



Screening of Seaweeds for Antibacterial and Antifungal Activities

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Abstract: Commonly occurring seaweeds, *Caulerpa scapelliformis*, *Ulva lactuca*, *Padina tetrastromatica*, *Stoechospermum marginatum* and *Acanthophora spicifera* collected from the Tuticorin coast, Tamil Nadu (India) were evaluated for antibacterial and antifungal activity by agar diffusion method. Four different solvents viz. petroleum ether, benzene, chloroform, and methanol were used for extraction. By using commercial medicine, Amikacin-AK30-30mcg/disc, Cephalexin-CN30-30mcg/disc, Ciprofloxacin-CIP5-5mcg/disc and Fluconazole-FLC25-25mcg/disc as control and zones of inhibition were compared. The seaweed extracted using petroleum ether as a solvent showed the best antifungal activity and was more effective than commercial antibiotic, fluconazole. None of the extracts were active against *Staphylococcus aureus*, a gram-positive bacterial pathogen. The methanol extract was active against *Salmonella typhi*, *Pseudomonas aeruginosa*, *Proteus vulgaris*, and *Escherichia coli* except for *Caulerpa scapelliformis* and *Ulva lactuca*. All the four extracts of *Acanthophora spicifera* showed antibacterial activity.

Keywords: Seaweeds, Antibacterial, Antifungal, Solvents

The marine environment has a rich biological and chemical diversity (Dayanidhi et al 2021). This diversity of marine organisms and habitat offers a wide variety of natural marine compounds. Several oceanic compounds show pharmacological activities and are helpful in the invention and discovery of novel bioactive compounds, primarily for deadly diseases like Cancer, Acquired immunodeficiency syndrome (AIDS) and Arthritis (Samuel et al 2011, Pham-Huy and Huy 2022). Many other compounds have been developed as analgesics or to treat inflammation (Giriwono et al 2019). Seaweed (macroalgae), a marine organism widely distributed in the coastal regions of all the continents. Several scientists have worked on the seaweed extracts, exhibiting antibacterial and antifungal activity (Hellio et al 2000, Lima-Filho et al 2002, Taskin et al 2007, Seenivasan et al 2010). Moreover, crude extracts of Indian seaweeds were found to be active only against Gram-positive bacteria whereas, majority of the pathogenic bacteria are gram-negative. Therefore, keeping this in view, the present work was designed to study the antibacterial and antifungal activity of four different solvents extracts of five seaweeds collected from Tuticorin coast, Tamil Nadu.

MATERIAL AND METHODS

Collection and preparation of selected seaweeds: Fresh seaweed samples of *Caulerpa scapelliformis*, *Ulva lactuca*, *Padina tetrastromatica*, *Stoechospermum marginatum* and *Acanthophora spicifera* were handpicked from the intertidal zone of Hare Island, Tuticorin (TN) located at 8.8°N, 78.3°E

The collected seaweed samples were cleaned with seawater and freshwater, dried in the shade, and powdered to extract antimicrobial compounds.

Preparation of organic seaweed extracts: The organic seaweeds extracts were prepared by following the method of El Shafay et al (2016). Powdered seaweed sample (5g) were soaked in 50 mL of four different solvents (analytical grade) i.e. petroleum ether, benzene, chloroform and methanol for three days. Then soaked samples were filtered and concentrated in a rotary evaporator at 35°C. The residual water was then removed with the help of vacuum pump (Agilent vacuum pump PVL401), followed by suspension of weighted crude extract in dimethyl sulfoxide (DMSO) to the final concentration of 50 mg/mL and stored in a refrigerator.

Bioassay: For bioassay, fungal pathogen *Candida albicans* ATCC 90028 and five gram-negative bacterial pathogens namely *Salmonella typhi* ATCC 35640, *Pseudomonas aeruginosa* ATCC 27853, *Proteus vulgaris* ATCC 33420, *Klebsiella pneumoniae* ATCC 700603, *Escherichia coli* ATCC 25922 and one pathogenic gram-positive bacteria namely *Staphylococcus aureus* ATCC 25923 were used. A loop full of the microorganism was inoculated in nutrient broth from the 24 hours incubated nutrient agar slant of each test organism at pH-7.4 to activate the bacterial strains used as test organisms. The broths were kept for incubation at 37°C for 24 hours so that the microorganism can grow till the log phase. The nutrient broth was maintained as control without inoculating the test organisms. These bacteria were obtained from Hi-Media Ltd., Mumbai. Amikacin-AK30-30mcg/disc,

