



Relative Abundance of Cyanobacteria in Soil of Khasi Hills, Meghalaya and Determination of Antioxidant Potential of some Dominant Species

Carefulness Macnelson Dirborne and Papiya Ramanujam¹

School of Biological Sciences, Department of Botany, University of Science and Technology Meghalaya
Baridua-793 101, India

¹School of Life Sciences, Department of Botany, North-eastern Hill University (NEHU), Mawlai Umshing-793 022, India
E-mail: cariedirborne@gmail.com

Abstract: The relative abundance and antioxidant potential of some dominant cyanobacteria in the selected landuse type of Khasi Hills, Meghalaya were evaluated. A total of 105 species were isolated. Species of the genus belonging to *Oscillatoria* were with relative abundance of 41 to 75%. However, species occurring throughout the year with high relative abundance were *Synechococcus aeruginosus* (77-82%), followed *Oscillatoria limosa* (75%) and *Anabaena spiroides* (72%). The genus of *Oscillatoria* (*O. curviceps* and *O. limosa*) were with high phenolic (77.05 and 73.15µgGAE/ml respectively) and flavonoid contents (65.10 and 47.33µgGE/ml respectively). These can also scavenge DPPH radical and superoxide anion more efficiently than the other species. *Oscillatoria* species which was observed with high relative abundance in this part of the region could be used as a good source of antioxidant.

Keywords: Relative abundance, Antioxidant, Cyanobacteria, *Oscillatoria*, Khasi hills

Soil ecosystems formed the most important non-aqueous habitat for the terrestrial algae. Tropical India provides a favourable environment for the growth of cyanobacteria (Chellappa et al 2004, Geethu and Shamina 2021). Cyanobacteria are the oldest phototrophic microorganisms and play an important role as primary producers and pioneer organisms in soil crust (Budel et al 2016) and improve carbon and nitrogen contents, aggregation and stability of soil and also secretes the exopolysaccharides (Rossi et al 2018). The search for natural antioxidant has gained much interest due to the toxicity of synthetic antioxidant (Andrade et al 2019). Several studies have demonstrated that the amount of carotenoids contributed to the antioxidant capacity of microalgae (Takaichi 2011). Cyanobacteria are known to be a source of bioactive substances with many potential antioxidant compounds (Singh et al 2017, Morone et al 2019). Antioxidants are effective because they can donate their own electrons to Reactive Oxygen Species (ROS) and thereby neutralizing its adverse effects (Kohen and Nyska 2002). Thus the study was conducted in order to isolate the cyanobacteria with high relative abundance which can then be used as antioxidants.

MATERIAL AND METHODS

Collection of samples: Soil was collected from lands under shifting cultivation (1st year, 2nd year and 3rd year), agricultural farmlands (potato/rice field, potato/maize field), forests and

plantation (Mawphlang Sacred Grove, pine forest and citrus plantation) from various locations of Khasi hills, Meghalaya. Surface soil and soil near the roots regions were collected randomly from 15 different parts of collection spots (10 m²) with each collection spot approximately 10 m apart from each other. Soil samples from each collection spots were mixed together to form a composite mixture for the study of cyanobacteria community structure in selected land use type.

Preparation of samples: About 10 g of each soil sample was placed in a flask and diluted 100-fold with distilled water. BG 11 culture media was used in liquid or solidified form with 1.5% agar were used for culture and isolation. All the isolated cyanobacteria were observed under trinocular microscope (Olympus BX41) and identified to the possible lower taxonomic level by consulting standard books and monographs (Desikachary 1959, Philipose 1967, Prescott 1982, John et al 2002). Classification of cyanobacteria was carried out in accordance with Komárek et al (2014) and Guiry and Guiry (2020). The dominant cyanobacteria were grown in controlled condition of temperature at 25±1°C in light intensity of 40 µmol photons m⁻² s⁻¹ for 30 days. Culture experiments were conducted under a regime of 16 hour light/8 hour dark.

Data analysis: Relative abundance of a species was calculated (Dey et al 2010),

$$\text{Relative abundance} = Y/X 100$$

Where, X = total number of samples collected, Y = number of samples from which soil cyanobacteria was isolated

Preparation of cyanobacteria extract: 1g of air dried material was extracted at room temperature using methanol. It was then filtered through a filter paper (Whatman 1) and evaporated to dryness. The extract was then dissolved in methanol and store in the refrigerator at 20°C.

