</i>	1027
4347	Effect of Date of Sowing and Varieties on Performance of Summer Fodder Pearlmillet ( <i>Pennisetum glaucum</i> L.) under North Gujarat Condition <i>Manisha M. Prajapati, Vikash Kumar, J.K. Patel and Veeresh Hatti</i>	1031
4348	Optimizing Planting Geometry and Nutrient Management for Enhanced Growth and Yield of Teff ( <i>Eragrostis tef</i> (Zucc.) Trotter) in Eastern Dry Zone of Karnataka Manjunath S. Melavanki, B. Boraiah, Mahantesh B. Nagangoudar and R.T. Chethan Babu	1037
4349	Assessment of Heavy Metal Pollution in Soil of Different Land Uses of in Semi-Arid Region of Jaipur, Rajasthan <i>Sonali Tiwari, Naveen Kumar, Priyanka Jatav and Archana Meena</i>	1042

Date of Publication 30.10.2024