Cephalexin-CN30-30mcg/disc, Ciprofloxacin-CIP5-5mcg/disc and Fluconazole-FLC25-25mcg/disc were used as control medicine. The bioassay was carried out under a sterile condition in a clinical laboratory by following the agar diffusion method as described by Perez et al (1990). Inoculums in the exponential phase of growth, equivalent to a 0.5 McFarland standard were swabbed on to the surface of Muller Hinton agar media. Different crude extract (2 mL) was transferred to a sterile 6 mm Whatman No.1. Filter paper disc (E-760) and after drying it was placed in the seeded agar plate, along with the control discs which was prepared with the solvents alone. The inhibition zone was observed after 24 hours of incubation at 37°C.

RESULTS AND DISCUSSION

Among five different seaweeds (*Caulerpa scalpelliformis*, *Ulva lactuca*, *Padina tetraströmatica*, *Stoechospermum marginatum* and *Acanthophora spicifera*) collected, the extracts of all seaweeds except *Caulerpa scalpelliformis* showed specific activity against the pathogens (bacteria i.e. *Salmonella typhi*, *Pseudomonas aeruginosa*, *Proteus vulgaris*, *Klebsiella pneumoniae* and *Escherichia coli* and

fungi i.e. *Candida albicans*) (Table 1, 2). The present study showed that petroleum ether extract of *Ulva lactuca* is active against fungal pathogen, *Candida albicans*, in accordance with the findings of Oranday et al (2004). Further, the petroleum ether extract of *Caulerpa scalpelliformis*, *Ulva lactuca*, and *Padina tetraströmatica*; the benzene extract of *Stoechospermum marginatum* were active against fungal pathogen *Candida albicans*. The different extracts of the red alga, *Acanthophora spicifera* were not found active against fungal pathogen, *Candida albicans*. The zone of inhibition indicates that extracts of all the seaweeds exhibit antifungal activities except *Acanthophora spicifera* which are quite comparable with the commercial antifungal agent i.e. fluconazole (Table 2). The petroleum ether extract of *Caulerpa scalpelliformis* was more effective (32mm) than the commercial antifungal agent fluconazole (27mm). Similarly, Kaur et al (2016) observed *Inula racemosa* root extract were active against phyto-pathogenic fungi (*Dreschlera oryzae*). Tuney et al (2006) found that the methanol extract of *Ulva rigida* collected from the coast of Urla (Izmir, Turkey) had no antifungal activity against *Candida albicans*. Karthick et al (2014) reported that *Caulerpa scalpelliformis* contains

Table 1. Antibacterial activity of sea weeds extracts from Hare Island of Tuticorin coast

Name of the bacterial pathogens	Control medicine			<i>Caulerpa scalpelliformis</i>				<i>Ulva lactuca</i>				<i>Acanthophora spicifera</i>				<i>Padina tetraströmatica</i>				<i>Stoechospermum marginatum</i>			
	AK30	CN30	CIP5	PE	BZ	CF	MN	PE	BZ	CF	MN	PE	BZ	CF	MN	PE	BZ	CF	MN	PE	BZ	CF	MN
<i>Salmonella typhi</i> ATCC 35640	16	20	21	-	-	-	-	-	-	-	-	15	-	-	-	-	-	-	-	-	-	-	16
<i>Pseudomonas aeruginosa</i> ATCC 27853	14	15	14	-	-	-	-	-	-	-	-	-	-	-	14	17	-	-	14	-	14	-	-
<i>Proteus vulgaris</i> ATCC 33420	16	20	21	-	-	-	-	16	-	-	-	-	-	-	-	-	-	-	15	-	-	-	-
<i>Klebsiella pneumoniae</i> ATCC700603	26	29	30	-	-	-	-	-	-	23	-	-	20	24	-	-	-	-	-	-	-	-	-
<i>Escherichia coli</i> ATCC 25922	16	20	22	-	-	-	-	-	-	-	-	-	-	-	16	-	-	-	-	-	-	-	16

*PE-Petroleum ether; BZ-Benzene; CF-Chloroform; MN-Methanol; FLC25-Fluconazole; AK30-Amikacin; CN30-Cephalexine; CIP5-Ciprofloxacin
 *Zone in mm indicates the distance from the border of the disc to the edge of the clear zone.
 (-): hyphen indicates no clear zone has been observed

Table 2. Antifungal activities of sea weeds extracts from Hare Island of Tuticorin coast