Analysis of antioxidant compounds and activity: The total phenolic content was determined by following Slinkard and Singleton (1977) using Folin & Ciocalteu reagent and gallic acid as standard whereas total flavonoid was determined by spectrophotometric method given by Quettier-Delue et al (2000). DPPH radical scavenging activity was measured by method described by Brand-Williams et al (1995). The superoxide anion radical scavenging activity was measured following the methods given by Nishikimi et al (1972). The inhibition concentration at 50% inhibition (IC_{50}) was the parameter used to compare the radical scavenging activity.

RESULTS AND DISCUSSIONS

Distribution and relative abundance of cyanobacteria: Altogether 105 species belonging to 5 orders were observed in soils of different landuse type. Oscillatoriales (38%) were recorded to be dominant order followed by Nostocales, Synechococcales, Chroococcales and Spirulinales (Fig. 1).

Relative abundance was higher in agricultural farmlands followed by forests and plantation and lower in shifting cultivated land. *Synechococcus aeruginosus* was observed in soil of all landuse except in sacred grove. The highest relative abundance was in pine forest (82%) followed by citrus plantation. This could be attributed to the ability of this genus to tolerate stress (Stuart et al 2009). Some species observed with high abundance were *Anabaena spiroides* (72%) followed by *An. variabilis*, *An. oscillaroides* and *An. spiroides*. Some *Scytonema* species such as *Sc. hyalinum*, *Sc. geitleri* and *Sc. schmidtii* were observed in the form of crust (Table 1). Species of *Nostoc*, *Anabaena*, *Cylindrospermum* and *Scytonema* are widespread in Indian soil and were known to contribute abundantly to soil fertility (Nayak and Prasanna, 2007). *Aphanothece castagnei* and *Gloeotheca rhodochlamys* were both observed in soils of potato/rice field with a higher abundance (66.6%). Species of *Pseudoanabaena batrachospermum* (52%) were observed in soil of potato/rice field. Species of *Oscillatoria* observed during rice cultivation in potato/rice field were *O. limosa* (75%) followed by *O. curviceps* (65%), *O. prombosidea* (45%), *O. princeps* (42%), *O. agardhii* (41%) (Table 1). Vijayan and Ray (2015) observed the dominance of

Oscillatoriales revealed the ecological status of agroecosystem as a result of agricultural practices.

Antioxidant compounds and potential of dominant species: *O. curviceps* has the maximum amount of phenolic (77.05 $\mu\text{gGAE/ml}$) and flavonoid content (65.10 $\mu\text{gGE/ml}$). *Gloeotheca rhodochlamys* was with lowest amount of phenolic (9.43 $\mu\text{gGE/ml}$). Kharkongor and Ramanujam (2017) recorded highest amount of phenolic compound could be the reason for a species to exhibit the highest DPPH radical and superoxide anion radical scavenging activity. It is already known that the smaller DPPH and Superoxide anion radical scavenging activity at % inhibition IC_{50} , the better is the ability of an algal species to scavenge these radical and anions. Thus *O. curviceps* (156.23 $\mu\text{g/ml}$) could scavenge DPPH radical more efficiently than *Aphanothece castagnei* (786.89 $\mu\text{g/ml}$). The same could also be accounted for superoxide anion scavenging activity in which *O. curviceps* (213.54 $\mu\text{g/ml}$) can scavenge superoxide anion more efficiently than *Synechococcus aeruginosus* (889.14 $\mu\text{g/ml}$). The antioxidant properties of some selected cyanobacteria in which DPPH radical scavenging activity was highest in *Oscillatoria* sp that could be used as excellent source of antioxidant. Guerreiro et al (2020) observed that cyanobacteria from paddy fields have high levels of carotene. Therefore, antioxidant activity could possibly be attributed to the presence of high amount of carotenoids in *Oscillatoria* sp. Martinez and Barbosa (2008) concluded that carotenoids could quench radicals by hydrogen atom transfer or by accepting electrons from radicals. Kharkongor and Ramanujam (2017) observed highest amount of phenolic compound could be the reason for a species to exhibited highest DPPH radical scavenging activity and superoxide anion radical scavenging activity (Table 2).

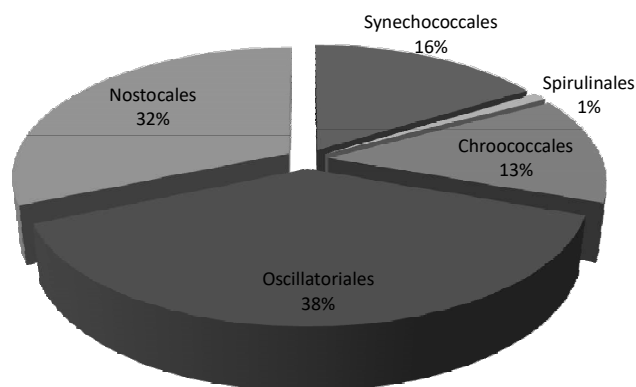


Fig. 1. Distributional pattern of different orders of Cyanobacteria

Table 1. Relative abundance and distribution of Cyanobacteria in soil of Khasi hills, Meghalaya

Taxa	Shifting cultivation			Farmlands		Forests and plantation		
	SC1	SC2	SC3	P/R	P/M	MSG	PF	CP
Phylum - Cyanobacteria Class - Cyanophyceae								
Order - Synechococcales								
<i>Aphanocapsa elachista</i> West & G.S. West	-	-	+	*	-	-	+	*
<i>A. incerta</i> (Lemmermann) G. Cronberg & Komárek	-	-	-	+	*	-	-	-
<i>Leptolyngbya battersii</i> Gomont	-	-	-	-	+	*	-	-
<i>L.faveolarum</i> (Gomont) Anagnostidis & Komárek	-	-	+	-	-	-	-	-
<i>L. vincentii</i> Komárek	-	-	-	+	*	-	-	-
<i>Merismopedia</i> sp	-	-	-	-	-	-	-	+
<i>Pseudoanabaena batrachospermorum</i> (Skuja) Anagnostidis & Komárek	-	-	-	+	**	-	-	+
<i>P. catenata</i> Lauterborn	-	-	-	-	-	-	-	+
<i>P. galeata</i> Böcher	-	-	-	+	*	+	*	-
<i>Pseudoanabaena</i> sp	-	-	-	-	-	-	-	+
<i>Schizothrix borealis</i> Komárek & Kovacic	-	-	-	-	-	+	*	-
<i>S. lateritia</i> Kützing ex Gomont	-	-	-	-	-	+	*	-
<i>S. telephoroides</i> Gomont	-	-	-	-	-	-	+	*
<i>S. tenuis</i> Woronichin	-	-	-	-	-	-	-	+
<i>S. tinctoria</i> Gomont ex Gomont	-	-	-	-	-	-	+	*
<i>Synechococcus aeruginosus</i> Nägeli	+	*	+	*	+	*	+	***
<i>Sy. elongatus</i> (Nägeli) Nägeli	-	-	-	+	*	-	-	-
Order - Spirulinales								
<i>Spirulina laxissima</i> G.S. West	-	-	-	+	*	-	-	-
Order - Chroococcales								
<i>Aphanothece castagnei</i> (Kützing) Rabenhorst	-	-	-	+	***	-	-	-
<i>Ap. densa</i> Silva	-	-	-	-	+	*	+	*
<i>Chlorogloea novacekii</i> Komárek & Montejano	-	-	-	-	-	+	-	***
<i>Chlorogloea</i> sp	-	-	-	-	-	-	-	+
<i>Chroococcus minor</i> (Kützing) Nägeli	-	-	-	+	**	-	-	***
<i>Ch. tenax</i> (Kirchner) Hieronymus	-	-	-	+	*	-	-	-
<i>Gloecapsa decorticans</i> (A. Braun) P. Richter	-	-	-	-	-	-	+	*
<i>G. alpina</i> Nägeli	-	-	-	-	-	+	*	-
<i>G. muralis</i> Kützing	-	-	-	+	**	-	-	-
<i>G. rupestris</i> Kützing	-	-	-	-	-	-	-	+
<i>Gloeothece rhodochlamys</i> Skuja	-	-	-	+	***	-	-	-
<i>Gl. tepidariorum</i> (A. Braun) Lagerheim	-	-	-	-	-	+	-	-
<i>Microcystis aeruginosa</i> Kützing (Kützing)	-	-	-	-	+	*	-	-
<i>M. smithii</i> Komárek & Anagnostidis	-	-	-	+	-	-	-	-
Order - Oscillatoriales								
<i>Geitlerinema splendidum</i> (Greville ex Gomont) Anagnostidis	+	-	-	+	-	-	-	-
<i>Ge. unigranulatum</i> Singh	-	-	-	+	-	-	-	-
<i>Lyngbya dendrobia</i> Brühl & Biswas	-	-	-	+	**	-	-	-
<i>L. martensiana</i> Meneghini ex Gomont	-	-	-	+	**	-	-	-
<i>L. putealis</i> Montagne ex Gomont	-	-	-	+	*	-	-	-
<i>L. semiplena</i> J. Agardh ex Gomont	-	-	-	-	-	+	*	-