	Control medicine			<i>Caulerpa scalpelliformis</i>				<i>Ulva lactuca</i>				<i>Acanthophora spicifera</i>				<i>Padina tetraströmatica</i>				<i>Stoechospermum marginatum</i>			
	FLC25	PE	BZ	CF	MN	PE	BZ	CF	MN	PE	BZ	CF	MN	PE	BZ	CF	MN	PE	BZ	CF	MN		
<i>Candida albicans</i> ATCC90028	27	32	-	-	-	24	-	-	-	-	-	-	-	22	-	-	-	-	22	-	-	-	

See Table 1 for details

tannin, flavonoids, glycosides, phenols, saponins, and terpenoids. This indicated that in the particular case that petroleum ether could be able to successfully extract antifungal compounds in an effective concentration.

The petroleum ether extracts of *Acanthophora spicifera* was sensitive against Gram-negative bacterial pathogen *Salmonella typhi* but comparatively lower than the commercial antibiotic agents i.e. Amikacin, Cephalexin and Ciprofloxacin. The methanol extract of *Stoechospermum marginatum* was active against Gram-negative bacterial pathogen *Salmonella typhi*. The methanol extract of a *Padina tetrastromatica* and benzene extract of *Stoechospermum marginatum* showed similar activities like Ciprofloxacin and Amikacin against Gram-negative bacterial pathogen *Pseudomonas aeruginosa*. However, petroleum ether extracts of *Padina tetrastromatica* was more effective (17 mm) than commercial antibiotics. The methanol extract of *Ulva lactuca* showed antibacterial activity against Gram-positive bacterium *Staphylococcus aureus* and Gram-negative bacterium *Escherichia coli* (Selvi et al 2001, Oranday et al 2004). The methanol extract of *Ulva lactuca* did not show such antibacterial activity against all these pathogenic bacteria whereas petroleum ether extract of *Ulva lactuca* was active against Gram-negative bacterial pathogen *Proteus vulgaris*. This could be attributed to the presence of lipophilic and phenolic compounds, especially steroids fatty acids, in *Ulva lactuca* organic extract towards antimicrobial activity (El-Baky et al 2008). The zone of inhibition (16 mm) indicates that the methanol extract of *Padina tetrastromatica* was similarly active against *Escherichia coli* as commercial antibiotic i.e. Amikacin. The chloroform extract of *Ulva lactuca* and *Acanthophora spicifera* and the benzene extract of *Acanthophora spicifera* showed lesser activity than needed against Gram-negative bacterial pathogen *Klebsiella pneumoniae*. The methanol extract of *Stoechospermum marginatum* and *Acanthophora spicifera* were active against Gram-negative bacterial pathogen *Escherichia coli*. All the extracts studied were inactive against the Gram-positive bacterial pathogen *Staphylococcus aureus* which might be attributed to the inhibited entry of antimicrobial agents into cell due to the presence of high percentage (90-95%) peptidoglycan, lipopolysaccharides and phospholipids in the bacterial cell wall. Marke et al (2022) also tested *Tinospora cordifolia* (natural herbal shrub) extracts for assessing antibacterial activity and suggested that phytochemical compounds present in this natural herbal shrub can be utilized for formulation of drugs against some gastrointestinal pathogens like *Staphylococcus aureus*, *Escherichia coli*, *Vibrio cholera*, *Salmonella typhi* and *Shigella*.

During the present study, different extracts of *Caulerpa scalpelliformis* were inactive against all the studied pathogenic bacteria whereas Selvi et al (2001) reported that the methanol extract of *Caulerpa scalpelliformis* showed antibacterial activity against Gram-positive bacterium *Staphylococcus aureus* and Gram-negative bacterium *Salmonella typhi*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae* and *Escherichia coli*. Non-conformance of the findings might be due to different natural factors such as environmental conditions (light, temperature or salinity), reproductive state, age of the seaweed, geographic location and seasonality (Perez et al 2016). Hence, information made during the present study revealed that seaweed extracts with different solvents exhibit antibacterial and antifungal properties which could be further utilized for production of organic pharmaceutical products. Moreover, this created a quest also for the discovery of antibiotic compounds of natural origin.

CONCLUSION

Seaweeds have huge potential for discovery of compounds with various bioactivities. Seaweeds collected from the coastal areas of Tuticorin exhibit antibacterial and antifungal properties. The test microorganism showed resistance towards seaweed extracts prepared with different organic solvents, which is indicating the availability of biologically active compounds at altered concentrations in seaweeds. The study is signifying the need to explore the possibilities of having bioactive compounds in seaweeds with pharmaceutical importance.

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