Cont...

Table 1. Relative abundance and distribution of Cyanobacteria in soil of Khasi hills, Meghalaya

Taxa	Shifting cultivation			Farmlands		Forests and plantation		
	SC1	SC2	SC3	P/R	P/M	MSG	PF	CP
<i>L. shackletoni</i> West & G. S. West	-	-	-	+*	-	-	-	-
<i>L. taylorii</i> Drouet & Strickland	-	-	-	-	-	+	-	-
<i>Lyngbya</i> sp	-	-	-	-	-	-	+	-
<i>Microcoleus chthonoplastes</i> Thuret ex Gomont	-	-	-	-	-	+*	-	-
<i>Microcoleus</i> sp	-	-	-	-	-	-	-	+*
<i>Oscillatoria agardhii</i> Gomont	-	-	-	+**	-	-	-	-
<i>O. pseudogeminata</i> G. Schmid	+	+*	-	-	+	-	+	+*
<i>O. santa</i> Kützing ex Gomont	-	-	-	-	+*	-	-	-
<i>O. tenuis</i> C. Agardh ex Gomont	-	-	+*	-	-	-	-	+*
<i>O. vizagapatensis</i> Rao	-	-	-	+	-	-	-	-
<i>O. curviceps</i> C. Agardh ex Gomont	-	-	+**	+***	+**	-	+*	-
<i>O. irrigua</i> Kützing ex Gomont	-	+*	+**	+*	-	-	-	-
<i>O. limnetica</i> Lemmermann	-	-	-	-	-	-	-	+**
<i>O. limosa</i> C. Agardh ex Gomont	-	-	-	+***	+**	+*	-	-
<i>O. princeps</i> Vaucher ex Gomont	-	-	-	+**	-	-	-	-
<i>O. tergestina</i> Kützing	-	-	-	+	-	-	-	-
<i>O. amphibia</i> Agardh ex Gomont	-	-	-	+*	-	-	-	-
<i>O. anguina</i> Bory ex Gomont	-	-	-	+*	-	-	-	-
<i>O. brevis</i> Kützing ex Gomont	-	+**	-	-	-	-	-	-
<i>O. germinata</i> Schwabe ex Gomont	-	-	-	-	-	+	-	+***
<i>O. nigra</i> Vaucher ex Gomont	-	-	-	-	-	+	-	-
<i>O. proboscidea</i> Gomont	-	+*	-	+**	-	-	-	-
<i>O. rubescens</i> De Candolle ex Gomont	-	-	-	+	-	-	-	+
<i>Oscillatoria</i> sp	-	-	-	+*	-	-	-	-
<i>Phormidium abronema</i> Skuja	-	-	-	+*	+*	-	-	-
<i>P. inundatum</i> Kützing ex Gomont	-	-	-	-	-	-	+*	-
<i>P. papyraceum</i> Skuja	-	-	-	+**	-	-	-	-
<i>P. retzii</i> Kützing ex Gomont	-	-	-	-	-	-	+	-
<i>P. tenue</i> Gomont	+	+*	-	-	+**	-	-	-
<i>P. corium</i> Gomont ex Gomont	-	-	-	-	-	-	-	+
<i>Phormidium</i> sp 1	-	-	+	-	-	-	-	-
<i>Phormidium</i> sp 2	-	-	-	+	-	-	-	-
<i>Plectonema</i> sp	-	-	-	-	-	-	-	+
<i>Symploca muscorum</i> Gomont ex Gomont	-	-	-	-	-	+	-	-
Order - Nostocales								
<i>Amorphonostoc pruniforme</i> (Kützing) Elenkin	-	-	-	+	-	-	-	-
<i>Anabaena constricta</i> (Szafer) Geitler	-	-	-	-	-	-	+	-
<i>An. catenula</i> Kützing ex Bornet & Flahault	-	-	-	-	-	+	+	-
<i>An. iyengarii</i> Kharadwaja	-	-	-	+**	-	-	-	-
<i>An. oryzae</i> F. E. Fritsch	-	-	-	+**	-	-	-	-

Cont...

Table 1. Relative abundance and distribution of Cyanobacteria in soil of Khasi hills, Meghalaya

Taxa	Shifting cultivation			Farmlands		Forests and plantation		
	SC1	SC2	SC3	P/R	P/M	MSG	PF	CP
<i>An. oscillarioides</i> (Bory ex Bornet) Flahault	-	-	+ **	-	-	+ **	-	-
<i>An. sphearica</i> Bornet & Flahault	-	-	-	+ **	-	-	-	-
<i>An. spiroides</i> Klebahn	-	-	+ **	-	+ **	-	+ ***	+ ***
<i>An. torulosa</i> Lagerheim ex Bornet & Flahault	-	-	-	+	-	-	-	-
<i>An. variabilis</i> Kützing ex Bornet & Flahault	-	-	-	+ *	+ **	-	-	+ ***
<i>Aulosira</i> sp	-	-	-	+ **	-	-	-	-
<i>Calothrix</i> sp 1	-	-	-	+ *	-	-	-	-
<i>Calothrix</i> sp 2	-	-	-	-	-	-	-	+ *
<i>Cylindrospermum michailovskoense</i> Elenkin	-	-	-	-	-	+ *	-	-
<i>Cy. muscicola</i> Kützing ex Bornet & Flahault	-	-	-	-	-	+ *	-	-
<i>Nostoc carneum</i> C.Agardh ex Bornet & Flahault	-	-	-	+ **	-	+ **	-	+ *
<i>N. commune</i> Vaucher ex Bornet & Flahault	-	-	-	-	-	+ *	-	-
<i>N. linckia</i> Bornet ex Bornet & Flahault	-	-	-	-	-	-	+ *	-
<i>N. muscorum</i> C. Agardh ex Bornet & Flahault	-	-	-	-	+ **	-	-	-
<i>N. padulosum</i> Kützing ex Bornet	-	-	-	+ *	-	-	-	-
<i>N. pruniforme</i> Hariot	-	-	-	-	-	-	+ *	-
<i>N. spongiaeformae</i> C. Agardh ex Bornet & Flahaut	-	-	-	+ **	-	-	-	-
<i>Scytonema geitleri</i> Bharadwaja	-	-	-	-	-	-	-	+ **
<i>Sc. hyalinum</i> Gardner	-	-	-	-	-	-	-	+ **
<i>Sc. mirabile</i> Bornet	-	-	-	-	-	+ *	-	-
<i>Sc. schmiditii</i> Gomont	-	-	-	-	-	-	-	+ **
<i>Scytonema</i> sp 1	-	-	-	+ **	-	-	-	-
<i>Scytonema</i> sp 2	-	-	-	-	-	+ *	-	-
<i>Stigonema hormoides</i> Bornet & Flahault	-	-	-	-	-	-	-	+ **
<i>St. mammilosum</i> Agardh ex Gomont	-	-	-	-	-	-	+ **	-
<i>St. ocellatum</i> Thuret ex Bornet & Flahault	-	-	-	-	-	+ *	-	-
<i>Tolypothrix</i> sp	-	-	-	-	-	-	-	+ *
<i>Westiellopsis</i> sp	-	-	-	-	-	+ *	-	-

(SC 1 - 1st year, SC 2 - 2nd year, SC 3 - 3rd year, P/R - Potato/Rice field, P/M - Potato/Maize field, MSG - Mawphlang Sacred Grove, PF - Pine Forest, CP - Citrus Plantation). Relative abundance - (*) indicates 20% to 39%, (**) indicates 40% to 59%, (***) indicates > 60%; ' + ' indicates present, ' - ' indicates absent)

Table 2. Antioxidant potential of some dominant cyanobacterial species

Cyanobacteria species	Total phenolics content (µgGAE/ml of extract)	Flavonoid content (µgGE/ml of extract)	DPPH Radical scavenging activity IC ₅₀ (µg/ml)	Superoxide anion scavenging activity IC ₅₀ (µg/ml)
<i>Anabaena oscillarioides</i>	23.12±1.67	12.34±0.98	456.34±1.54	543.78±0.34
<i>An. spiroides</i>	24.00±2.12	11.23±1.65	567.56±0.45	678.10±1.08
<i>An. variabilis</i>	24.12±1.87	11.19±1.34	456.21±1.12	657.05±1.87
<i>Aphanothece castagnei</i>	25.26±1.19	15.23±2.34	786.89±4.23	345.54±3.09
<i>Gloeothece rhodochlamys</i>	27.56±1.21	12.43±1.45	689.19±5.67	452.16±8.12
<i>Oscillatoria curviceps</i>	77.05±1.23	65.10±1.89	156.23±9.54	213.54±4.12
<i>O. germinata</i>	54.10±1.45	48.34±3.23	167.65±7.23	256.19±1.23
<i>O. limosa</i>	73.15±3.56	47.33±5.34	172.29±10.21	237.45±9.65
<i>Scytonema geitleri</i>	57.34±5.78	42.76±1.42	598.21±2.65	256.87±1.87
<i>Sc. hyalinum</i>	40.17±2.22	19.45±1.98	456.34±2.98	548.21±1.98
<i>Sc. schmiditii</i>	41.86±3.89	21.24±2.67	324.56±12.09	439.10±8.43
<i>Synechococcus aeruginosus</i>	21.16±4.32	10.28±1.88	764.17±3.12	889.14±1.87

Values are mean ± SE of three replicates

CONCLUSION

All the twelve cyanobacterial species have some antioxidant activity. *O. curviceps* has the maximum amount of phenolic and flavonoid content. This species also scavenged DPPH radical and superoxide anion more efficiently than all the other species. Other species with potentially higher scavenging activity was observed for *O. germinata*, *O. limosa* and *Scytonema geitleri*. The high antioxidant activity along with high relative abundance of these species can be an excellent source of raw materials from this part of the region.

REFERENCES

- Andrade MA, Lima V, Sanches-Silva A, Vilarinho F, Castilho MC, Khwaldia K and Ramos F 2019. Pomegranate and grape by-products and their active compounds: Are they a valuable source for food applications? *Trends Food Science and Technology* **86**: 68-84.
- Brand-Williams W, Cuvelier ME and Berset C 1995. Use of free radical methods to evaluate antioxidant activity. *Journal of Food Science and Technology* **28**(1): 25-30.
- Büdel B, Dulić T, Darienko T, Rybalka N and Friedl T 2016. Cyanobacteria and algae of biological soil crusts. In: Weber B, Büdel B, Belnap J (eds) *Biological soil crusts: an organizing principle in drylands*. Springer International Publishing, Cham, pp 55–80
- Chellappa S, Chellappa N and Marinho IR 2004. Freshwater phytoplankton assemblages and bloom of toxic cyanophyceae of Campo Grande reservoir of Rio Grande do Norte of Brazil. *Indian Hydrobiology* **7**: 151-171.
- Desikachary T 1959. *Cyanophyta*, I. C. A. R. Monograph on algae, New Delhi.
- Dey HS, Tayung K and Bastia AK 2010. Occurrence of nitrogen fixing Cyanobacteria in local rice fields of Orissa, India. *Ecoprint* **17**: 77-85.
- Geethu V and Shamina M 2021. Filamentous cyanobacteria from western ghats of north kerala, India. *Bangladesh Journal of Plant Taxonomy* **28**(1): 83-95.
- Guerreiro A, Andrade MA, Menezes C, Vilarinho F and Dias E 2020. Antioxidant and Cytoprotective Properties of Cyanobacteria: Potential for Biotechnological Applications. *Toxins* **12**: 548.
- Guiry MD and Guiry GM 2020. *AlgalBase. World-wide electronic publication, National University of Ireland, Galway*. <http://www.algalbase.org>.
- John DM, Whitton BA and Brook AJ 2002. *The freshwater algal flora of the British Isles: An identification guide to freshwater and terrestrial algae*, Cambridge University Press, Cambridge.
- Kharkongor D and Ramanujam P 2017. Antioxidant Activities of Four Dominant Species of *Trentepohlia* (Trentepohliales, Chlorophyta). *International Journal of Complementary and Alternative Medicine* **8**(5): 00270
- Kohen R and Nyska A 2002. Oxidation of biological systems: oxidative stress phenomena, antioxidants, redox reactions, and methods for their quantification. *Toxicologic Pathology* **30**(6): 620-650.
- Komárek J, Kastovsky J, Mares J and Johansen JR 2014. Taxonomic classification of cyanoprokaryotes (Cyanobacterial genera) using a polyphasic approach. *Preslia* **86**: 295-335.
- Martínez A and Barbosa A 2008. Antiradical power of carotenoids and vitamin E: testing the hydrogen atom transfer mechanism. *Journal of Physical Chemistry B* **112**(51): 16945-16951.
- Morone J, Alfeus A, Vasconcelos and Martins R 2019. Revealing the potential of cyanobacteria in cosmetics and cosmeceuticals: A new bioactive approach. *Algal Research* **41**: 101541.
- Nayak S and Prasanna R 2007. Soil pH and its role in Cyanobacterial abundance and diversity in rice field soils. *Applied Ecology and Environmental Research* **5**(2): 103-113.
- Nishikimi M, Appaji N and Yagi K 1972. The occurrence of superoxide anion in the reaction of reduced phenazine methosulfate and molecular oxygen. *Biochemical and Biophysical Research Communications* **46**(2): 849-854.
- Philipose MT 1967. *Chlorococcales*, I.C.A.R., Krishi Bhawan, New Delhi.
- Prescott GW 1982. *Algae of Western Great Lakes Area, Michigan State University, Otto Koelt Science Publishers, Koenigstein, West Germany*.
- Quettier-Deleu C, Gressier B, Vasseur J, Dine T, Brunet C, Luyckx M, Cazin M, Cazin JC, Bailleul F and Trotin F 2000. Phenolic compounds and antioxidant activities of buckwheat (*Fagopyrum esculentum* Moench) hulls and flour. *Journal of Ethnopharmacology* **72**(1): 35-42.
- Rossi F, Mugnai G, and De Philippis R 2018. Complex role of the polymeric matrix in biological soil crusts. *Plant and Soil* **429**: 19-34.
- Singh R, Parihar P, Singh M, Bajguz A, Kumar J, Singh S, Singh VP and Prasad SM 2017. Uncovering potential applications of cyanobacteria and algal metabolites in biology, agriculture and medicine: Current status and future prospects. *Frontiers in Microbiology* **8**: 515.
- Slinkard K and Singleton VL 1977. Total phenol analysis: automation and comparison with manual methods. *American Journal of Enology and Viticulture* **28**(1): 49- 55.
- Stuart RK, Dupont CL, Johnson DA, Paulsen IT and Palenik B 2009. Coastal strains of Marine Synechococcus species exhibit increased tolerance to copper shock and a distinctive transcriptional response relative to those of open-ocean strains. *Applied and Environmental Microbiology* **4**(5): 753-758.
- Takaichi S 2011. Carotenoids in algae: Distributions, biosyntheses and functions. *Marine Drugs* **9**(6): 1101-1118.
- Vijayan D and Ray JD 2015. Ecology and diversity of Cyanobacteria in Kuttanadu paddy wetlands, Kerala, India. *American Journal of Plant Sciences* **6**: 2924-2938.
- Zenova GM, Shtina EA, Dedysh SN, Glagoleva OB, Likhacheva AA and Gracheva TA 1995. Ecological relations of algae in biocenoses. *Mikrobiologia* **64**: 121-133.